

Article ID: 175998
DOI: 10.5586/aa/175998

Publication History
Received: 2023-09-06
Accepted: 2023-11-28
Published: 2023-12-19

Handling Editor
Piotr Sugier; Maria
Curie-Skłodowska University in
Lublin, Lublin, Poland;
<https://orcid.org/0000-0002-1448-1517>

Authors' Contributions
BZ: Research concept and design; BZ: Collection and/or assembly of data; RN: Data analysis and interpretation; BZ, RN: Writing the article, RN: Critical revision of the article; BZ: Final approval of the article


Funding
This paper was supported by the European Union Next-Generation EU [Piano Nazionale di Ripresa e Resilienza (PNRR) – Missione 4 Componente 2, Investimento 1.4 – D.D. 1032 17/06/2022, CN0000022]. This manuscript reflects only the authors' views and opinions; neither the European Union nor the European Commission can be considered responsible for them.

Competing Interests
No competing interests have been declared.

Copyright Notice
© The Author(s) 2023. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits redistribution, commercial and noncommercial, provided that the article is properly cited.

REVIEW

Arcopilus aureus: A valuable endophytic associate of hazelnut

Beata Zimowska ^{1*}, Rosario Nicoletti ^{2,3}

¹Department of Plant Protection, Subdepartment of Plant Pathology and Mycology, University of Life Sciences, K. St. Leszczyńskiego 7, 20-069 Lublin, Poland

²Council for Agricultural Research and Economics, Research Centre for Olive, Fruit and Citrus Crops, Via Torrino, 81100 Caserta, Italy

³Department of Agricultural Sciences, University of Naples 'Federico II', Via Università 100, 80055 Portici, Italy

* To whom correspondence should be addressed. Email: beata.zimowska@up.lublin.pl

Abstract

Endophytic fungi are increasingly being considered for their ecological role and effects on plant protection and nutrition. Their contribution in improving fitness can be particularly relevant in the case of semi-extensive tree crops, such as hazelnut. In this manuscript, the occurrence and properties of the species *Arcopilus aureus* are reviewed, following its finding as a component of the endophytic microbiome of hazelnut in the distant geographic and climatic contexts of Poland and Italy, representing an indication of its widespread association with this plant and a possible role in protection against biotic adversities.

Keywords

Corylus avellana; taxonomy; Chaetomiaceae; defensive mutualism; antagonism; biological compounds; endophytic fungi

1. Introduction

Components of the endophytic phytobiomes establish mutualistic associations with their hosts where, in exchange of nutrients and habitat, they provide protection against various biotic and abiotic stresses. Their adaptation to life inside the tissues of other eukaryotes might trigger the capacity to synthesize novel bioactive secondary metabolites through the interaction with the host's genome (Ekbic et al., 2022; Gakuubi et al., 2021; Nicoletti & Fiorentino, 2015; Rokas et al., 2020).

During the recent years, an explosion of research focused on the characterization of endophytes and their occurrence and functions, as well as on their aptitude as providers of ecosystem services have been observed (e.g. Dimitrova & Nacheva, 2021; Malinowski & Belesky, 2019; Sumi et al., 2022). Promises of a new green revolution aiming at the implementation of new techniques, sustainable solutions, and disruptive innovation trigger increasing interest of researchers all over the world in biodiversity and in bioprospecting endophytes as an alternative source of bioactive compounds to be used for industrial applications. Among others, members of *Chaetomium* and related genera represent a model of endophytic fungi as a rich source of unique bioactive metabolites that can be considered for diverse biotechnological deployment (Rao et al., 2023). This article particularly focuses on the species *Arcopilus aureus* (Chivers) X. Wei Wang & Samson (formerly *Chaetomium aureum*), following the finding of endophytic isolates from hazelnut (*Corylus avellana* L.) in the course of our ongoing studies carried out in Poland and in Italy.

2. From *Chaetomium* to *Arcopilus*: A taxonomic excursus on the Chaetomiaceae and the identification of *Arcopilus aureus*

The family Chaetomiaceae (Sordariomycetes, Sordariales) was appointed to contain fungi that create non-stromatic sexual bodies with a membranaceous wall, covered

by perithecial hairs similar to hyphal outgrows, fasciculate and evanescent asci, and single-celled, soft, brown or gray-brown ascospores; *Chaetomium globosum* Kunze was assumed as the type species (Kunze & Schmidt, 1817; Winter, 1885). Using classical methods, several attempts were made to study morphological details for species identification. The most relevant features taken into account were the characters of ascomata. Particularly, the structure of the ascomatal hairs was used as the most important feature in the first monographs, respectively describing 10 species (Zopf, 1881) and 28 species (Chivers, 1915). The appearance of ascomatal outgrows together with the shape and size of the sexual bodies and ascospores, in addition to the conspicuous characteristics, were used as the major separation characteristics by other taxonomists. According to these features, Skolko and Groves (1953) recognized 53 species, while Ames (1961) described 85 distinct species of *Chaetomium*.

Over the course of taxonomic redefinition Chaetomiaceae, the most crucial amendment was made by Josef von Arx. Instead of focusing on the diverse perithecial hairs, he laid emphasis on the features of asci and ascospores, the presence of germ pores on ascospores, and the structures of the ascomatal wall to distinguish species (Rong & van Warmelo, 1988). According to his taxonomical system, *Chaetomium* was kept for species creating ostiolate perithecia covered by relatively well-developed outgrows, and this feature allowed discriminating *Chaetomium* from other genera (von Arx et al., 1986).

Other taxonomical parameters, such as hyphal growth, colony color and size, and thickness of the hyphae within the colony, have been studied to distinguish species within *Chaetomium* (Plomley, 1959; Sharma & Pandey, 2010). Growth dynamics, colony form, morphology and appearance of mycelium and sexual spores varied greatly depending on the carbon source. Lignocellulose agar (LCA) and potato dextrose agar (PDA) have been pointed as the best media for culturing *Chaetomium* due to the flourishing growth of colonies and the appearance of all crucial taxonomical features (Sharma & Pandey, 2010). *Chaetomium sensu stricto* is characterized by the creation of globose, ellipsoidal to ovate or obovate perithecia, which most often are ostiolate, or non-ostiolate in some species. The teleomorph body wall is usually formed of *textura intricata* or *epidermoidea* stretch, or of *textura angularis* in some species; the perithecial outgrows are hypha-like, flexuous, undulate, coiled to simply branched or dichotomously branched, generally with verrucose stretch, and soft in some species. The asci are clavate or fusiform with eight biseriate or irregularly arranged sexual spores. The ascospores are limoniform to globose (erratical in a few species), bilaterally flattened, and usually more than 7 µm in length. An *Acremonium*-like asexual morph is formed in some species (Udagawa, 1960; Wang et al., 2016a, 2016b; Whiteside, 1957). Indeed, recent phylogenetic studies have shown that several fungi belong to the Chaetomiaceae which are only known for their asexual morphs and were previously classified in the genera *Acremonium*, *Staphylotrichum*, *Humicola*, or *Trichocladium* (von Arx et al., 1984; Wang et al., 2016a, 2016b).

Considering the microscopic features of the teleomorph for defining the species concept in *Chaetomium* has proven to be challenging, since they are widely evanescent and disappear before maturation of the sexual spores. Therefore, it is usually recommended to observe hyaline ascospores within asci. For spotting asci, diligent attention should be paid to the creation of ascomata. It is relevant to make slides from young perithecium at the precociousness of the culture, usually within two weeks, which we also did to identify our strains. When the specimens are getting mature for a longer time, most perithecia become matured and this makes it difficult to notice asci. For species that form non-ostiolate sexual bodies, hyaline young perithecia are mainly a good choice for ascus studies (Abdel-Azeem, 2020; Berkson, 1966; Wang et al., 2022).

The high phylogenetic heterogeneity of Chaetomiaceae determined the necessity to reclassify all taxa employing molecular tools. The ITS region of rDNA is recommended as the best primary barcode for identification and grouping of *Chaetomium* species (Lee & Hanlin, 1999). However, it could be assisted by a secondary barcode, and LSU (D1/D2 domains of the 28S nrDNA), *rpb2* (partial RNA polymerase II second largest subunit), and *tub2* (β-tubulin) sequences were proposed as additional barcodes. Nowadays, fifty genera incorporating 275 species are recognized in Chaeto-

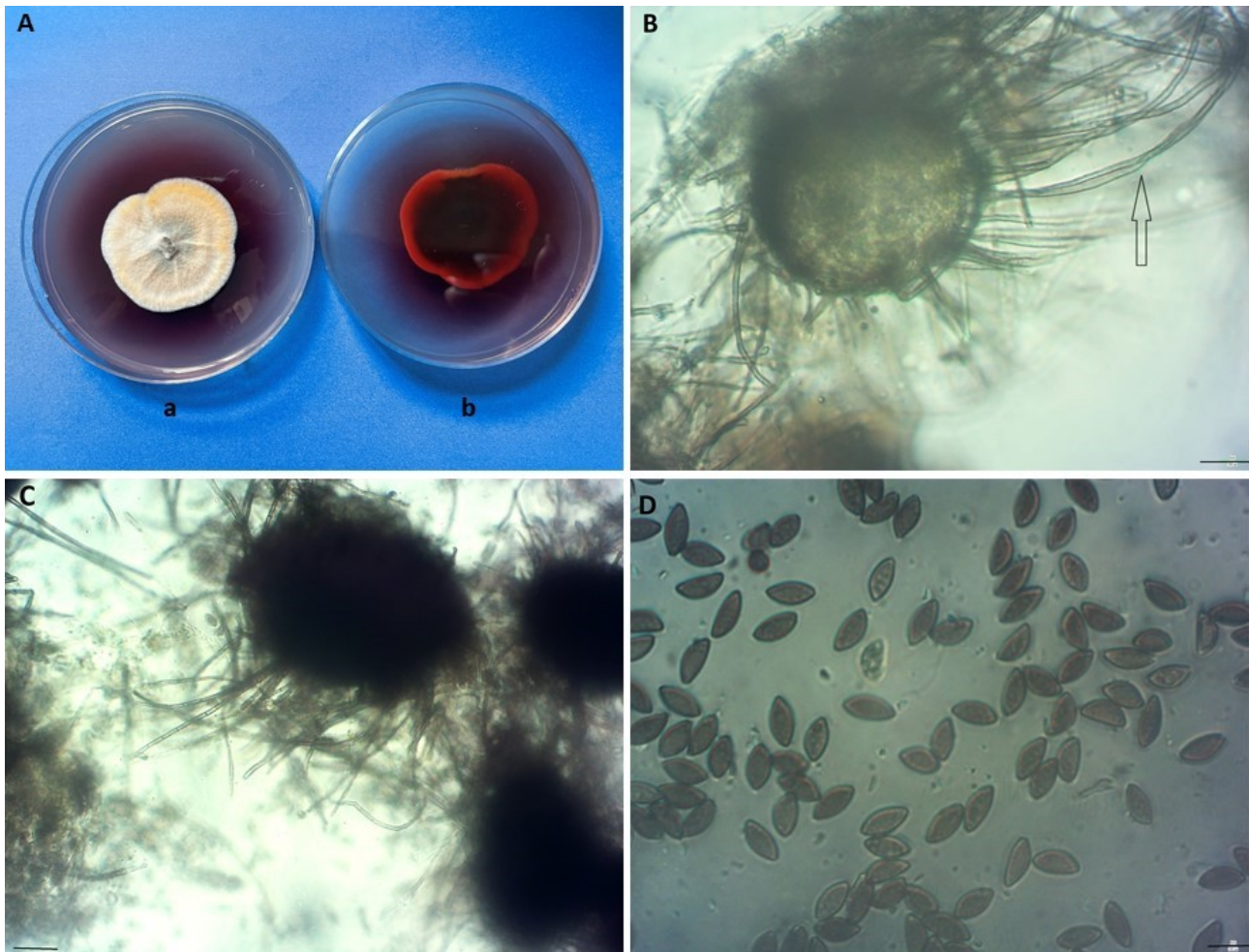


Figure 1 *Arcopilus aureus* Tv 121. (A) colony on PDA (a-averse, b-reverse); (B) young perithecium with a thin wall and characteristic arcuate hairs (arrow); (C) old perithecium; (D) ascospores. Scale bars = 10 µm.

miaceae; *Chaetomium* is the largest genus, accommodating 43 species (Wang et al., 2022).

Since Gustav Kunze officially introduced the genus *Chaetomium* (Kunze & Schmidt, 1817), remarkable progress has been made in the taxonomy of Chaetomiaceae. The comprehensive usage of molecular tools addressed the phylogenetic relationships for taxa with similar morphological features, with the imagery of five new genera, including *Arcopilus*; this name refers to the arcuate terminal outgrowths of perithecia which characterize most of the members in this genus, including the type species *A. aureus* (Wang et al., 2016a).

Despite the revolution following the employment of molecular tools, classical methods of identification should still be considered as the first step for classifying new strains. Our morphological observations were made after culturing the endophytic isolates of *A. aureus* from secondary branches of hazelnut Tv121 (from Motycz near Lublin, Poland) and A2269N (from the Astroni Nature Reserve near Napoli, Italy) on PDA (Difco) in darkness at 22 °C. After 7 days, the colonies were 30–36 mm in diameter, depicting yellow, orange to red color with red pigment diffusing into the medium (Figure 1A). The perithecia were 75–155 µm high and 80–150 µm in diameter, with an initially bright and thin wall (Figure 1B), then becoming brown and thick (Figure 1C). Characteristic arcuate hairs were observed, with an incurved apical part, circinate to coiled (Figure 1B). The ascospores were dark brown when mature, fusiform, elongate fusiform or limoniform, sometimes bilaterally flattened, with one or two apical germ pores, (8–10.5(–11) × (5–6(–6.5) µm) at both ends (Figure 1D).

Identification of isolate Tv121 was also carried out through rDNA-ITS sequencing. To this end, total genomic DNA extraction from young mycelium taken from pure culture, PCR amplification, and sequencing were performed according to a previously published protocol (Zimowska et al., 2021). The original small subunit ribosomal RNA gene and rDNA-ITS (ITS1 – 5.8S – ITS2) sequence sourced in this study has been deposited in GenBank with the reference number OR178408. The identification of the fungal symbiont was acknowledged by checking the homology of this sequence with other sequences available in GenBank.

3. Occurrence and ecological roles

With a worldwide distribution, species of the family Chaetomiaceae display elevated phenotypical and ecological diversity; they are mostly saprobic and occur in compost, soil, seed, dung, indoor environments, and rotting plant materials (Samson et al., 2019; Wang et al., 2016a). For its part, *A. aureus* has been diffusely mentioned from soil, dung, and other miscellaneous substrates (Abdullah & Azzo, 2015; Doveri, 2016; Kubatova, 2006; Kuter et al., 1983; Lee et al., 2019; Nakashima et al., 1991a, 1991b; Pornsuriya et al., 2010; Price et al., 1994; Quyet et al., 2018; Taniguchi et al., 1984; von Arx et al., 1986; Wang et al., 2013, 2016a). It has even been recovered from highly radioactive polluted soil and forest litter of the prohibited zone around the Chernobyl nuclear power plant (Ukraine) (Zhdanova et al., 2001) and from lead contaminated soil in Brazil (da Silva et al., 2018).

More significantly, *A. aureus* is characterized by a widespread occurrence as a plant associate. In fact, it has been reported from seeds of pepper and cucumber in the United States (Skolko & Groves, 1953), oats in Kansas (USA) and Canada (Conners, 1967; Hansing & Hartley, 1966), maize in the Chinese province of Liaoning (Gao et al., 2005), roots of sugarcane in Cuba (Hernández-Gutiérrez et al., 1995) and China (Raza et al., 2019), and bark of the cotton tree (*Bombax ceiba*) in Pakistan (Abbas et al., 2017). Some concern has been raised in China for its possible pathogenic aptitude, based on reports as a leaf spot agent on leaves of pineapple (*Ananas comosus*) in Hainan (Luo et al., 2012) and on the medicinal herb *Pseudostellaria heterophylla* in Guizhou (Yuan et al., 2021).

Moreover, this species has been identified as an endophyte in omnifarious geographic and ecological contexts. Particularly, there are records from stalks of bullgrass (*Paspalum fasciculatum*) in Costa Rica (Danielsen & Jensen, 1999), petioles of the Chinese fan palm (*Livistona chinensis*) in Thailand (Jiao et al., 2016), black pine (*Pinus nigra*) in Spain (Martínez-Álvarez et al., 2016), various organs of grapevine in Brazil (de Faria Silva et al., 2022) and India (Dwibedi & Saxena, 2018), and from leaves and stems of the medicinal shrub *Phlogacanthus thyrsoformis* in India (Sharma et al., 2020). More findings have been reported from Yunnan (China), from leaves of *Vaccinium dunalianum* (Fan et al., 2020) and roots of the orchid *Cypripedium flavum* (Zang et al., 2004). In this region, an isolate from another orchid (*Gastrodia elata*) was identified as *Arcopilus* sp. (Duan et al., 2022); however, a blast in GenBank of its ITS sequence shows 100% homology with 18 isolates of *A. aureus*, besides an isolate of *Arcopilus globulus*. This latter match may have raised doubts in the authors about the correctness of the classification, which were dispelled considering that this species is now designated as a synonym of *A. aureus* (Wang et al., 2022). In a survey carried out on the longbranch frostweed (*Crocyanthemum canadense*) in Nova Scotia (Canada), *A. aureus* was found to represent the most frequent endophytic associate colonizing every kind of tissues (root, leaf, lower and upper stem) in 56% of the examined plants collected at five out of six sites (Byers et al., 2021). Not too far geographically, this species was previously reported as one of four fungi colonizing roots of all the three plant species examined in the pine barrens of New Jersey, i.e. pitch pine (*Pinus rigida*), switchgrass (*Panicum virgatum*), and rosette grass (*Dichanthelium acuminatum*), implying a likely aptitude for a horizontal spread among plants thriving in the same ecosystem (Luo et al., 2017).

Arcopilus aureus is also part of the endophytic communities of trees of notable economic importance, such as olive (*Olea europaea*) (Nicoletti et al., 2020) and chestnut (*Castanea sativa*) (Nicoletti et al., 2021). An endophytic strain from the latter plant has

been identified in Portugal as a prospective antagonist in the control of bark canker caused by *Cryphonectria parasitica* (Coelho et al., 2022). Moreover, an endophytic isolate from *P. nigra* reduced the disease progress resulting after inoculation of the canker agent *Fusarium circinatum* on *Pinus radiata* seedlings, indicating that it can find application in biological control of this pathogen (Martínez-Álvarez et al., 2016).

Another strain of *A. aureus* was previously identified as an effective biocontrol agent of the rice blast pathogen *Magnaporthe grisea* and the sheath blight pathogen *Rhizoctonia solani* both *in vitro* and *in vivo*. Particularly, its metabolites suppressed mycelial growth in *R. solani* and germination of spores and appressorium formation in *M. grisea*; moreover, they minimized the disease indexes of rice blast in the field and rice sheath blight both in the greenhouse and in the field (Wang et al., 2013). Culture filtrates of an isolate from *Guzmania* displayed strong inhibitory effects against *Phytophthora infestans*, *Bipolaris sorokiniana*, *Fusarium culmorum*, and *F. oxysporum*, while the bioactivity was moderate against *Botrytis cinerea*, *B. fabae*, *Pyrenophora graminea*, and *Neocosmospora solani*. These inhibitory effects persisted in the ethyl acetate extract of the culture filtrates (Linkies et al., 2021). Ethyl acetate, hexane, and methanolic extracts obtained from cultures of a soil strain determined some extent of inhibition in oospore formation in *Pythium aphanidermatum* (Pornsuriya et al., 2010). Finally, a Chinese strain of *A. aureus* was able to inhibit mycelial growth of *Phytophthora capsici* in dual cultures *in vitro* and to induce coiling of pathogen's hyphae. Moreover, its fermentation broth and crude extract also almost completely inhibited mycelial growth of *P. capsici*, while treatment of pepper plants with fermentation broth activated defense enzymes, such as phenylalanine ammonia lyase, peroxidase, polyphenol oxidase, increasing their resistance. In addition, the fermentation broth significantly promoted growth of pepper seedlings, leading to an increase in fresh and dry weight (Liu et al., 2013).

4. Secondary metabolites and other biotechnological aspects

By reason of the recent separation from *Chaetomium*, the production of secondary metabolites by *A. aureus* has been considered and reviewed together with other species of this genus (Zhang et al., 2012). Particularly, a huge research activity has been carried out on the biochemical properties of *C. globosum*; over 200 secondary metabolites have been isolated and identified from isolates of this species, including terpenoids, alkaloids, tetramic acids, diketopiperazines, steroids, xanthenes, bis(3-indolyl)-benzoquinones, azaphilones, anthraquinones, pyranones, and orsellides, and many of them present interesting properties, such as anticancer, antimicrobial, antimalarial, cytotoxic, and antiviral activities (Dwibedi et al., 2023; Rao et al., 2023). In general, these fungi exhibit potent antimicrobial activities that encourage their use as biological control agents.

The data concerning secondary metabolites of *A. aureus* are more limited, and no information about this aspect was reported in the fundamental revision by Wang et al., (2016a). However, the analysis of the available literature indicates that the first bioactive secondary metabolite identified as a product of a strain of this species was oosporein (3,3',6,6'-tetrahydroxy-5,5'-dimethyl-2,2'-bi-*p*-benzoquinone) (Lloyd et al., 1955). This compound, responsible for the red pigmentation of *A. aureus* cultures, was previously known as a product of other fungi, and it has been investigated as a phytotoxin and a mycotoxin affecting poultry (Cole et al., 1974; Manning & Wyatt, 1984); later on, it has been shown to exert antiviral and antifungal effects (Nagaoka et al., 2004; Terry et al., 1992), with the latter reported to increase at acidic pH (Taniguchi et al., 1984). Although it was not directly extracted, it was considered as likely responsible for the antifungal effect of culture filtrates and extracts from the strain examined in the previously mentioned work by Linkies et al. (2021). Moreover, it has been reported to exert protective effects in plants against aluminum stress based on its chelating properties (Haruma et al., 2022).

Another quinone derivative, cochlioquinol II, first reported as a phytotoxin from the Bermuda grass pathogen *Bipolaris cynodontis* (Lim, 1998), was found to be produced by an endophytic isolate from grapevine and reported to be responsible for the yellow pigmentation of this strain, along with riboflavin (de Faria Silva et al., 2022). Better

known as pigments and for possible application as food colorants are compounds belonging to the polyketide class of azaphilones (Pimenta et al., 2021), which were discovered as products of an endophytic strain isolated in Morocco; particularly, isochromophilone IV and VII, sclerotioramin and sclerotiorin were extracted from cultures of this strain along with the tricyclic compound SB236050 and the new resorcinol derivative chaetorcinol (Kabbaj et al., 2015). Extracted from an endophytic strain from *G. elata* along with the biosynthetic analogs maristachone B, 8-zinniol methyl ether, and the novel arcopiniols A-C (Duan et al., 2022), zinniol is another product which was first characterized as a phytotoxin from several *Alternaria* and *Phoma* species (Cotty & Misaghi, 1984; Sugawara & Strobel, 1986); however, this biological effect has been questioned after more recent experimental assessments (Qui et al., 2010).

It is widely reported that many endophytic fungi can synthesize *in vitro* products originally characterized from their host plants (Nicoletti & Fiorentino, 2015). Among other endophytic fungi isolated from grapevine, a strain of *A. aureus* was characterized as the best resveratrol producer in an Indian study (Dwibedi & Saxena, 2018). This finding might have prompt biotechnological application, considering the notable beneficial effects of this polyphenolic flavonoid on human health, as well as its use as a pharmacophore.

Finally, an unidentified product ($C_{31}H_{35}O_8N$), obtained from a strain classified as *Chaetomium laterale* var. *diporum* (currently a synonym of *A. aureus*), was discovered to almost totally inhibit mycelial growth of widespread pathogens, such as *Sclerotinia sclerotiorum* and *Botrytis cinerea*, at the concentration of $10 \mu\text{g mL}^{-1}$; however, its antifungal activity was much lower when assessed against other plant pathogenic fungi (Nakashima et al., 1991a, 1991b).

The limited extent of the available information concerning secondary metabolites of *A. aureus* calls for further insights, also in the aim of assessing possible chemotaxonomic relevance supporting species discrimination. Some of the above products were reported to exert phytotoxic effects; however, the spectrum of their bioactivity should be determined to establish if this property is general or specific, as well as if the doses released *in vivo* can effectively induce damage to the host plants. Moreover, clues concerning the fungitoxic effects also call for further assessments of the possible role of some products in defensive mutualism.

Besides the bioactive properties of secondary metabolites, biotechnological applications concerning isolates of *A. aureus* have not been substantially exploited so far. Some isolates have been reported as sources of xylanases (Iizuka & Kawaminami, 1969), cellulases (Ghora & Chaudhuri, 1975), and trehalase (Sumida et al., 1989), which is not so remarkable, considering that these are general properties of microbial strains recovered from any kind of environmental and geographical context. With reference to bioremediation, an isolate of *A. aureus* isolated in Brazil from semiarid soil with lead concentrations above regulatory limits exhibited better performance in laboratory assays in terms of fungal growth in a lead-containing substrate, capability to use pectin as a carbon origin, as well as basophilic and thermotolerant properties. Particularly, a reduction of free Pb in soil (61 and 54%, respectively) could be detected 60 days after inoculation. These findings support the use of this species as a potential tool for the bioremediation of contaminated sites (da Silva et al., 2018). Moreover, it has been proved to be highly tolerant to glyphosate (Hu et al., 2005).

5. Conclusions

The examination of the available literature concerning *A. aureus* depicts its widespread geographic distribution and frequent occurrence as a plant symbiont, particularly in the form of an endophytic associate. In this respect, even the limited available data are indicative of a good adaptation to this ecological role, considering that it has been recovered from both roots and the above-ground parts of plants of various taxonomic affiliations. Some clues of the capacity to spread horizontally within heterogeneous ecological contexts portend an even wider and more pervasive occurrence, which stimulates further insights aiming at a more thorough assessment of its symbiotic interactions with the host.

The present paper represents the first report of the endophytic association of *A. aureus* with *C. avellana*, as inferable from the previous reports summarized in our recent review on endophytic fungi of this plant (Nicoletti & Zimowska, 2023). Indeed, the obtainable data on the occurrence and functions of endophytic fungi of hazelnut are scanty, and a good part of them refers to countries outside the European Union, such as China and Iran. Some published studies mainly concern the perspective of a pharmaceutical application, starting from the finding that a hazelnut endophytic strains of *Penicillium aurantiogriseum* is able to synthesize the antitumor drug taxol in axenic cultures (Yang et al., 2014). Ongoing studies concerning the metabolomic profile of our strains of *A. aureus* could bring new outcomes regarding the potential biotechnological exploitation of this fungus.

Most importantly, the association of *A. aureus* with hazelnut calls for further assessments in terms of its role in defensive mutualism. The antagonistic properties resulting from some published studies and our preliminary evidences support the possible implication of this fungus in protection of hazelnut from biotic adversities. Indeed, this crop is expected to face hard times in this respect after the announcement of phase-out of copper products in the European Union (Nicoletti et al., 2022). Hence, all components of the associated microbiome should be carefully evaluated for their contribution to plant health and fitness. In this respect, it is quite relevant considering that *Chaetomium*-like fungi have been already introduced in the management of plant diseases, and even some commercial formulations (e.g. Ketomium®) have been released in several countries (Soytong et al., 2001).

References

- Abbas, S. Q., Noureen, H., Abbas, E., Abbas, A., & Iftakhar, T. (2017). New fungal record on *Bombax ceiba* Linn. from Pakistan. III. *International Journal of Biology and Biotechnology*, 14(1), 69–74.
- Abdel-Azeem, A. M. (2020). *Recent developments on genus Chaetomium*. Springer International Publishing.
- Abdullah, S. K., & Azzo, N. M. (2015). Two new records of *Chaetomium* species isolated from soil under grapevine plantations and a checklist of the genus in Iraq. *Journal of Agricultural Technology*, 11(7), 1515–1522. <http://www.ijat-aatsea.com>
- Ames, L. M. (1961). *A monograph of the Chaetomiaceae* (No. 2). Army Research Office.
- Berkson, B. M. (1966). Cytomorphological studies of the ascogenous hyphae in four species of *Chaetomium*. *Mycologia*, 58(1), 125–130.
- Byers, P. R., Evans, R. C., Migicovsky, Z., & Walker, A. K. (2021). Fungal symbionts of endangered *Crocianthemum canadense* (Cistaceae) in Nova Scotia. *Botany*, 99(7), 403–419. <https://doi.org/10.1139/cjb-2020-0187>
- Chivers, A. H. (1915). Preliminary diagnoses of new species of *Chaetomium*. *Proceedings of the American Academy of Arts and Sciences*, 48(4), 83–88.
- Coelho, V., Machado, J. M., Guerra, I., & Gouveia, M. E. (2022). *In vitro* antagonism evaluation of endophyte *Chaetomium aureum* against *Cryphonectria parasitica*. In *9th International Conference on Agriculture, Animal Sciences and Rural Development*. ISPEC Publishing House.
- Cole, R. J., Kirksey, J. W., Cutler, H. G., & Davis, E. E. (1974). Toxic effects of oosporein from *Chaetomium trilaterale*. *Journal of Agricultural and Food Chemistry*, 22, 517–520.
- Connors, I. L. (1967). *An annotated index of plant diseases in Canada and fungi recorded on plants in Alaska, Canada and Greenland* (Vol. 1251, pp. 1–381). Research Branch Canada Department of Agriculture Publication.
- Cotty, P. J., & Misaghi, I. J. (1984). Zinniol production by *Alternaria* species. *Phytopathology*, 74(7), 785–788.
- Danielsen, S., & Jensen, D. F. (1999). Fungal endophytes from stalks of tropical maize and grasses: Isolation, identification, and screening for antagonism against *Fusarium verticillioides* in maize stalks. *Biocontrol Science and Technology*, 9(4), 545–553.
- da Silva Júnior, F. M. R., Volcão, L. M., Hoscha, L. C., & Pereira, S. V. (2018). Growth of the fungus *Chaetomium aureum* in the presence of lead: Implications in bioremediation. *Environmental Earth Sciences*, 77, Article 275. <https://doi.org/10.1007/s12665-018-7447-x>
- de Faria Silva, L. A., Ferreira Alves, M., Florêncio Filho, D., Takahashi, J. A., Soares Santos, L., & Almeida De Carvalho, S. (2022). Pigment produced from *Arcopilus aureus* isolated from grapevines: Promising natural yellow colorants for the food industry. *Food Chemistry*, 389, Article 132967. <https://doi.org/10.1016/j.foodchem.2022.132967>

- Dimitrova, N., & Nacheva, L. (2021). An optimized micropropagation protocol by ex vitro rooting of Pear rootstock OHF 333 (*Pyrus communis* L.). *Acta Agrobotanica*, 74, Article 744. <https://doi.org/10.5586/aa.744>
- Doveri, F. (2016). Description of *Chaetomium aureum*, *Corynascus sepedonium* and *Coniochaeta hansenii* newly recorded from Italy and a key to coprophilous Chaetomiaceae and Coniochaetaceae. *Ascomycete.org*, 8(1), 7–24.
- Duan, H. J., Xie, F., Xia, D. D., Xie, T. P., Zhang, Y., Sun, Y., Lu, J. M., Zhou, H., & Ding, Z. T. (2022). Zinniol analogues from the endophytic fungus *Arcopilus* sp. YUD20001 associated with *Gastrodia elata*. *Phytochemistry Letters*, 51, 140–144. <https://doi.org/10.1016/j.phytol.2022.08.014>
- Dwibedi, V., Rath, S. K., Jain, S., Martínez-Argueta, N., Prakash, R., Saxena, S., & Rios-Solis, L. (2023). Key insights into secondary metabolites from various *Chaetomium* species. *Applied Microbiology and Biotechnology*, 107(4), 1077–1093. <https://doi.org/10.1007%2Fs00253-023-12365-y>
- Dwibedi, V., & Saxena, S. (2018). *Arcopilus aureus*, a resveratrol-producing endophyte from *Vitis vinifera*. *Applied Biochemistry and Biotechnology*, 186, 476–495. <https://doi.org/10.1007/s12010-018-2755-x>
- Ekbic, H. B., Yaman, E., Özenç, D. B., & Ekbic, E. (2022). Effects of hazelnut husk compost and tea residue compost on quality and performance of 5 BB American grapevine rootstock saplings. *Acta Scientiarum Polonorum Hortorum Cultus*, 21(5), 15–23.
- Fan, M., Chen, X., Luo, X., Zhang, H., Liu, Y., Zhang, Y., Wu, J., Zhao, C., & Zhao, P. (2020). Diversity of endophytic fungi from the leaves of *Vaccinium dunalianum*. *Letters in Applied Microbiology*, 71(5), 479–489. <https://doi.org/10.1111/lam.13345>
- Gakuubi, M. M., Munusamy, M., Liang, Z. X., & Ng, S. B. (2021). Fungal endophytes: A promising frontier for discovery of novel bioactive compounds. *Journal of Fungi*, 7(10), Article 786. <https://doi.org/10.3390/jof7100786>
- Gao, X., Lu, G., Sun, X., Liu, S., & Chen, R. (2005). Diversity of corn seed-borne fungi. *Journal of Fungal Research*, 3(2), 42–46.
- Ghora, B. K., & Chaudhuri, K. L. (1975). Studies on the adaptive nature of cellulolytic enzyme from *Chaetomium aureum* Chivers. *Experientia*, 31, 147–148.
- Hansing, E. D., & Hartley, A. (1966). Oat seed fungi and their control. *Proceedings of the Association of Official Seed Analysts*, 56, 143–147. <http://www.jstor.org/stable/23432071>
- Haruma, T., Doyama, K., Lu, X., Arima, T., Igarashi, T., Tomiyama, S., & Yamaji, K. (2022). Oosporein produced by root endophytic *Chaetomium cupreum* promotes the growth of host plant, *Miscanthus sinensis*, under aluminum stress at the appropriate concentration. *Plants*, 12(1), Article 36. <https://doi.org/10.3390/plants12010036>
- Hernández-Gutiérrez, A., Mena Portales, J., Herrera Figueroa, S., López Mesa, M. O., Mercado Sierra, A., Ortiz Medina, J. L., & Rodríguez Morejón, K. (1995). Additions to the mycobiota of sugarcane (*Saccharum* sp. hybrid) in Cuba. II. *Revista Iberoamericana de Micología*, 12(3), 68–69.
- Hu, B., Hsieh, H., Yao, J., & Fu, C. (2005). Study on the tolerance of some soil fungi to glyphosate. *Journal of the Experimental Forest of National Taiwan University*, 19(1), 17–23.
- Iizuka, H., & Kawaminami, T. (1969). Studies on xylanase from microorganisms: Part II. Isolation and selection of xylanase-producing microorganisms and the identification of a new species of *Streptomyces*. *Agricultural and Biological Chemistry*, 33(9), 1257–1263.
- Jiaojiao, S., Wattanachai, P., & Kasem, S. (2016). Isolation and identification of endophytic fungi from 10 species palm trees. *Journal of Agricultural Technology*, 12(2), 349–363. <http://www.ijat-aatsea.com>
- Kabbaj, F. Z., Lu, S., Faouzi, M. E. A., Meddah, B., Proksch, P., Cherrah, Y., Altenbach, H. J., Aly, A. H., Chadli, A., & Debbab, A. (2015). Bioactive metabolites from *Chaetomium aureum*: Structure elucidation and inhibition of the Hsp90 machine chaperoning activity. *Bioorganic & Medicinal Chemistry*, 23(1), 126–131. <https://doi.org/10.1016%2Fj.bmc.2014.11.021>
- Kubátová, A. (2006). *Chaetomium* in the Czech Republic and notes to three new records. *Czech Mycology*, 58(3-4), 155–171. <https://doi.org/10.33585/cmy.58301>
- Kunze, G., & Schmidt, J. K. (1817). *Chaetomium*. *Mykologische Hefte nebst einem allgemein-botanischen Anzeiger*. Voss.
- Kuter, G. A., Nelson, E. B., Hoitink, H. A. J., & Madden, L. V. (1983). Fungal populations in container media amended with composted hardwood bark suppressive and conducive to *Rhizoctonia* damping-off. *Phytopathology*, 73(10), 1450–1456.
- Lee, S., & Hanlin, R. T. (1999). Phylogenetic relationships of *Chaetomium* and similar genera based on ribosomal DNA sequences. *Mycologia*, 91(3), 434–442. <https://doi.org/10.1080/00275514.1999.12061037>

- Lee, S. H., Park, H. S., Nguyen, T. T., & Lee, H. B. (2019). Characterization of three species of Sordariomycetes isolated from freshwater and soil samples in Korea. *Mycobiology*, 47(1), 20–30. <https://doi.org/10.1080/12298093.2019.1574372>
- Lim, C. H. (1998). Structures and biological activities of phytotoxins produced by the plant pathogenic fungus *Bipolaris cynodontis* cynA. *Journal of Pesticide Science*, 23(3), 281–288. <https://doi.org/10.1584/jpestics.23.281>
- Linkies, A., Jacob, S., Zink, P., Maschmer, M., Maier, W., & Koch, E. (2021). Characterization of cultural traits and fungicidal activity of strains belonging to the fungal genus *Chaetomium*. *Journal of Applied Microbiology*, 131(1), 375–391. <https://doi.org/10.1111/jam.14946>
- Liu, Y., Yin, C., Tian, Y., Liu, X., Zhang, X., & Gao, K. (2013). Identification of the antagonistic fungus strain HTC and its potential for biocontrol of pepper phytophthora blight. *Acta Phytophylacica Sinica*, 40(5), 437–444.
- Lloyd, G., Robertson, A., Sankey, G. B., & Whalley, W. B. (1955). The chemistry of fungi. Part XXV. Oosporein, a metabolite of *Chaetomium aureum* Chivers. *Journal of the Chemical Society*, 2163–2165.
- Luo, Z., Fan, H., Li, X., Liu, Y., Hua, M., Wang, X., Yu, N., Zhou, W., & He, F. (2012). Study on several fungal leaf diseases of pineapple of Hainan. *Southwest China Journal of Agricultural Sciences*, 25(5), 1703–1707.
- Luo, J., Walsh, E., Miller, S., Blystone, D., Dighton, J., & Zhang, N. (2017). Root endophytic fungal communities associated with pitch pine, switchgrass, and rosette grass in the pine barrens ecosystem. *Fungal Biology*, 121(5), 478–487. <https://doi.org/10.1016/j.funbio.2017.01.005>
- Malinowski, D. P., & Belesky, D. P. (2019). Epichloë (formerly Neotyphodium) fungal endophytes increase adaptation of cool-season perennial grasses to environmental stresses. *Acta Agrobotanica*, 72(2), Article 1767. <https://doi.org/10.5586/aa.1767>
- Manning, R. O., & Wyatt, R. D. (1984). Comparative toxicity of *Chaetomium* contaminated corn and various chemical forms of oosporein in broiler chicks. *Poultry Science*, 63(2), 251–259.
- Martínez-Álvarez, P., Fernández-González, R. A., Sanz-Ros, A. V., Pando, V., & Diez, J. J. (2016). Two fungal endophytes reduce the severity of pitch canker disease in *Pinus radiata* seedlings. *Biological Control*, 94, 1–10. <https://doi.org/10.1016/j.biocontrol.2015.11.011>
- Nagaoka, T., Nakata, K., Kouno, K., & Ando, T. (2004). Antifungal activity of oosporein from an antagonistic fungus against *Phytophthora infestans*. *Zeitschrift für Naturforschung C*, 59(3-4), 302–304.
- Nakashima, N., Moromizato, Z., & Matsuyama, N. (1991a). *Chaetomium* spp., antagonistic microorganisms to phytopathogenic fungi. *Journal of the Faculty of Agriculture, Kyushu University*, 36(1-2), 109–111.
- Nakashima, N., Moromizato, Z., & Matsuyama, N. (1991b). The antifungal substance produced by *Chaetomium trilaterale* var. *diporum* RC-5 isolated from sclerotia of *Sclerotinia sclerotiorum*. *Japanese Journal of Phytopathology*, 57(5), 657–662.
- Nicoletti, R., Beccaro, G. L., Sekara, A., Cirillo, C., & Di Vaio, C. (2021). Endophytic fungi and ecological fitness of chestnuts. *Plants*, 10(3), Article 542. <https://doi.org/10.3390/plants10030542>
- Nicoletti, R., Di Vaio, C., & Cirillo, C. (2020). Endophytic fungi of olive tree. *Microorganisms*, 8(9), Article 1321. <https://doi.org/10.3390/microorganisms8091321>
- Nicoletti, R., & Fiorentino, A. (2015). Plant bioactive metabolites and drugs produced by endophytic fungi of Spermatophyta. *Agriculture*, 5, 918–970. <https://doi.org/10.3390/agriculture5040918>
- Nicoletti, R., Petriccione, M., Curci, M., & Scortichini, M. (2022). Hazelnut-associated bacteria and their implications in crop management. *Horticulturae*, 8(12), Article 1195. <https://doi.org/10.3390/horticulturae8121195>
- Nicoletti, R., & Zimowska, B. (2023). Endophytic fungi of hazelnut (*Corylus avellana*). *Plant Protection Science*, 59(2), 107–123. <https://doi.org/10.17221/133/2022-PPS>
- Pimenta, L. P., Gomes, D. C., Cardoso, P. G., & Takahashi, J. A. (2021). Recent findings in azaphilone pigments. *Journal of Fungi*, 7(7), Article 541. <https://doi.org/10.3390/jof7070541>
- Plomley, N. J. B. (1959). Formation of the colony in the fungus *Chaetomium*. *Australian Journal of Biological Sciences*, 12(1), 53–64.
- Pornsuriya, C., Soyong, K., Kanokmedhakul, S., & Lin, F. C. (2010). Efficacy of antifungal metabolites from some antagonistic fungi against *Pythium aphanidermatum*. *Journal of Agricultural Technology*, 6(2), 299–308. <http://www.ijat-rmutto.com>
- Price, D. L., Simmons, R. B., Ezeonu, I. M., Crow, S. A., & Ahearn, D. G. (1994). Colonization of fiberglass insulation used in heating, ventilation and air conditioning systems. *Journal of Industrial Microbiology*, 13, 154–158.

- Qui, J. A., Castro-Concha, L. A., García-Sosa, K., Miranda-Ham, M. L., & Peña-Rodríguez, L. M. (2010). Is zinniol a true phytotoxin? Evaluation of its activity at the cellular level against *Tagetes erecta*. *Journal of General Plant Pathology*, 76, 94–101. <https://doi.org/10.1007/s10327-010-0222-9>
- Quyet, N. T., Thanh, N. D., Cuong, H. V., Huong, B. T. L., & Soyong, K. (2018). Molecular identification of *Chaetomium* species from soil in Vietnam. *International Journal of Agricultural Technology*, 14(2), 225–232.
- Rao, Q. R., Rao, J. B., & Zhao, M. (2023). Chemical diversity and biological activities of specialized metabolites from the genus *Chaetomium*: 2013–2022. *Phytochemistry*, 210, Article 113653. <https://doi.org/10.1016/j.phytochem.2023.113653>
- Raza, M., Zhang, Z. F., Hyde, K. D., Diao, Y. Z., & Cai, L. (2019). Culturable plant pathogenic fungi associated with sugarcane in southern China. *Fungal Diversity*, 99, 1–104. <https://doi.org/10.1007/s13225-019-00434-5>
- Rokas, A., Mead, M. E., Steenwyk, J. L., Raja, H. A., & Oberlies, N. H. (2020). Biosynthetic gene clusters and the evolution of fungal chemodiversity. *Natural Product Reports*, 37(7), 868–878. <https://doi.org/10.1039/C9NP00045C>
- Rong, I. H., & van Warmelo, T. (1988). Ascosporeogenesis in some selected species of *Chaetomium*. *Phytophylactica*, 20(4), 353–356.
- Samson, R. A., Houbraken, J., Thrane, U., Frisvad, J. C., & Andersen, B. (2019). *Food and indoor fungi*. Westerdijk Fungal Biodiversity Institute.
- Sharma, G., & Pandey, R. R. (2010). Influence of culture media on growth, colony character and sporulation of fungi isolated from decaying vegetable wastes. *Journal of Yeast and Fungal Research*, 1(8), 157–164. <http://www.academicjournals.org/JYFR>
- Sharma, R., Tangjang, S., & Shukla, A. C. (2020). Diversity of fungal endophytes from *Phlogacanthus thyrsoformis* (Roxb. ex Hardw.) Mabb., a promising medicinal plant in Arunachal Pradesh, northeast India. *The Journal of Indian Botanical Society*, 99(1-2), 46–55. <https://doi.org/10.5958/2455-7218.2020.00004.2>
- Skolko, A. J., & Groves, J. W. (1953). Notes on seed-borne fungi. VII *Chaetomium*. *Canadian Journal of Botany*, 31, 779–809.
- Soyong, K., Kanokmedhakul, S., Kukongviriyapa, V., & Isobe, M. (2001). Application of *Chaetomium* species (Ketomium) as a new broad spectrum biological fungicide for plant disease control. *Fungal Diversity*, 7, 1–15.
- Sugawara, F., & Strobel, G. (1986). Zinniol, a phytotoxin, is produced by *Phoma macdonaldii*. *Plant Science*, 43(1), 19–23.
- Sumi, A. A., Khayer, A., Jhan, P. K., & Rubel, M. H. (2022). Medicinal plants as biocontrol agents: An in vitro study on black rot pathogen of cabbage (*Xanthomonas campestris* pv. *campestris*). *Acta Agrobotanica*, 75, Article 759. <https://doi.org/10.5586/aa.759>
- Sumida, M., Ogura, S., Miyata, S., Arai, M., & Murao, S. (1989). Purification and some properties of trehalase from *Chaetomium aureum* MS-27. *Journal of Fermentation and Bioengineering*, 67(2), 83–86.
- Taniguchi, M., Kawaguchi, T., Tanaka, T., & Oi, S. (1984). Antimicrobial and respiration inhibitory activities of oosporein. *Agricultural and Biological Chemistry*, 48(4), 1065–1067.
- Terry, B. J., Liu, W. C., Cianci, C. W., Proszynski, E., Fernandes, P., Bush, K., & Meyers, E. (1992). Inhibition of herpes simplex virus type 1 DNA polymerase by the natural product oosporein. *Journal of Antibiotics*, 45, 286–288.
- Udagawa, S. I. (1960). A taxonomic study on the Japanese species of *Chaetomium*. *Journal of General and Applied Microbiology*, 6(4), 223–251.
- von Arx, J. A., Dreyfuss, M., & Müller, E. (1984). A reevaluation of *Chaetomium* and the Chaetomiaceae. *Persoonia*, 12(2), 169–179.
- von Arx, J. A., Guarro, J., & Figueras, M. J. (1986). The ascomycete genus *Chaetomium*. *Beihefte zur Nova Hedwigia*, 84, 1–162.
- Wang, X. W., Han, P. J., Bai, F. Y., Luo, A., Bensch, K., Meijer, M., & Houbraken, J. (2022). Taxonomy, phylogeny and identification of Chaetomiaceae with emphasis on thermophilic species. *Studies in Mycology*, 101, 121–243. <https://doi.org/10.3114/sim.2022.101.03>
- Wang, X. W., Houbraken, J., Groenewald, J. Z., Meijer, M., Andersen, B., Nielsen, K. F., & Samson, R. A. (2016a). Diversity and taxonomy of *Chaetomium* and *Chaetomium*-like fungi from indoor environments. *Studies in Mycology*, 84, 145–224. <https://doi.org/10.1016/j.simyco.2016.11.005>
- Wang, X. W., Lombard, L., Groenewald, J. Z., Li, J., Videira, S. I. R., Samson, R. A., & Crous, P. W. (2016b). Phylogenetic reassessment of the *Chaetomium globosum* species complex. *Persoonia*, 36, 83–133. <https://doi.org/10.3767/003158516X689657>
- Wang, Y. L., Liu, S. Y., Mao, X. Q., Zhang, Z., Jiang, H., Chai, R. Y., Qiu, H. P., Wang, J. Y., Du, X. F., Li, B., & Sun, G. C. (2013). Identification and characterization of rhizosphere

- fungal strain MF-91 antagonistic to rice blast and sheath blight pathogens. *Journal of Applied Microbiology*, 114(5), 1480–1490. <https://doi.org/10.1111/jam.12153>
- Whiteside, W. C. (1957). Perithecial initials of *Chaetomium*. *Mycologia*, 49(3), 420–425.
- Winter, G. (1885). *Ascomyceten: Gymnoasceen und Pyrenomyceten*. Dr. L. Rabenhorst's *Kryptogamen Flora von Deutschland, Oesterreich und der Schweiz* (pp. 1–928). Eduard Kummer, Zweite Auflage. <https://doi.org/10.5962/bhl.title.1356>
- Yang, Y., Zhao, H., Barrero, R. A., Zhang, B., Sun, G., Wilson, I. W., Xie, F., Walker, K. D., Parks, J. W., Bruce, R., Guo, G., Chen, L., Zhang, Y., Huang, X., Tang, Q., Liu, H., Bellgard, M. I., Qiu, D., Lai, J., & Hoffman, A. (2014). Genome sequencing and analysis of the paclitaxel-producing endophytic fungus *Penicillium aurantiogriseum* NRRL 62431. *BMC Genomics*, 15(1), 1–14. <http://www.biomedcentral.com/1471-2164/15/69>
- Yuan, Q. S., Wang, X., Wang, L., Ou, X., Jiang, W., Kang, C., Guo, L., & Zhou, T. (2021). First report of *Arcopilus aureus* causing leaf black spot disease of *Pseudostellaria heterophylla* in China. *Plant Disease*, 105(12), Article 4168. <https://doi.org/10.1094/PDIS-03-21-0647-PDN>
- Zang, M., Wang, R. L., & Hu, H. (2004). Bulbils exist in root of *Cypripedium flavum*. *Plant Diversity*, 26, 1–3. <https://journal.kib.ac.cn/EN/Y2004/V26/I01/1>
- Zhang, Q., Li, H. Q., Zong, S. C., Gao, J. M., & Zhang, A. L. (2012). Chemical and bioactive diversities of the genus *Chaetomium* secondary metabolites. *Mini Reviews in Medicinal Chemistry*, 12(2), 127–148. <https://doi.org/10.2174/138955712798995066>
- Zhdanova, N. N., Zakharchenko, V. A., Artyshkova, L. V., Schkolny, A. T., & Vasilevskaya, A. I. (2001). Mycobiota of radionuclides polluted soils of the limits zone of the ChNPP in 14 years after the accident. *Mikologiya i Fitopatologiya*, 35(6), 1–8.
- Zimowska, B., Becchimanzi, A., Krol, E. D., Furmanczyk, A., Bensch, K., & Nicoletti, R. (2021). New *Cladosporium* species from normal and galled flowers of Lamiaceae. *Pathogens*, 10(3), Article 369. <https://doi.org/10.3390/pathogens10030369>
- Zopf, W. (1881). *Zur Entwicklungsgeschichte der Ascomyceten: Chaetomium* (No. 5). Für die Akademie in Commission bei W. Engelmann in Leipzig.