G. P. NOVELLI

ROLE OF FREE RADICALS IN SEPTIC SHOCK

Institute of Anaesthesiology and Intensive Care of the University of Florence, Italy

The available data supporting the role of oxygen radicals (OR^0) as fundamental — though not unique — mediators in the pathogenesis of sepsis and septic shock are reviewed.

The main questions are 1) Are there any signs of OR^0 overgeneration during sepsis and septic shock? 2) Are there any indirect signs of OR^0 damage during sepsis and septic shock? 3) Does OR^0 inactivation affect the evolution and severity of sepsis and septic shock? 4) May the pathophysiology of sepsis and septick shock be viewed as a consequence of OR^0 overgeneration?

All these questions might receive affirmative answers; thus the relationships of OR^0 with other mediators of sepsis and septic shock are discussed.

It emerges that all the molecules which are being considered as mediators of sepsis and septic shock (cytokines, endothelins, platalet activating factor, complement) may act on granulocytes and other phagocytic cells to stimulate OR^0 production. There are also very close relationships with nitric oxide, another free radical.

The data in favour of an antioxidant therapy of sepsis and septic shock in humans are too few to be conclusive.

Key words: oxygen free radicals, septic shock, multiple organ failure, antioxidants.

INTRODUCTION

Oxygen radicals (OR^0) play a fundamental role in the complex pathogenesis of systemic inflammatory response syndrome (SIRS), sepsis, septic shock and multiple organ dysfunction syndrome (MODS). Their precise significance has not been completely clarified, probably due to the lack of methods for their direct detection and quantitation.

 OR^0 (O_2^- , H_2O_2 , OH^0 , O_2^*) are the highly reactive intermediates of the mono- or divalent reduction of molecular oxygen. Owing to the presence of an uncoupled electron in the outer orbital they react with the very nearest molecules, thus initiating a lot of biochemical processes which lead to profound disturbances of cell biochemistry such as lipid peroxidation, protein

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denaturation, polysaccharide depolymerization, genome oxidation, cellular and interstitial structure destabilization etc.

The generation of small amounts of OR^0 is a physiological process which is neutralized by the activity of scavenging substances (superoxide dismutases, peroxidases, vitamin E, vitamin C, methionine), so that no acute damage occurs; when the OR^0 generation massively increases, the balance with the defence systems fails and damage occurs.

The putative sources of OR^0 are: a) The mitochondrial respiratory chains during energy deprivation; b) The metabolic cascade of arachidonic acid overactivated by phospholipases; c) The enzyme xanthine-oxidase derived from xanthine-dehydrogenase in the presence of proteases; d) The granulocytes and other phagocytes activated by complement, bacteria, endotoxin, proteases, lysosomal enzymes, etc. All these events occur during SIRS, sepsis, septic shock and MODS; hence the involvement of OR^0 unbalance and overgeneration in their pathogenetic chains would seem highly probable.

Unfortunately, all the available data on the involvement of OR^0 in the aforementioned conditions are indirect and derived from assessment of byproducts of OR^0 reactions or from the consequences of OR^0 inactivation, without any direct measurements of OR^0 themselves, thus giving rise to some scepticism.

The aim of this paper is to present the main available data supporting the role of OR^0 in the pathogenesis of sepsis and septic shock as fundamental, though not unique, mediators.

Some questions need to be answered and discussed in order to arrive at conclusions and putative therapeutic applications.

ARE THERE ANY SIGNS OF OR⁰ OVERGENERATION DURING SEPSIS AND SEPTIC SHOCK?

The direct signs of OR^0 overgeneration following experimental stimuli capable of producing sepsis and septic shock are few, but the data concerning damage produced by OR^0 are quite numerous.

Direct signs of OR^0 overgeneration

Neutrophils taken from endotoxemic rats produced significantly more O_2^- than controls, both spontaneously and after exogenous stimulation (1). Jackson and coworkers (2), using murine macrophages incubated with endotoxin, recorded an electron spin responance signal of oxygen-centered radicals which disappeared after immission of superoxide dismutase into the system. According to Brackett and coworkers (3), the process of OR^0 generation after endotoxin administration is not immediate but takes some ten minutes, thus suggesting the existence of an intermediate mechanism. Increased H_2O_2 concentration has been measured in the expired breath of patients with

acute respiratory failure (4). Finally, Vespasiano and coworkers (5) measured spontaneous O_2^- production by neutrophils taken from septic patients. Its low value was due to the exhaustion of OR^0 — generating mechanisms as a consequence of their overstimulation; the O_2^- production in fact recovered in direct proportion to the improvement in severity scores of the illness.

Recently, Galley and coworkers (6) measured xanthine oxidase activity in blood. It was absent in healthy subjects, but had significantly increased in septic ones, more in non-survivors than in survivors. Since xanthine-oxidase is an O_2^- producer; its presence might be interpreted as indicative of enhanced superoxide production.

Indirect signs of OR⁰ overgeneration

The list of the biochemical markers of OR^0 overactivity during sepsis, septic shock etc., is long. Malondialdehyde and alkenales, alkanes in exhaled air, chemiluminescende, consumption of physiological scavengers etc. have been cited in laboratory animals and in humans (see 7).

Ethane in exhaled air, an atraumatic marker of lipid peroxidative reactions, increased after endotoxin injection in rats (8). Antioxidant deficiency has been observed in the bronchoalveolar fluid of patients with sepsis and acute lung injury (9—11). High blood levels of malondialdeyde were measured in shocked patients (12), where other investigators have measured conjugate dienes, vitamin E consumption (13, 14), vitamin C consumption (15), lipid peroxides (16). Increased cell membrane stiffness, a typical effect of OR^0 damage (17), has been reported during septic shock in rats or humans (8, 18).

An increase in malondialdehyde, hydroxyalkenales and other products of lipid peroxidation has been reported in erythrocytes and plasma from patients surviving a period of shock (18, 19).

Physiological anti-oxidant capacity was reduced in patients with acute respiratory insufficiency. Blood levels of catalase, superoxide dismutase (20) and ceruloplasmin (21) increased, while glutathione and vitamin E decreased (22). Moreover, the plasma concentration of hydroxynonenal (a very specific peroxidation product) was increased (23).

In conclusion, many indirect signs of OR^0 overgeneration and lipid peroxidation have been reported during sepsis and septic shock in both laboratory animals and humans.

DOES OR⁰ INACTIVATION AFFECT THE EVOLUTION AND SEVERITY OF SEPSIS AND SEPTIC SHOCK?

This is a widely-explored field and experiments have been presented using scavengers, antioxidants, trappers etc. to modify the evolution of sepsis, septic shock and related conditions, which are mostly experimental but also clinical.

In our laboratory this approach has been adapted since 1984 on a laboratory model of lethal endotoxemia or trauma (24, 25).

Sping-trapping nitrones (PBN, POBN, DMPO) prevented and reverted traumatic and endotoxin shock in rats (*Tab. 1*) and prevented the microcirculatory troubles provoked by endotoxin (*Tab. 2*), the OR⁰-induced stiffness of cell membranes and the exhalation of ethane, a by-product of lipid peroxidation (4).

Table. 1. Effect of spin-trapping nitrones (active or inactivated) on survival of rats after endotoxin or trauma. Survival of rats (survived/examined) after injection of *Escherichia coli* endotoxin (30 mg/kg i.p.) or trauma (rotating drum). Animals were pretreated i.p. with one of three spin-trapping nitrones (PBN, POBN and DMPO) or with the same nitrones which had been inactivated by exposure to light and air. Control experiments were performed using saline alone (from Novelli, 1992).

	PBN 150 mg/kg		PC 100 1)BN mg/kg	DM 100 1	Saline	
	Active	Inactive	Active	Inactive	Active	Inactive	
Endotoxin	9/10	0/10	8/10	0/10	8/10	0/10	1/10
Trauma	12/12	0/9	10/10	0/11	12/12	0/11	0/9

Table 2. Effect of spin-trapping nitrones on the microcirculatory derangements provoked by endotoxin. Non-quantitative effects of a lethal dose of *Escherichia coli* endotoxin on some variables of the microcirculation of the mesocecum of rats and on arterial pressure and pH. The modifications caused by each of three nitrones (PBN, DMPO, POBN) tested by i.p. injection or topical application to the mesocecum are summarized (from Novelli, 1992).

Microcirculation									
	Vasomotion				Capillary flow				1
	30	60	120	30	60	120	BP	pH	Outcome
Endotoxin	1	↓	↓↓	Ļ	Ļ	↓↓↓ ·	↓↓	↓↓	Diet
Endotoxin + i.p. nitrones	=	=	=	=	=		=	=	Survived
Endotoxin + topical nitrones	=	=	=	=	=	=	ĻĻ	↓↓	Died
i.p. nitrones	=	=	=	=	=	=	=	=	Survived
topical nitrones	_	=	=	=	=	=	=	0	Survived

These same nitrones have been reported to affect favourably mortality after large doses of endotoxin in rats (26, 27) and to protect the myocardium against reprefusion-induced arrhythmias in rats (28–30).

Other reports from our laboratory have demonstrated a protective action of large doses of reduced glutathione (500 mg/kg) against endotoxin or traumatic shock in rats (31). Reduced glutathione is a potent physiological intracellular antioxidant that was effective in improving systemic and pulmonary hemodynamics during acute respiratory insufficiency in humans (personal unpublished data).

The multiple organ dysfunction syndrome (MODS) can also be viewed as a consequence of continuous, prolonged OR^0 generation. MODS can be provoked in rats by sterile intraperitoneal zymosan in parafin, which causes prolonged complement activation (32); the mortality of this model of MODS was lessened in a dose-dependent manner by large doses of reduced glutathione or by spin-trapping nitrones. Antioxidants also significantly prevented pulmonary interstitial edema (as assessed by computerized tomography) and ethne exhalation (33, 34).

In a similar model of intraperitoneal zymosan injection in rats, the degree of lung and liver peroxidation as assessed with malondialdehyde was correlated to the severity of the illness and with mortality (35). Many years ago, Bitterman and coworkers (36) reported that intravenous superoxide dismutase improved the damaging effects of OR^0 in a model of superior mesenteric artery occlusion shock in rats. The iron-chelator deferoxamine, an inhibitor of OR^0 reactions through iron chelation, exerted an impressive protective effect in early sepsis following cecal ligation and puncture in rats (11). Dimethyl sulfoxide, a hydroxyl scavenger that readily penetrates cell membranes, induced significant beneficial modifications of the course of endotoxemia in rats (37).

N-acetyl-cysteine improved acute lung injury (38) and septic shock (39) in some patients, but available data are few and debatable (see below).

In conclusion, there is evidence that drugs which inhibit OR^0 reactions exert a beneficial action on the evolution of experimental sepsis and septic shock.

MAY THE PATHOPHYSIOLOGY OF SEPSIS AND SEPTIC SHOCK BE VIEWED AS A CONSEQUENCE OF OR⁰ OVERGENERATION?

The basic pathophysiological disorder of sepsis, septic shock and MODS is a diffuse impairment of oxygen consumption and tissue perfusion, mostly focused on microcirculatory and capillary dysfunction. The question is whether such disorders can be viewed as consequences of OR^0 overactivity.

Capillary function and permeability have been studied in our laboratory on the mesocecum of rats with fluorescein-labeled albumin injected i.v. and polarized microscopy. In the absence of any stimulus the capillaries and their walls were outlined by the fluorescence but after a lethal dose of i.v. endotoxin fluorescent albumin progressively leaked out of capillaries and invaded the interstitium. This increase in capillary permeability was prevented by nitrones or glutathione (*Tab. 3*), thus confirming that it was due to overgeneration OR^{0} (40) whose main source is located inside the neutrophils which are activated and which adhere to the post-capillary veins (41).

Table 3. Measure of capillary permeability as assessed by counting the leakage points of fluorescent albumin on the mesocecum of rats. Experiments were performed on a) endotoxin alone, b) endotoxin preceded by reduced glutathione 250 or 500 mg/kg i.p., c) endotoxin preceded by the spin-trapping nitrone PBN (from Novelli, 1993).

Leakage points at various intervals									
	0.00 min.	10 min.	20 min.	30 min.	40 min.				
Endotoxin	0	3 ± 1	7±3	15 ± 6	∞				
Endotoxin + GSM 50 mg/kg	0	0	0	1±2	1 ± 2				
Endotoxin+GSM 250 mg/kg	0	0	1 ± 3	3 ± 4	5±3				
Endotoxin + PBN	0	0	1 ± 2	1 ± 1	2 ± 1				

This finding might be considered analogous to the early increase in pulmonary interstitial water observed by Suffredini and coworkers (42) after intravenous injection of small doses of endotoxin in healthy volunteers. Recently, protein extravasation from the capillaries was demonstrated to be the first step in experimental multiple organ failure provoked by intraperitoneal sterile zymosan (43). An increase in xanthine oxidase activity associated with increased free-radical concentrations and evidence of free radical damage and increased lactate concentrations have all been reported in patients with sepsis and organ dysfunction as compared with both healthy subjects and non-infected critically-ill patients (6).

In conclusion, the overall picture of sepsis ans septic shock can be viewed as a consequence of OR^0 generation not only in laboratory animals but also in humans.

WHAT IS THE RELATIONSHIP BETWEEN OR⁰ OVERGENERATION AND OTHER MEDIATORS OF SEPSIS AND SEPTIC SHOCK?

The above findings demonstrate unbalanced OR^0 overgeneration during sepsis and septic shock that might provoke the typical pathophysiology, and is parallel to the production of the commonly-accepted mediators.

1. Polymorphonuclear granulocytes have a pathogenetic role in the disturbances of sepsis and septic shock as a consequence both of mechanical obstacles to microcirculation and of OR^{0} production ("respiratory burst"). In fact, leukocyte depletion prevented pulmonary and cellular damage after endotoxin injection (44, 14).

Chaotic OR^{0} generation from the granulocytes is the consequence of their activation by complement fractions which are in turn activated by the well-known mediators of sepsis and shock: proteases, lysosomal enzymes, coagulation by-products, bacteria, endotoxins, etc. The source of these mediators is the site of infection or inflammation. The intestine also transfers endotoxins and bacteria into the systemic circulation but at the same time it has been claimed to produce OR^{0} (45).

2. Endotoxins directly stimulate granulocytes and monocytes to produce OR^{0} (2, 46, 47).

3. The tumor necrosis factor (TNF), interleukins (IL-1, IL-6) endothelins and platelet-activating factor (PAF) and nitric oxide (NO) have been claimed as pathogenetic mediators of sepsis and septic shock. None of them however has a unique role in provoking these conditions and discussion is underway concerning their hierarchical relationship. In any case, endothelins stimulate granulocytes to produce OR^0 (50) as does IL-1 (51). PAF also activates OR^0 production by granulocytes through the complement. TNF is a stimulator of OR^0 production by granulocytes and monocytes (52, 53), but it also increases vascular endothelium susceptibility to peroxidative damage (54).

 OR^{o} overgeneration is thus the final step in the actions of all the mediators of sepsis and one of the mechanisms of their destabilizing action on the whole organism.

4. NO, that is a free radical itself and is linked to OR^0 , is an important mediator of sepsis and septic shock (Szabo) (55). The inhibition of its production by i-NOS improved endotoxin and septic shock in both laboratory animals (46, 57) and humans (48, 58). Moreover, the spin-trapping nitrone phenyl-butyl-nitrone (PBN), which we have discovered prevented death due to endotoxin (25), is also a selective inhibitor of i-NOS induction in mice (59, 60).

In any case, NO reacts with superoxide to form peroxynitrite, another free radical whose cytotoxicity is higher than that of NO itself (Brunelli, 1995) (61) and which like all the OR^0 impairs contractile protein activity through disturbances of the calcium transport systems (62). Moreover, the cytokines released during sepsis (TNF, IL-1) are potent inducers of i-NOS and NO production.

It thus seems very difficult to distinguish between the roles of OR⁰ and NO in the pathogenesis and pathophysiology of sepsis and septic shock, also because of the lack of drugs which specifically and selectively act on one or the other system.

FUTURE THERAPEUTIC POSSIBILITIES

The concept of protection by some antioxidant drug against the chaotic, damaging action of OR^0 seems simple and attractive. So far, however, no drug with an anti-oxidant activity has been proved to be effective in controlling acute OR^0 -derived diseases like sepsis and septic shock (see 63—66).

Many drugs that inhibit OR^0 sources or their consequences have been tested in animal models, and they may have favourably affected the evolution of sepsis and septic shock; no one has however been submitted to a controlled clinical trial.

In our patients, large doses of reduced glutathione improved acute lung injury due to sepsis (personal unpublished data), but these observations did not go beyond a preliminary, absolutely non-controlled study.

N-acetylcysteine has a well-established antioxidant profile (67) and was given to patients with acute lung injury by Bernard in 1991 (38) and to septic shock patients by Spies and coworkers (39). The latter was a prospective, randomized, doubleblind study on 58 patients these were given very large doses of N-acetylcysteine as a bolus, followed by infusion. The drug led to increased oxygen consumption in all the treated patients but gastric intramucosal pH, hemodynamics and survival improved in only half of them. The only difference between responders and non-responders to N-acetylcysteine was the interval between the onset of sepsis and the administration of the drug (non-responders: 61 hours; responders: 37 hours), thus suggesting that after a certain time-lag the damage becomes irreversible and no longer able to be influenced by simple OR^0 inactivation.

CONCLUSION

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There is evidence that the pathogenetic chain leading to sepsis and septic shock involves OR^0 as final mediators of biochemical cell and tissue damage. Their pharmacological inactivation has been poorly evaluated in humans, but in any case it might be effective only in the earliest stages of sepsis, before the setting in of structural damage leading to irreversible oxygen consumption deficiency.

REFERENCES

- 1. Simons RK, Maier R, Lennards ES. Neutrophil function in a rat model of endotoxin-induced lung injury. Arch Surg 1987; 122: 197-203.
- 2. Jackson SK, Stark JM, Rowlands CC, Evans JC. Electron spin resonance detection of oxygen-centred radicals in murine macrophages stimulated with bacterial endotoxin. Free Rad Biol Med 1989; 7: 165-170.
- 3. Brackett DJ, Lai EK, Lerner MR, Wilson MF, McCay PB. Spin trapping of free radicals produced "in vivo" in heart and liver during endotoxemis. *Free Rad Res Comm* 1989; 7: 315-324.
- 4. Novelli GP. Oxygen radicals in experimental shock: effects of spin-trapping nitrones in ameliorating shock pathophysiology. Crit Care Med 1992; 20: 499-507.
- 5. Vespasiano MC, Lewandoski JR, Zimmerman JJ. Longitudinal analysis of neutrophil superoxide anion generation in patients with septic shock. Crit Care Med 1993; 81: 666-672.

- 6. Galley HF, Davies MJ, Webster NR. Xanthine oxidase activity and free radical generation in patients with sepsis syndrome. Crit Care Med 1996; 24: 1649-1653.
- 7. Redi H, Gasser H, Hallstrom S, Schlag G. Radical related cell injury. In Pathophysiology of shock, sepsis and organ fuilure, 1993. Springer-Verlag, Berlin Schlag & Redl (ed.) pp. 92-110.
- 8. Sznaider JI, Fraiman A, Hall JB et al. Increased hydrogen peroxide in the expires breath of patients with acute hypoxemic respiratory failure. Chest 1989; 96: 606-612.
- 9. Pacht ER, Timerman AP, Lykens MG, Merola AJ. Deficiency of alveolar fluid glutathione in patients with sepsis and the adult respiratory distress syndrome. *Chest* 1991; 100: 1397-1403.
- 10. Lykens MG, Davis WB, Pacht ER. Antioxidant activity of bronchoalveolar lavage fluid in the adult respiratory distress syndrome. Lung Cell Mol Physiol 1992; 6: L169-175.
- 11. Moch M, Schroppel B, Schoenberg MH et al. Protective effects of hydroxyethyl starch-deferoxamine in early sepsis. Shock 1995; 4: 425-432.
- 12. Ospici P, Cannistraci M, Foschi A et al. Studio sul comportamento della malondialdeide plasmatica e tissutale in soggetti in stato di shock. Acta Anaesth Ital 1983; 34: 867-872.
- 13. Gangeri G, Sganga G, De Gaetano A et al. Oxygen free radicals damage in sepsis and multiple organ failure: plasma levels of conjugates dienes and endogenous antioxidants. Clin Intensive Care 1992; 3: 16.
- 14. Kapoor R, Prasad K. Role od polymorphonuclear leukocytes in cardiovascular depression and cellular injury in hemorrhagic shock and reinfusion. *Free Rad Biol Med* 1996; 21: 609-618.
- 15. Borrelli E, Roux-Lombard P. Grau GE et al. Plasma concentrations of cytokines, their soluble receptors and antioxidant vitamins can predict the development of multiple organ failure in patients at risk. Crit Care Med 1996; 24: 392-397.
- 16. Keen RR, Stella L, Flanigan DP, Lands WE. Differential detection of plasma hydroperoxides in sepsis. Crit Care Med 1991; 19: 1114-1119.
- 17. Patel JM, Block ER. The role of oxidant gases on membrane fluidity and function in pulmonary endothelil cells. Free Rad Biol Med 1988; 4: 121-134.
- 18. Poli G, Biasi F, Chiarpotto E et al. Lipid peroxidation in human disease: evidence of red cell oxidative stress after circulatory shock. Free Rad Biol Med 1989; 6: 167-170.
- 19. Biasi F, Chiarpotto E, Lanfranco G et al. Oxidative stress in the development of human ischemic heptitis during circulatory shock. Free Rad Biol Med 1994; 17: 225-233.
- 20. Leff JA, Parson PE, Day CE. Serum antioxidants as predictors of adult respiratory distress syndrome. *Lancet* 1993; 341: 777-780.
- 21. Krser-Staples JA, Kew RR, Webster RO. Ceruloplasmin and transferrin levels are altered in serum and bronchoalveolar lavage fluid of patients with the adult respiratory distress syndrome. Am Rev Respir Dis 1992; 145: 1009-1015.
- 22. Richard C, Lemonnier F, Thibauld M, Auzepy P. Vitamin E deficiency and lipoperoxidation during adult respiratory distress syndrome. Crit Care Med 1990; 18: 4-9.
- Quinlan GJ, Lamb NJ, Ewans TW, Gutteridge JMC. Plasma fatty acid changes and increased lipid peroxidation in patients with adult respiratory distress syndrome. *Crit Care Med* 1996; 24: 241-146.
- 24. Novelli GP, Angiolini P, Tani R, Consales G, Bordi L. Phenyl-t-butyl-nitrone is active against traumatic shock in rats. Free Rad Res Comm 1985; 1: 321-327.
- 25. Novelli GP, Ursini F (eds). Oxygen free radicals in shock. Karger, Basel, 1986.
- McKechnie K, Furman BL, Parrat JR. Modification by oxygen free radical scavengers of the metabolic and cardiovascular effects of endotoxin infusion in conscious rats. *Circulatory Shock* 1986; 19: 429-439.
- 27. Hamburger SA, McCay PB. Endotoxin-induced mortality in rats is reduced by nitrones. *Circulatory Shock* 1989; 29: 329-334.
- 28. Hearse DJ, Tosaki A. Free radicals and reperfusion induced arrhythmics: protection by spin traż agent PBN in the rat heart. Circulation Res 1987; 60: 375-383.

- 29. Bolli R, Patel BS, Jeroudi MD, Lai EK, McCay PB. Demonstration of free radical generation in "stunned" myocardium of intact dogs with the use of the spin-trap α-phenyl-N-tert-butylnitrone. J Clin Invest 1988; 82: 476-485.
- Bradamante S, Monti E, Paracchini L, Piccinini F. Protective activity of the spin trap tert-butyl-alpha-phenyl-nitrone (PBN) in reperfused rat heart. J Mol Cell Cardiol 1992; 24: 375-386.
- 31. Falsini S, Cellai MP, Angiolini P, Cavuta M, Novelli GP. Glutatione ridotto e L-cisteina nello shock endotossinico nel ratto. *Min Anest* 1994; 60: 413-418.
- 32. Goris RJA, Boekholtz WKF, Van Bebber IPT, Noytinck JKS, Schillings PHM. Multiple organ failure and sepsis without bacteria. An experimental model. Arch Surg 1986; 121: 897-901.
- Di Filippo A, Paternostro E, Scardi S, Consalvo M. Insufficienza multipla d'organo sperimentale nel ratto: effetti protettivi del glutatione ridotto. Acta Anaesth Ital 1992; 43: 358-368.
- 34. Di Filippo A, Scardi S, Consalvo M, Ridolfi N, Pellegrini G, Paternostro E, Novelli GP. L'etano espirato come marker non invasivo della evoluzione della Multiple Organ Dysfunction Syndrome (MODS) sperimentale. *Min Anest* 1994; 60: 295-303.
- 35. Demling R, Nayak U, Ikegami K, Lalonde C. Comparison between lung and liver lipid peroxidation and mortality after zymosan peritonitis in the rat. Shock 1994; 2: 222-227.
- 36. Bitterman H, Aoki N, Lefer AM. Anti-shock effects of human superoxide dismutase in splanchnic artery occlusion shock. Proc Soc Exptl Biol Med 1988; 188: 265-271.
- 37. Brackett DJ, Lerner MR, Wilson MF. Dimethyl sulfoxides antagonizes hypotensive metabolic and pathologic responses induced by endotoxin. *Circulatory Shock* 1991; 33: 156-163.
- Bernards GR. N-acetylcysteine in experimental and clinical acute lung injury. Am J Med 1991;
 91 (Suppl 3): 54-59.
- 39. Spies CD, Reinhart K, Witt I et al. Influence of n-acetylcysteine on indirect indicators of tissue oxygenation in septic shock patients: results from a prospective, randomized, double-blind study. Crit Care Med 1994; 22: 1738-1746.
- 40. Novelli GP, Casali R, Bonizzoli M et al. Aumento della permeabilità capillare provocato dall'endotossina: protezione con antiossidanti e glutatione. Min Anest 1993; 59: 211-216.
- 41. Suzuki M, Asako H, Kubes P et al. Neutrophil-derived oxidants promote leukocyte adherence in postcapillary venules. *Microvasc Res* 1991; 42: 125-138.
- 42. Suffredini AF, Shelhamer JH, Neumann RD, Brenner M, Baltaro RJ, Parrillo JE. Pulmonary and oxygen transport effects of intravenously administered endotoxin in normal humans. Am Rev Resp Dis 1992; 145: 1398—1403.
- 43. Nieuwenhuijzen GAP, Knapen MFCM, Oyen WJG, Hendriks T, Corstens FHM, Goris RJA. Organ damage is preceded by changes in protein extravasation in an experimental model od multiple organ dysfunction syndrome. *Shock* 1997; 7: 98-104.
- 44. Heflin HC, Brigham KL. Prevention by granulocyte depletion of increased vascular permeability of sheep lung following endotoxemia. J Clin Invest 1981; 68: 1253.
- 45. Turnage RH, Magee JC, Guice KS, Myers SI, Oldham KT. Complement activation by the hydroxyl radical during intestinal reperfusion. *Shock* 1994; 2: 445-450.
- 46. Domenici-Lombardo L, Adembri C, Consalvo M et al. Evolution of endotoxin induced acute lung injury in the rat. Int J Exp Path 1995; 76: 381-390.
- 47. Jajdacsy-Balla A, Doi EM, Lerner MR et al. Dose-response effect of in vivo administration of endotoxin on polymorphonuclear leukocytes oxidative burst. Shock 1996; 5: 357-361.
- 48. Petros A, Bennett D, Vallance P. Effect of nitric oxide synthase inhibitors on hypotension in patients with septic shock. Lancet 1991; 338: 1557-1558.
- 49. Schilling JM, Cakmakci M, Battig U, Geroulanos S. A new approach in the treatment of hypotension in human septic shock by N-monomethyl-L-arginine an inhibitor of the nitric oxide synthethase. *Intensive Care Med* 1993; 19: 227-231.

- 50. Huribal M, Kumar R, Cunningham ME, Sumpio BE, McMillen MA. Endothelin-stimulated monocyte supernatants enhance neutrophil superoxide production. *Shock* 1994; 1: 184–187.
- 51. Meier B, Radeke HH, Selle S et al. Human fibroblasts release reactive oxygen species in response to interleukin-1 or tumor necrosis factor-α. Biochem J 1989; 263: 539-545.
- 52. Yamauchi N, Watanabe N, Kuriyama H et al. Suppressive effects of intracellular glutathione on hydroxyl radical production induced by tumor necrosis factor. Int J Cancer 1990; 46: 884-888.
- 53. Lloyd SS, Chang AK, Taylor FB, Janzen EG, McCay PB. Free radicals and septic shock in primates: the role of tumor necrosis factor. *Free Rad Biol Med* 1993; 14: 223-242.
- 54. Varani J, Bendelow MJ, Sealey DE et al. Tumor necrosis factor enhances the susceptibility of vascular endothelium to neutrophil mediated killing. Lab Invest 1988; 59: 292-295.
- 55. Szabo C. Alterations in nitric oxide production in various forms of circolatory shock. New Horizon 1995; 3: 2-32.
- 56. Tiemermann C, Vane J. Inhibition of nitric oxide synthesis reduces the hypothension induced by bacterial lipopolysaccharides in the rat in vivo. *Europ J Pharmacol* 1990; 182: 591-595.
- 57. Kilbourn RG, Jubran A, Gross SS. Reversal of endotoxin-mediated shock by N-methyl-L-arginine, an inhibitor of nitric oxide synthesis. *Biochem Biophys Res Commun* 1990; 172: 1132-1138.
- 58. Lorente JA, Landin L, De Pablo R, Renes E, Liste D. L-arginine pathway in the sepsis syndrome. Crit Care Med 1993; 21: 1287-1295.
- 59. Miyajima T, Kotake Y. Skin-trap N-tert-butyl nitrone (PBN) inhibits induction of nitric oxide synthase in endotoxin-induces shock in mice. *Biochem Biophys Res Commun* 1995; 215: 114-121.
- 60. Miyajima T, Kotake Y. Optimal time and dosage of phenyl-N-tert-butyl nitrone (PBN) for the inhibition of nitric oxide synthase induction in mice. *Free Rad Biol Med* 1996; 22: 463-470.
- 61. Brunelli L, Crow JP, Beckman JS. The comparative toxicity of nitric oxide and peroxynitrite to Escherichia Coli. Arch Biochem Biophys 1995; 316: 327-334.
- 62. Ishida H, Ichimori K, Hirota Y, Fukahori M, Nakazawa H. Peroxynitrite-induced cardiac myocyte injury. Free Rad Biol Med 1996; 20: 343-350.
- 63. Rice-Evans CA, Diplock AT. Current status of antioxidant therapy. Free Rad Biol Med 1993; 15: 77-96.
- 64. Schiller HJ, Reilly PM, Bulkley GB. Antioxidant therapy. Crit Care Med 1993; 21: S92-102.
- 65. Maxwell SRJ. Prosoects for use of antioxidant therapies. Drugs 1995; 49: 345-361.
- 66. Aruoma Ol. Characterization of drugs as antioxidant prophylactics. *Free Rad Biol Med* 1996; 20: 675-705.
- 67. Aruoma Ol, Halliwell B, Hoey BM, Butler J. The antioxidant action of N-acetylcysteine: its reaction with hydrogen peroxide, hydroxyl radical, superoxide and hypochlorous acid. Free Rad Biol Med 1989; 6: 593-597.

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Author's address: G. P. Novelli, Institute of Anaesthesiology and Institute Care, Careggi Hospital, Viale Morgagni 85, 1-50134 Florence, Italy.