Review article

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LABILE PRODUCTS OF VASCULAR ENDOTHELIUM AS MEDIATORS AND MODULATORS OF THE FUNCTIONS OF THE CENTRAL NERVOUS SYSTEM

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A rapid development of the knowledge about vascular endothelial cell function as an "endocrine gland" releasing the labile, highly biologically active products, caused a major reapprisal of our concepts concerning the pathophysiology of our body. The publication summarizes the present understanding of the involvement of nitric oxide (NO), endothelins (ETs) and arachidonic acid products in the mechanisms underlying the regulation of the tonus of vessels supplying blood to the CNS, their known modulatory and mediatory role in CNS functions such as a development and memory, peripheral nonadrenergic noncholinergic, or sensory neurotransmission. The regulation intracellular Ca⁺⁺ion levels as a proposed mechanism for the neuroprotective, as well as the neurotoxic effect of the described endothelial products is presented. The supposed therapeutical usefullness of compounds which can modulate their biosynthesis, substitute their activity, or modify its degradation are also summarized.

Key words: endothelins, prostanoids, nitric oxide, central and peripheral nervous system

INTRODUCTION

The rapidly developing knowledge of the function of the vascular endothelial cells in maintaining the integrity of the vascular wall, patency of vessels, and fluidity of blood, completely changed the concept of their role in physiology and pathology of our body. Endothelial cells function is related to their capacity to synthetize a vast number of substances such as: prostacyclin (PGI_2) and the other arachidonic acid metabolites (1-3), "endothelium derived relaxing factor" (EDRF) (4), proved to be nitric oxide (NO) (5),

endothelins (6), activators (t-PA) (7), and inhibitors (PAI) (8) of the plasma fibrinolytic activity, cytokines (9), adhesion and chemotaxis promoting selectins and integrins (10-12), growth promoting factors (13), a platelet activating factor (PAF) (14), a wide range of antioxidant enzymes (15) and others. Most of these substances released continuously, support the blood/vessel wall homeostasis and the sufficient blood supply to the tissues.

Thus, hyperoxia, hypoxia or anoxia of tissues (4, 15), tissue damage related to inflammation, such as oedema and cytotoxicity due to the presence of activated phagocytic cells (6, 17) are strongly influenced by the biological efficiency of the endothelium.

The endothelial regulatory system is still not fully understood. Three components of this system will be reviewed here: EDRF/NO, endothelins, and some metabolites of arachidonic acid in the aspect of blood supply and the function of the central nervous system.

EDRF/NO

EDRF identified as nitric oxide (NO) (5), or rather as its free radical (NO') (17) is biosynthesized from L-arginine (L-Arg) by oxidative desimination (18, 19) or perhaps from peptides which contain L-Arg (20). The co-product of this reaction, citrulline, is further metabolised back to arginine, forming "a half-urea cycle" (21). (Fig. 1). This enzymic process takes place not only in vascular

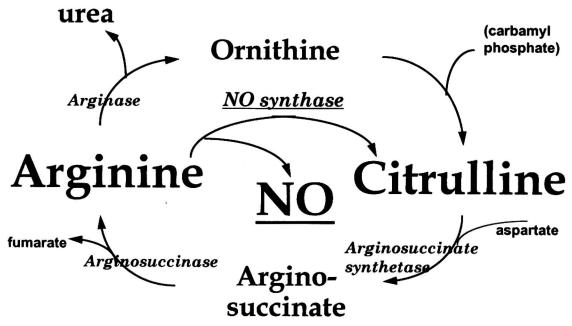


Fig. 1. Metabolism of arginine in the body; "the urea cycle". NO-synthase (NOS) is forming NO in the "part-urea cycle".

endothelium (4, 19), but also in macrophages (22), neutrophils, blood monocytes (23), mastocytes, hepatocytes, Kupfer cells (24, 25), non-adrenergic non-cholinergic (NANC) nerve endings (26), and neurons (27, 28).

Fig. 2. Structural formulae of L-arginine (L-Arg), and some of NOS inhibitors: N^ω-monomethyl-L-arginine (L-NMMA); N-iminoethyl-L-ornithine (L-NIO); N^ω-nitro-L-arginine (L-NA); N^ω-nitro-L-arginine methyl ester: (L-NAME); N^ω-amino-L-arginine (L-NAA); asymmetrical N^ω-dimethyl-L-arginine (ADMA).

Some structural analogs of L-Arg (Fig. 2) (22, 23) have been used as the NO-synthase (NOS) antagonist to study the biochemistry and physiology of the NOS-NO pathway.

Recently it has become apparent, that there are at least two types of NOS. One is constitutive, membrane bound (in endothelium) (29), or cytosolic (in brain), Ca⁺⁺/calmodulin dependent, and releases NO in smaller amounts for short periods of time in response to receptor or physical stimulation (25, 30, 31). NO released by this enzyme acts as the transductive responses mechanism,

mediating several physiological responses through stimulation of soluble cytosolic guanylate cyclase, and elevating the cellular c-GMP levels (32, 33). Acetylcholine, bradykinin, substance P, thrombin, adenosine diphosphate, and 5-hydroxytryptamine, are the best known activators of the NO release, by constitutive NOS from endothelial cells, elevating intracellular Ca⁺⁺via receptor mediated increase of inositol 1,4,5-triphosphate and diacylglycerol levels (25, 30, 31). NO released in this way, plays an important role in maintaining the basal tonus of the vessel, counteracting the activity of all endogenous and exogenous vasoconstrictors (25, 30, 34, 35). It has been recently postulated that the endogenously formed L-Arg metabolite ADMA (Fig. 2) is responsible for the hypertension observed in patients with severe kidney insufficiency (36) or eclampsia (37).

Intercellular liberation of NO constitutes the common principle of pharmacological action for glyceryl trinitrate and other organic nitrates recognized recently as the "NO-donors" (38). In contrast to endogenous, the generation of NO from exogenous NO-donors is a reductive process that requires cysteine, glutathione and special enzymes, and takes place in some cells, for example in the vascular smooth muscle, but not in platelets (39). Another NO-donor, SIN-1A, the metabolite of molsidomine, releases NO in an aqueous solution in a non-enzymatic-, pH-, temperature- and time-dependent manner, causing the relaxation of the vasculature without any signs of tachyphylaxis (40).

Another biologically potent activity of the NO formed by the constitutive NOS in endothelium is the inhibition of platelet aggregability, release reaction, and its adhesion to endothelium and polymorphonuclear leukocytes (PMNs) (41—44). In the antiplatelet activity EDRF/NO synergises with PGI₂ on the basis of accumulation of the intracellular c-GMP and cAMP levels (43, 45). No synergistic activity in vasodilatory properties of both autacoids has been demonstrated (46).

NO molecule is short living and decomposes in the aqueous solution to nitrite and nitrate ions, deprived of the biological activity of nitric oxide. NO is instantly destroyed by superoxide anions (O_2^*) (47). Oxyhaemoglobin is a powerful scavenger of NO, methylene blue while inhibiting guanylate cyclase hinders biological effect of EDRF/NO (48). Superoxide dismutase (SOD) and inhibitors of c-GMP phosphodiesterase potentiate the biological effects of NO (47, 48).

Another izoform of NOS is induced after activation of macrophages, monocytes, neutrophils, fibroblast, endothelial and a number of other cells by cytokines (TNF- α , IL-1, IF- γ or LPS), and once expressed, sythesizes the larger (comparing to inducible NOS) amounts of NO for long periods of time (25, 30, 31, 49, 50, 51). Furthermore, this enzyme which is cytosolic, Ca⁺⁺/calmodulin independent, requires tetrahydrobiopterin as a cofactor and its induction is

inhibited by corticosteroids and some cytokines (IL-8, IL-10) (52, 53). So far, the only clearly established role of this NO is as a cytotoxic molecule for invading microorganisms and tumor cells, thus mediating the so called nonspecific immunity of our body (25, 54-57). The release of NO *via* this enzyme is also included in pathological vasodilatation, observed in septic shock (58, 59) and an endothelium and tissue damage accompanying the inflammatory/immune reactions (16, 17). The existence of a human inducible NOS (iNOS) is strongly suggested by the elevation of nitrate in the plasma and urine of individuals who have received cytokines (61, 62), or become septic (63).

The cytotoxicity of NO released by iNOS is linked to several processes. NO binds to all Fe-S enzymes, which results in inhibition of many cell oxydoreductases and *cis*-aconitase (64).

NO disruption of ferritin may acount for the Fe release from target cells incubated with activated macrophages (64). The free Fe may promote lipid peroxidation (65).

A prominent action of macrophage derived NO on tumor cells is the inhibition of their synthesis of DNA by inhibition of ribonucleotide reductase (64). Superoxide anion may be another important target of the toxic activity of NO. Reaction of O_2° with NO results in formation of peroxynitrite, which decays to nitrogen dioxide and hydroxyl radical, considered as the strongest oxidant in biological systems (17).

The interaction of NO with sulfhydryls has brought the hypothesis that S-nitrosothiols could be long-living reservoirs of bioactive NO (66). On the other hand, S-nitrosylation inactivates SH-dependent bacterial dehydrogenases and nitrosylation of proteins leads to the formation of N-nitrosoamines with cancerogenic properties (67).

cNOS and iNOS differ also in the susceptibility to inhibition by various L-Arg analogs, most evident in the relative selectivity of L-NA for cNOS (68). Also compounds that bind calmodulin, such as calcineurin or trifluoroperazine, are the only selective inhibitors on cNOS activity known so far (69). The above mentioned inhibition of the iNOS induction by corticosteroids (52), may add one more mechanism to the understanding of an anti-inflammatory activity of these compounds.

In the central nervous system the biological activity of NO is not restricted to the vessels, but the discoveries of the last five years have demonstrated the function of NO as a messenger and a synaptic plasticity modulator in CNS and in peripheral NANC and sensory neurons (21, 25, 28, 31). The participation of NO in neurotoxicity and opioid dependence has also been suggested (70, 71).

Neurotransmission by agents such as ACh, glutamate, and glycine has long been known to be associated with calcium requiring elevation of c-GMP levels in the brain, and particularly in cerebellum (72). In 1982 L-Arg was identified as the endogenous activator of the soluble guanylate cyclase in neuroblastoma

cells (73). These observations together with the discovery of the L-Arg/NO pathway in vascular endothelium led to the concept of the existence of such a pathway in the central nervous system. Knowles et al., demonstrated that addition of L-Arg to rat synaptosomal cytosol in the presence of NADPH, resulted in the formation of NO and cytrulline and was accompanied by the stimulation of soluble guanylate cyclase (74). This enzyme, calcium/calmodullin dependent, is inactive in a resting (80 nM) concentrations of Ca++ synaptosomes (25) whereas it was fully active at Ca++ concentrations of 400 nM (74). Interestingly enough, physiological Ca⁺⁺ levels, which are essential for the action of the brain NOS, were found to inhibit the brain soluble guanylate cyclase (74). This could represent a control mechanism whereby guanylate cyclase is not activated in those CNS cells stimulated to produce NO, but only in the effector cells (74). The brain, but not endothelial cNOS contains flavins (FAD and FMN), and thus may act as a cytochrome P-450 reductase (75). The brain NOS is competitively inhibited by L-MNNA, L-NA and L-NIO, but not by L-canavinine (76). Histoimmunochemical studies using antibodies to the cNOS (28) followed by the measurement of the NOS activity in the cytosolic fraction of different rat brain regions (77) have showed that the highest concentration of NOS was present in the cerebellum, followed by the hypothalamus and mildbrain, striatum, and hipocampus, with the lowest activity found in the medulla oblongata. The granule cells have been suggested to be the principal neurons in the cerebellum, which release NO in response to exogenous excitatory aminoacid (NMDA) receptors (28, 78). An increased activity in the excitatory pathways has long been known to cause increased levels of cGMP particularly in cerebellar cortex (21, 78).

The neurotransmitter released in all of the main excitatory synapses in the cerebellum acts through excitatory aminoacid receptors, and is probably glutamate (21). There are two categories of recognised excitatory aminoacid receptors: the ones connected with ion channel (ionotropic receptors), and those coupled to G proteins (metabotropic receptors). The ionotropic receptors are subdivided into three types, according to their selective agonists: NMDA, AMPA (quisqualate), or kainate. These types of ionotropic receptors are habituary termed NMDA and others are referred to as: non-NMDA receptors (21). In developing rat cerebellum NMDA receptors mediate most, if not all of the cGMP response to glutamate (79). In adult mice "basal" cerebellar, as well as after pharmacological intervention increased cGMP levels are reduced by the selective NMDA agonists (80). Up to now it has not been proved that either exogenous glutamate or the endogenous neurotransmitter elicits cGMP accumulation through non-NMDA receptors. In fact, glutamate has been shown to be a potent inhibitor of the elevations in cGMP that are induced by the exogenous non-NMDA agonist — kainate (81). This observation, with the evidence that the accumulation of cGMP levels is observed not in neurones

that are stimulated by the glutamate receptor agonist, but in surrounding cells only, raised the concept of existence of the mobile, permeable molecule mediating the intracellular communication (79, 81, 82). Nitric oxide with its lipophylic properties, and the short-lasting activity was a good candidate for such an action. The activation of NMDA receptors raises the cytosolic Ca⁺⁺ levels due to the receptor-operated ion channels. The same Ca⁺⁺ influx is believed to mediate the brain cNOS activity and to initiate many physiological and pathological effects of NMDA receptor activation (21).

Brain NOS is inhibited by the L-Arg metabolites. This effect can be reversed in vitro by the supplementation of the exogenous L-Arg (21). Interestingly enough, in the immature cerebellum, the inhibition curve for L-NA (but not L-NMMA) shows two components (83). One component is evident for very low ($IC_{50} = 6nM$) concentration, but with maturation, this component is lost, leaving the other component ($IC_{50} = 600nM$) observable in the adult. This suggests the existence of two NO-synthases, which are differently sensitive to L-NA, (but not to L-NMMA), and that one of them exists only during the development period (21,83).

Cerebellum is that part of the brain where the NO synthase is concentrated. Cerebellum contains two main neurone types: the large Purkinje cells, and small but numerous granule cells. The granule cells are a major site of NMDA receptor-mediated NO formation, and of cGMP accumulation in the developing tissue (84, 77, 78). NO generated presynaptically in granule cells appears to have at least two potential targets located in postsynaptic membranes. One is in astrocytes, and the other one in the Bergman glial cell bodies (21). The regional distribution of NOS does not entirely match that of NMDA receptors, and for example, neurones in the deep cerebellar nuclei express many NMDA receptors, but NMDA is unable to induce measurable increases in cGMP levels there (82). Cells in these nuclei do however, respond to the exogenous NO-donors (85), suggesting a different stimulus for NO formation, or the inhibition of guanylate cyclase by accumulating intracellular Ca⁺⁺, as mentioned above (74).

Both the localisation of NOS as well as the effector cells reactivity and its relation to physio- and pathological meaning need further investigations. As it is presently understood, NO is the one of the supposed candidates for mediating (via NMDA receptor) the processes of memory and learning (21). It could be involved in triggering the long-lasting changes in synaptic strength on which certain forms of learning are believed to depend (21). The other mechanism the NMDA-activity dependent reorganisation of the afferent fibers, with respect to the target neurones during brain maturation and development, is also suggested to be mediated by NO (86). The astrocytes activation has been demonstrated to influence the plasticity of neuronal synaptic connections (87) which may be related to the changes in the membrane ion channels, which are

dependent on the cGMP level. This effect is also supposed to be mediated by NO in CNS (21). Moreover, the endogenously formed inhibitors of NOS such as LNMMA, ADMA and D'MA have been recently isolated from the bovine brain (88).

The L-Arg/NO pathway may also play a role in the pathology of the central nervous system. It has been demonstrated that an excessive NMDA receptor activation, with the consequent increase in intracellular Ca⁺⁺ accumulation, contributes to glutamate neurotoxicity by an enhanced production of NO (89, 90). Thus, it appears, that the biological responses to NO could be biphasic, as it is observed in glutamate and related excitatory amino acids: that is, physiological or pathological effects may occur. An of NMDA receptor can influence neuronal development, differentiation and plasticity (21, 91-93) via NO-stimulated cGMP accumulation, as it has been suggested for rodent cerebellar tissue (21, 78, 79, 94). The observations that NMDA and kainate enhance the viability of cerebellar granule neurons grown under low potassium conditions (90) confirm the trophic effect of excitatory amino acids on cerebellar granular cells (95, 96). However, Boje and Skolnick demonstrated that the exogenous NO-donor, SNAP (S-nitroso-N-acetylpenicillamine) was toxic for the cultured cerebellar granule neurons (90). This toxicity was enhanced by SOD, which protects NO, and abolished in the presence of oxyhemoglobin, which scavenges NO, pointing to toxic effect on nitric oxide itself. The NO-mediated neurotoxicity after glutamate was soon after that (97). It was also suggested that the stimulation of NMDA receptors induced by ischaemic insult, could raise cytosolic calcium followed by the overproduction of NO (21). However, NMDA antagonists exert few, if any, protective effects of stratial infarctus induced by occlusion of middle cerebral artery (98), the inhibition of NO biosynthesis by L-NAME, or L-NA which protected the rat brain against the focal cerebral ischaemia (99, 100). These observations may suggest that the NO production by ischaemic insult may be at least in part unrelated to the NMDA receptor overstimulation. They also suggest that the inhibition of the CNS NOS is beneficial for the treatment of the cerebral ischaemia.

Within the discrete pattern of NO-synthesizing neurons in brain, NOS was found to colocalize with the cholinergic brain stem-thalamic system, which is thought to regulate the state-dependent activity of the thalamocortical circuit (28). It was demonstrated that the release of NO onto thalamocortical neurons results in an alteration in voltage dependence of the hyperpolarization-activated cation conductance, probably mediated via cGMP (101). The administration of L-Arg into the lateral cerebral ventricle in rats resulted in behavioural stimulation, electrocortical desynchronization with occasional isolated high voltage spikes, but not motor seizures (102). The simultaneous administration of low doses of NMDA resulted in behavioural and

electrocortical seizures, and these effects were prevented by pretreatment of the rats with L-NA (102). Thus, the L-Arg/NO pathway may also participate in the pathomechanism of epileptic seizures associated with brain insult. Zhang and co-workers (103) demonstrated that hyperoxia induced convulsions in rats which are associated with the decrease in cerebral norepinephrine and GABA content, and accumulation of L-Arg. Pretreatment of rats with pargyline (an MAO inhibitor), or L-NA, completely protected from seizures, inhibited the accumulation of L-Arg, and depletion of epinephrine, but not of GABA. Moreover, Oury and co-workers even suggested, that the superoxide anion formed during O2 toxicity may have some protective activity via inactivation of NO overproduced in brain during hyperoxia (104). Comparing to normal mice, they demonstrated that, the increased mortality during the 25 minut exposure of the transgenic animal, overexpressing human extracellular SOD, to hyperbaric oxygen can be prevented by the pretreatment with L-NA (104). Thus, the oxygen toxicity may be also related to the overproduction of NO in CNS.

The central and peripheral neurotoxicity may be also mediated by biologically active substances released not only by cells constitutively present in the neural tissue, but also by the migrating, phagocytizing cells. Hartung et al. (105) have recently reviewed the role of nitric oxide, oxygen radicals, arachidonic acid metabolites, proteases in inflammatory demyelination of neurons, mediated by cytokines (IL-1, II N-γ TNF-α), as well as complement activated astrocytes, microglial cells and macrophages. Microglial cells transformation to phagocytosing brain macrophages takes place in neuronal and/or terminal degeneration after the nerve lesion (106), and the induction of NOS by phagocytosis, and the above mentioned cytokines, has recently been demonstrated in cloned murine microglial cells (107).

In the peripheral nervous system the postulated role of NO as a neurotransmitter released by "non-adrenergic, non-cholinergic" (NANC) nerves (108–110) has been confirmed by the reaction of anti-cNOS antibody with neurons in the myenteric plexus in the intestine and medulla of the adrenal glands (111). The activation of these neurons leads to the inhibition of the tonus of the smooth muscle in the gastrointestinal tract (esophagus, stomach, duodenum, ileocolonic junction) where it mediates adaptive relaxation (108–110), penile corpus cavernosum, (108–111), airways (109, 112), pulmonary arteries (113) mediated by the release of NO, and the accumulation of cGMP in effector smooth muscle cells. However extensively studied, there is no clear-cut mechanism explaining how the initiation of NANC activation is induced. Electrical stimulation of the vagal NANC nerves leads to the profund relaxation of the esophagus, stomach fundus and corpus promoting the propulsion of food (108, 109). The rise in intragastric pressure leads to a sudden relaxation of the stomach fundus, which prevents the further

increase in the lumen pressure, arguing for the local inhibitory reflex activated by the extension of the stomach (110). Ganglionic nicotinic transmission leading to the NO-mediated relaxation of the stomach and guinea-pig trachea for the local sensory reflex was postulated (110, 112).

Interestingly enough, all the known activators of NO release from endothelium such as Substance P, VIP, CGRP, 5-HT and neuropeptide-Y containing neurons have been immunolocalized in the myenteric plexus (114), that suggests the participation of these mediators in the local reflexes with the final activation of NO release from neuronal or from the effector tissue. The resistance of pressure induced gastric relaxation (110), or the nicotinic receptor activated relaxation of guinea-pig trachea (112) to hexametonium can also be explained by an axon reflex (115), mediated by NO released from sensory nerves. In rabbits, a nociceptive effect of acetylcholine has been shown to be mediated through nicotinic receptors present on perivascular sensory nerve endings (116).

In men, iontophoresed ACh can stimulate peripheral nociceptive C-fibres to produce flare (neurogenic vasodilatation) which is abolished by an anaesthetic, and is absent in denervated skin (117, 118). Neurogenic inflammation involves vasodilatation, plasma protein extravasation and oedema. It can be also elicited by antidromic stimulation of the unmyelinated C-fibres of sensory nerves by topical application of chemical irritants, such as capsaicin, mustard oil, xylene (119, 120). Pretreatment of rats with high doses of capsaicin, which selectively destroys C-fibres (121), completely prevents the ability to elicit neurogenic inflammation by electrical stimulation (122). The capsaicin-sensitive somatic neurones include the polymodal nociceptors, which are sensitive to a variety of noxious chemical, thermal and mechanical stimuli, mediating autonomic reflexes (123, 124). Lembeck was the first who suggested that Substance P is the primary afferent neurotransmitter of inflammation (120). Subsequent studies demonstrated the accuracy of this proposal. Substance P along with neurokinin A, and CGRP (125-127) was found in C-fibres sensory nerves, indicating that it may play a role in neurogenic inflammation. Both symptoms: vasodilatation and oedema, of neurogenic inflammation, are attenuated by pretreatment with L-NA and L-NMMA, but not by their D-stereoisomers (127, 128). These results may add to the understanding of the hypothesis concerning the participation of NO in the control of basal tonus of vasculature (25, 31, 35, 36). It well may be that NO controlling the tonus is released by activation of the local sensory reflex arches. The afferent sensory neurones containing the above tachykinins as well as bradykinin, 5-HT, histamine, all known mediators of NO release from endothelium, were found to control vascular tonus and permeability in numerous tissues other than the skin, such as coronary vasculature (129), hepatic (130), airway and nasal mucosa (131, 132), meninges (133), eyes (134), joints (135-137), urinary bladder (135, 136), gastrointestinal tract (137, 138). It well can be that functionary

hyperaemia (139), as well as reactive hyperaemia (140) observed in working tissues or in tissues during reperfusion following short, seconds-lasting ischaemia, could be mediated by NO released from sensory nerves or from endothelium activated by tachykines, released from these nerves on the way of the local reflex. The local biochemical (pH, pO₂) and physical (pressure, stretch) properties, are good candidates for the initiation of such a reflex.

It is interesting that the neurogenic inflammation can be inhibited at the presynaptic level with opioid receptor agonists (141). Ferreira and co-workers (142–144) demonstrated that the local analgesic effect of substances such as ACh, or morphine is mediated *via* stimulation of the L-Arg/NO/c-GMP pathway. They also showed, that NO-donors, such as NaNP, or Sin-1 antagonized carrageenin and PGE₂ hyperalgesia in rats, and applied locally, caused analgesia and the reduction of the arm volume in patients with thrombophlebitis (144). The potentiation of the antinociceptive effect of β-endorphin in mice by L-Arg was also reported (145).

On the other hand, the pretreatment of animals with the NOS inhibitor: L-NMMA, reduced the carageenin-induced oedema in rat paw (146), and the L-NA administered intrathecally enhanced morphine nociception in the rat spinal cord (147) (Fig. 3). Thus, the results concerning the role of NO in

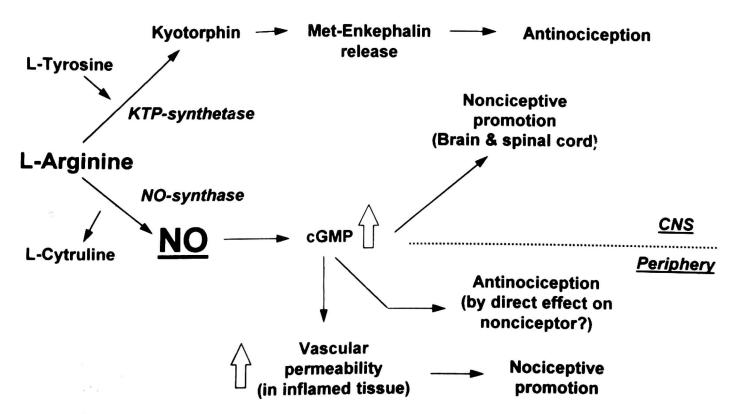


Fig. 3. The suggested participation of L-Arg and NO in central and peripheral antinociception. (according to A. Kawabata et al. 1993).

peripherally mediated nociception are not consistent. A histochemical study has suggested that NO may act as a messenger from sensory neurons in dorsal root ganglia to their satellite cells, where c-GMP levels increase in response to NO (148). Thus, it has been suggested that L-Arg may act as a nociceptive

promoter through the enhancement in NO production at spinal and supraspinal levels (147-149). Thus, it is obvious that NO-pathway may be differently involved in nociceptive processes at the level of the pheripheral and central nervous system.

It is also suggested, that L-Arg itself plays a dual role in the nociceptive processing in brain, being nociceptive via the NO/cGMP pathway, and antinociceptive via the kyotorphin-Met-enkephalin pathway (149, 150) (Fig. 3). Kyotorphin (L-tyrosyl-L-arginine), an endogenous peptide isolated from bovine brain, produces naloxone-reversible antinociception by enhancing Met-enkephalin release, and is localized in synaptosomes of CNS (151). It is suggested that L-Arg acts as a precursor for the biosynthesis of kyotorphin, that results in antinociception (147, 148). The therapeutical significance of L-Arg is also supported by clinical findings, presenting that intravenous infusion of L-Arg, produces potent analgesia in the naloxone-reversible manner in patients with various types of chronic pain (152, 153).

The participation of NO in specificity of the regulation of the cerebrovascular tonus was also reported (154). On the contrary to the coronary, and mesentery vascular beds, the relaxation caused by transmural neural stimulation of basilar, middle cerebral and posterior cerebral arteries of dogs were suppressed by L-NA. This suppression was reversed in the presence of exogenous L-Arg (154, 155). Thus, NO may play a crucial role in the genesis of neurally induced vasodilatation of cerebral arteries, such as migraine.

Hypoxia increased the c-GMP level in main rabbit cerebral arteries, and increased the cerebral blood flow (156). This effect is prevented in the presence of methylene blue, the known inhibitor of guanylate cyclase. Exogenous L-Arg dilates rat pial arterioles by NO-dependent mechanisms, and increases blood flow during focal cerebral ischaemia (157). Thus, L-Arg or NO-donors were suggested to be useful for the increase of cerebral blood flow during ischaemic stroke in men (157).

Endothelins and the nervous system

In March 1988 Yanagisawa and co-workers (6) described an endothelium derived vasoconstrictor peptide, endothelin, with the regional homologies to a group of neurotoxins, and suggested its action as an endogenous modulator of voltage-dependent ion channels. Subsequent studies resulted in the identification of several endothelin (ET) peptide isoforms and the genes that encode them. The ETs (Fig. 4) include three isopeptides ET-1, ET-2 and ET-3. They all contain 21 aminoacids and two disulfide bonds. ET-1 differs from ET-2 by two and from ET-3 by six amino acids, and each peptide is derived from a seperate gene (158). In humans, ET-1 m-RNA codes for a 212-amino acid precursor (prepro-ET-1), which undergoes proteolytic cleavage to

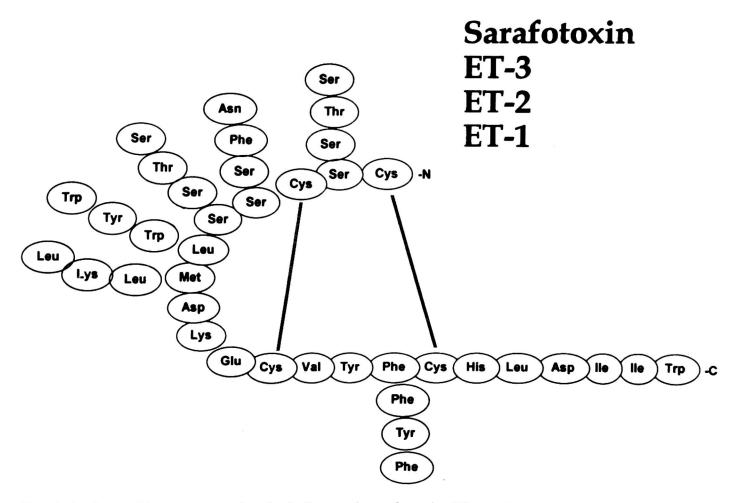


Fig. 4. Amino acid sequences of endothelins and sarafotoxin. The main sequence ET-1 is presented, when the differences in substituing amino acids are consecutively presented for ET-2, ET-3 and sarafotoxin.

a 38-aminoacid residue (pro- or "big" ET-1), and subsequently, through the action of a metalloprotease (ET-converting enzyme) to ET-1 (159). The ETs are structurally and functionally homologous to another mammalian peptides: vasoactive intestinal contractor (160) and to sarafotoxins found in the venom of the snake and burrowing asp, *Atractaspis engaddensis* (161). ET-1 is the only ET produced by human and porcine endothelial cells, while both ET-1 and ET-3 have been identified in neural tissue such as brain, spinal cord and dorsal root ganglia (162, 163). Ischaemia, thrombin, transforming growth factor-β, norepinephrine, phorbol esters and calcium ionophore A23187, were reported to activate release of ETs from endothelium (164, 165). In experimental animals ET-1 has a greater vasoconstrictor potency than any other known vasoactive hormone causing the long-lasting contraction of the vascular smooth muscle (164, 166, 167).

Although three peptides induce the potent vasoconstriction in vitro, the transient depressor response followed by a sustained pressor response in vivo is observed. ET-1 is the most active vasoconstrictor, while ET-3 is the most potent vasodilator (158, 168, 169). In addition to that, endothelins have been reported to produce a wide spectrum of biological effects, such as: stimulation of proliferation of vascular smooth muscle cells and fibroblasts (170),

contraction of human airway and intestinal smooth muscle (171, 172), positive inotropic and chronotropic effects on the myocardium (173), release of eicosanoids from vascular beds (172), stimulation of atrial natriuretic peptide (174). They stimulate the vasopressin and oxitocine release from hypothalamus (175) and modulate norepinephrine release from sympathetic terminals (176). ET-1 is equipotent with ET-3 in releasing EDRF/NO from vascular endothelium (172, 177, 178), and the anti-platelet and fibrinolytic properties of ET-3 in vivo, may be partially related to this effect (179). Radioimmunoassay has demonstrated low plasma levels of ET-1 (ca 1 pg/ml for healthy volunteers). It increased (up to 20 pg/ml) in acute stages of renal failure (180), endotoxic shock (181), myocardial infarction, pulmonary hypertension (182, 183) and uraemia (184). During acute asthmatic episodes, ET-1 levels rise in bronchial exudates (185).

ETs affect target cells by binding to receptors on the cell surface. Genes encoding multiple ET receptors have been cloned and expressed, revealing both ET-1 preferring (ET_A) and isopeptide-nonselective (ET_B) receptor subtypes, and the corresponding mRNAs have been detected also in mammalian brain (186 – 188). Activation of receptors by ETs in smooth muscle as well as in cultured astrocytes neublastoma and C6 glioma cells, stimulates Ca⁺⁺ influx and intracellular Ca⁺⁺ mobilisation, activates phospholipases A, and C, protein kinase C, activates Na+-H+ exchange, induces transcription of the c-fos protooncogene, and inhibits Na+-K+-ATP-ase (189-192). In contrast, ET-1 does not stimulate Ca++ influx or alter [Ca++], levels in rat brain synaptosomes (193), indicating, that neuronal ET receptors involved in Ca⁺⁺ signalling may be localized preferentially with postsynaptic elements. It is suggested that in neuroblastoma x glioma NG108-15 cells responses to ET-1 involve receptor-mediated Ca++ influx and mobilisation of Ca++ from inositol phosphate-sensitive intracellular stores, while plateau responses result from Ca⁺⁺ influx through dihydropyridine-sensitive voltage-gated channels (175, 194).

ETs were found to modify some of neuronal functions. ET-1 applied locally into area postrema of rats caused an increase, followed by a decrease in mean arterial blood pressure (195). ET-1 inhibits the release of norepinephrine from sympathetic neurons (196), and stimulates the release of acetylcholine from parasympathetic neurons (197), vasopressin from hypothalamus (198) and aspartate from cerebellar granule cells (199).

ETs seem to play an important role in the pathology of CNS connected with the functional effects on cerebrovascular and neural tissues. ETs induce longlasting constrictions of mammalian cerebral arteries and arterioles (200-202), including human vertebral, basilar, and middle cerebral arteries (203) as it was shown *in vitro*, and pial arterioles and basilar artery *in vivo* (204). It was demonstrated that ETs being large molecules, did not cross the

blood/brain barrier, and thus, they may act from the adventital-side of cerebral blood vessels (204, 204). This suggestion is confirmed by observations, that intercisternal, but not inraarterial application of ET-1 decreases the blood flow in canine basilar arteries (204), and argues for the action of the locally biosynthesized ETs in the CNS pathology.

Ischaemia was primarily found to activate ETs production by porcine aortic endothelial cells (6), coronary artery endothelium in culture (206), isolated mesenteric arteries (207), and the increase of the plasma ETs-levels were reported during myocardial infarction (208) and hypertension (209). Antibodies to ET-1 have been reported to attenuate the experimental myocardial (210), and renal (211) ischaemia. On the other hand, carbon dioxide induced hypoxia did not activate ETs release from cerebral vessel endothelial cells in culture (212), and unlike the patients with subarachnoid haemorrhage, the ETs-like radioimmunoactive material was not detected in cerebrospinal fluid (CSF) (213, 214). Even the clinical significance of the reported elevations of plasma (215, 216), and CSF (215, 216) ETs-like material following subarachnoid haemorrhage is unclear, since no correlation between the ETs level in CSF and the incidence of vasospasm was found (216). Vasospasm and ischaemic deficits in hemorrhage are commonly related to the subarachnoid blood (217). ETs might promote ischaemic neuronal injury by the high input of Ca++ ions, and indirectly, by stimulating the release of excitotoxic amino acids (199), while vasospasm may also be related to the presence of hemoglobin, which scavenges, and thus prevents the vasodilatory action of EDRF/NO (25). The isolated cerebral arteries of rats with subarachnoid haemorrhage were found to be more sensitive to vasoconstrictory properties of ETs (218). Alternatively ETs were reported to activate NO release and cGMP accumulation not only in the different vascular beds (172, 177, 178), but also in neuroblastoma x rat glioma hybrid cells (219). Thus, the observed vascular and the delayed ischemic deficit results from the complicated paralelly running processes.

It seems that ETs are mostly involved in pathological events in our body. They may mediate fatal contractions of our microvasculature and hypertension. They may promote the tumor growth and atherosclerosis development by mitogenic activity. They promote the cell death by overloading with Ca⁺⁺ ions. And the activation of vasodilatory EDRF/NO or PGI₂ or PGE₂ (220) may not counteract its pathogenicity.

Arachidonic acid metabolites

Eicosanoids are biological mediators derived from arachidonic acid (AA), an essential 20-carbon fatty acid of membrane phospholipids (221 - 223). AA may be metabolized by either cyclooxygenases (COX) forming prostacyclin (PGI₂), prostaglandins (PGs) or thromboxane (TXA₂), or by lipoxygenases

(LOX), leading to formation of leukotrienes (LTs) and dihydroxy fatty acids (HETEs). Lipoxygenases with specificity for the 5-, 12-, or 15- position have been characterized (222), and lipoxins (LXs) are compounds resulting from AA metabolism by two different LOX (221). LXs are mostly involved in the immune/inflammatory processes in our body (221).

The levels of prostanoids in brain tissue and cerebrospinal fluid are rather low (222, 223), however, the injury to a neural tissue such as trauma, ischaemia, hypoxia, reperfusion or subarachnoid haemorrhage causes a considerable increase in amounts of free arachidonic acid and its metabolites in brain (224, 225) and CSF (225, 226).

There are marked species differences in the ability of AA conversion to the different eicosanoids. In the brain of gebril and rabbits the main PGs are as follows (in the decreasing order): PGD_2 , $PGF_{2\alpha}$, $PGE_2 > 6$ -keto $PGF_{1\alpha}$, TXB_2 (227, 228), while in canine brain the low concentrations of PGD₂, with the relatively high 6-keto $F_{1\alpha}$ were found. In human brain $PGF_{2\alpha}$ is the predominating PGs followed by PGE2 and low concentrations of 6-keto $PGF_{1\alpha}$ (229). A relatively high level of 6-keto $PGF_{1\alpha}$ in the CSF argues for the biosynthesis of PGI₂ by the choroid plexus and pial vessels (224). The ability of a rabbit, gebril and human brain to generate LTs has been recently 232). demonstrated (230,231, In contrary to other sulfidopeptide-LTs (LTC₄, LTE₄, LTD₄) level is relatively low. However, when synthesized, they participate in the development of the brain injuries, such as vasoconstriction, oedema, and seizure activity (232, 233, 234).

The postischaemic production of prostanoids was extensively studied in gebril brain, because of the specificity of brain blood redistribution (the carotid arteries are the only way of supply). PGD_2 predominated in cortex and hypothalamus, PGE_2 and 6-keto $PGF_{1\alpha}$ in hippocampus. The pathophysiological meaning of this event is not clear. The increase of $PGF_{2\alpha}$ was observed in the development of cytotoxicity whereas the late accumulation of PGE_2 in cerebral tissue was coupled with vasogenic brain oedema (235, 236). In patients with stroke (237, 238), or with aneurysmal subarachnoid haemorrhage (239, 240), an imbalance between vasoconstrictory PGD_2 , $PGF_{2\alpha}$, and TXA_2 , and vasodilatatory PGI_2 levels in cerebrospinal fluid is suggested. It may be responsible for the development of vasoconstriction (241), since only PGI_2 from among the other prostanoids dilates the vessels of CNS (242).

Cerebral ischaemia is associated with the generation of oxygen free radicals, which accelerate during the reperfusion phase. Superoxide anion, hydroxyl radical, and singlet oxygen destroy endothelium and its products such as PGI₂ and EDRF/NO. Since both autacoids act synergistically in inhibiting platelet activity, the local aggregation of plateles, accompanied by the decreased plasma fibrinolytic activity, promotes local thromboembolic complications

(243). The metabolic and functional protection of the ischaemic brain of the spontaneously hypersensitive rats was observed during pretreatment with the PGI₂-analogue or TXA₂ synthase inhibitor OKY-046 or trapidil (244). So far, a selective pharmacological inhibition of TXA2 synthase has been shown to be beneficial in only a few experimental models of cerebral ischaemia. The insufficient amounts of endogenous PGI2 biosynthesized during the brain insult may be more important in brain injury pathology. In dogs with subarachnoid haemorrhage, ibuprofen (a COX inhibitor), prevented cerebral vasospasm (245), whilst indomethacin decreased basal cerebral blood flow, probably due to the inhibition not only of TXA2, but also PGI2 formation (246). In the treatment of experimental brain ischaemia in dogs, the best therapeutical results were obtained when PGI₂ was simultaneously administered with indomethacin and additionally with heparin (247 – 249). The beneficial effects of PGI₂ in brain injury are supposed to be related to vasodilatatory, antiplatelet/fibrinolytic and "cytoprotective" properties of this eicosanoid. On the basis of experimental work, it is stressed, that the beneficial effect of PGI₂ administration is observed only when it is administered before, or during the initial period of brain injury (250, 251). The intensive research in these fields is in progress and the other combinations (for example, with calcium channel blockers) have recently been proposed (252).

As the brain cannot repair itself by increasing the number of neurons, the early preventive therapeutical intervention is necessary in case of any signs of cerebellar insufficient blood supply. The surgery with carotid endarterectomy is the common intervention with recommended antiplatelet and fibrinolytic therapy (253-255).

Aspirin alone, or combined with dipyridamole, sulphinpyrazone, warfarin, heparin, streptokinase or ticlopidine is commonly used in secondary stroke prevention (256, 257). The latest meta-analysis of seven randomized, controlled trials, in which the effectiveness of aspirin in the treatment of 6409 patients with TIA and minor strokes was examined, demonstrated the significant risk reduction for total death, total strokes and cardiovascular death in patients receiving this compound (257). These trials have proved that aspirin is preventive in patients with TIA and minor strokes.

The preliminary trials of the treatment of ischaemic stroke with PGI₂ pointed at a significant alleviation of neurological deficit which occurred at 6 and 54 hour after the treatment with PGI₂, however, this improvement in two weeks after the treatment was no more statistically different from the group of patients receiving the conventional therapy (258). The placebo-controlled, randomized trials (259, 260) demonstrated that between the period of 2 weeks and 18 months there is no evidence for a therapeutic benefit from prostacyclin given even in the maximally tolerated dosage, either intermittently or continuously for 30–64 hours to patients with an acute,

completed stroke. Thus, prostacyclin alone seems to be too weak to prevent all signs of ischaemic insult in completed stroke, but it does not exclude its usefulness (also in combined therapy with TXA₂, or LOX inhibitors) in patients with TIA.

CONCLUSIONS

Discovery of the neuronal source and the functions of the labile substances, found previously to originate from endothelial cells, opened the large possibility for the research, and new ways to understanding the physiology and pathology of our nervous system. These substances affect neurons, glia, endothelial cells, vascular smooth muscle and platelets by elevating cyclic nucleotide levels, modifying $[Ca^{++}]_i$ levels, and the release of neurotransmitters. Their disturbed biosynthesis contributes to the development of atherosclerosis, as well as to ischaemic neuronal injury associated with stroke and subarachnoid haemorrhage. For most of these substances (maybe, excluding PGI_2 , and TXA_2), in spite of intensive study, it is too early for the final statement concerning the therapeutical implications of drugs modifying their biosynthesis.

REFERENCES

- 1. Moncada S, Gryglewski R, Bunting S, Vane JR. An enzyme isolated from arteries transforms prostaglandin endoperoxides to an unstable substance that inhibits platelet aggregation. *Nature* 1976; 263: 663-665.
- 2. Buchmann MR, Hass TA, Lagarde M, Guichardant M. 13-Hydroxyoctadecadienoic acid is the vessel wall chemorepellant factor, LOX. J Biol Chem 1985; 260: 16056-16059.
- 3. Hepkins NK, Oglesby TD, Bundy GL, Gorman RR. Biosynthesis and metabolism of 15-hydroperoxy 5, 8, 11, 13-eicosatetraenoic acid by human umbilical vein endothelial cells. J Biol Chem 1984; 259: 14048-14053.
- 4. Furchgott RF, Zawadzki JV. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature* 1980; 288: 373-376.
- 5. Palmer R. J. M, Ferridge A.G, Moncada S. Nitric oxide release from vascular endothelial cells accounts for the biological activity of endothelium-derived relaxing factor. *Nature* 1987; 327: 524-526.
- 6. Yanagisawa M, Kurihara H, Kimura S et al. A novel potent vasoconstrictor peptide produced by vascular endothelial cells (see comments). *Nature* 1988; 322: 411-415.
- 7. Collen D, Lijnen HR. Tissue-type plasminogen activator. Mechanisms of action and thrombolytic properties. *Haemostasis* 1986; 16 Suppl 3: 25-32.
- 8. Van Moutrik JA, Lawrance DA, Loskutoff DJ. Purification of an inhibitor of plasminogen activator (anti-activator) synthetised by endothelial cells. *J Biol Chem* 1984; 259: 14914-14921.

- 9. Morganti Kossmann MC, Kossmann T, Wahl SM. Cytokines and neuropathology. *Trends Pharmacol Sci* 1992; 13: 286-291.
- 10. Bevilacqua MP. Endothelial-leukocyte adhesion molecules. *Annu Rev Immunol* 1993; 11: 767-804.
- 11. Bevilacqua MP, Nelson RM. Selectins. J Clin Invest 1993; 91: 379-387.
- 12. Strieter RM, Kasahara K, Allen RM, et al. Cytokine-induced neutrophil-derived interleukin-8. Am J Pathol 1993; 141: 397-407.
- 13. Ross R, Raines EW, Bowen Pope DF. The biology of platelet-derived growth factor. *Cell* 1986; 46: 155-169.
- 14. Zimmerman GA, McIntyre TM, Prescott SM. Production of platelet-activating factor by human vascular endothelial cells: evidence for a requirement for specific agonists and modulation by prostacyclin. *Circulation* 1985; 72: 718-727.
- 15. Block ER, Patel JM, Sheridan NP. Effect of oxygen and endotoxin on lactate dehydrogenase release, 5-hydroxytryptamine uptake, and antioxidant enzyme activities in endothelial cells. *J Cell Physiol* 1985; 122: 240-248.
- 16. Tsao PS, Ma XL, Lefer AM. Activated neutrophils aggravate endothelial dysfunction after reperfusion of the ischemic feline myocardium. *Am Heart J* 1992; 123: 1464-1471.
- 17. Beckman JS, Beckman TW, Chen J, Marshall PA, Freeman BA. Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. *Proc Natl Acad Sci USA* 1990; 87: 1620–1624.
- 18. Palmer RM, Ashton DS, Moncada S. Vascular endothelial cells synthesize nitric oxide from L-arginine. *Nature* 1988; 333: 664-666.
- 19. Palmer RM, Moncada S. A novel citrulline-forming enzyme implicated in the formation of nitric oxide by vascular endothelial cells. *Biochem Biophys Res Commun* 1989; 158: 348-352.
- 20. Thomas G, Mostaghim R, Ramwell PW. Endothelium dependent vascular relaxation by arginine containing polypeptides. *Biochem Biophys Res Commun* 1986; 141: 446-451.
- 21. Garthwaite J. Glutamate, nitric oxide and cell-cell signalling in the nervous system. *Trends Neurosci* 1991; 14: 60-67.
- 22. Hibbs JBJ, Taintor RR, Vavrin Z, Rachlin EM. Nitric oxide: a cytotoxic activated macrophage effector molecule [published erratum appears in Biochem Biophys Res Commun 1989 Jan 31; 158 (2): 624]. Biochem Biophys Res Commun 1988; 157: 87-94.
- 23. Salvemini D, de Nucci G, Gryglewski RJ, Vane JR. Human neutrophils and mononuclear cells inhibit platelet aggregation by releasing a nitric oxide-like factor. *Proc Natl Acad Sci USA* 1989; 86: 6328-6332.
- 24. Ignarro LJ. Endothelium-derived nitric oxide: actions and properties. FASEB J 1989; 3. 31-36.
- 25. Moncada S, Palmer RM, Higgs EA. Nitric oxide: physiology, pathophysiology, and pharmacology. *Pharmacol Rev* 1991; 43: 109-142.
- 26. Gillespie JS, Liu XR, Martin W. The effects of L-arginine and NG-monomethyl L-arginine on the response of the rat anococcygeus muscle to NANC nerve stimulation. *Br J Pharmacol* 1989; 98: 1080-1082.
- 27. Garthwaite J, Charles SL, Chess Wiliams R. Endothelium-derived relaxing factor release on activation of NMDA receptors suggests role as intercellular messenger in the brain. *Nature* 1988; 336: 385-388.
- 28. Bredt DS, Hwang PM, Snyder SH. Localization of nitric oxide synthase indicating a neural role for nitric oxide. *Nature* 1990; 347: 768-770.
- 29. Pollock JS, Forstermann U, Mitchell JA, et al. Purification and characterization of particulate endothelium-derived relaxing factor synthase from cultured and native bovine aortic endothelial cells. *Proc Natl Acad Sci USA* 1991; 88: 10480-10484.

- 30. Moncada S, Palmer RM, Higgs EA. Biosynthesis of nitric oxide from L-argine. A pathway for the regulation of cell function and communication. *Biochem Pharmacol* 1989; 1709-1715.
- 31. Nathan C. Nitric oxide as a secretory product of mammalian cells. FASEB J 1992; 6: 3051-3064.
- 32. Kondo K, Mitchell JA, de Nucci G, Vane JR. Simultaneous measurement of endothelium-derived relaxing factor by bioassay and guanylate cyclase stimulation. *Br J Pharmacol* 1989; 98: 630-636.
- 33. Rapoport RM, Draznin MB, Murad F. Endothelium-dependent vasodilator-and nitrovasodilator-induced relaxation may be mediated through cyclic GMP formation and cyclic GMP-dependent protein phosphorylation. Trans Assoc Am Physicians 1983; 96: 19-30.
- 34. Ress DD, Palmer RM, Moncada S. Role of endothelium-derived nitric oxide in the regulation of blood pressure. *Proc Natl Acad Sci USA* 1989; 86: 3375-3378.
- 35. Chyu KY, Guth PH, Ross G. Effect of N omega-nitro-L-arginine methyl ester on arterial pressure and on vasodilator and vasoconstrictor responses: influence of initial vascular tone. *Eur J Pharmacol* 1992; 212: 159-164.
- 36. Vallance P, Leone A, Calver A, Collier J, Moncada S. Accumulation of an endogenous inhibitor of nitric oxide synthesis in chronic renal failure. *Lancet* 1992; 339: 572-575.
- 37. Fickling S. A, Williams D, Vallance P, Nussey S. S, Whitley G. St. J. Plasma concentrations of endogenous inhibitor of nitric oxide synthesis in normal pregnancy and pre-eclampsia. *Lancet* 1993; 342:
- 38. Schroder H, Noack E, Muller R. Evidence for a correlation between nitric oxide formation by cleavage of organic nitrates and activation of guanylate cyclase. *J Mol Cell Cardiol* 1985; 17: 931-934.
- 39. Gerzer R, Drummer C, Karrenbrock B, Heim JM. Inhibition of platelet activating factor-induced platelet aggregation by molsidomine, SIN-1, and nitrates in vitro and ex vivo. *J Cardiovasc Pharmacol* 1989; 14 Suppl 11: S115-S119.
- 40. Kuhn M, Forstermann U. Endothelium-dependent vasodilatation in human epicardial coronary arteries: effect of prolonged exposure to glyceryl trinitrate or SIN-1. *J Cardiovasc Pharmacol* 1989; 14 Suppl 11: S47-S54.
- 41. Azuma H, Ishikawa M, Sekizaki S. Endothelium-dependent inhibition of platelet aggregation. Br J Pharmacol 1986; 88: 411-415.
- 42. Furlong B, Henderson AH, Lewis MJ, Smith JA. Endothelium-derived relaxing factor inhibits in vitro platelet aggregation. *Br J Pharmacol* 1987; 90: 687-692.
- 43. Radomski MW, Palmer RM, Moncada S. Comparative pharmacology of endothelium-derived relaxing factor, nitric oxide and prostacyclin in platelets. *Br J Pharmacol* 1987; 92: 181-187.
- 44. Dembińska-Kiec A, Żmuda A, Wenhrynowicz O, Stachura J, Peskar B. A, Gryglewski R. J. Selectin-P mediated adherence of platelets to neutrophils is regulated by prostanoids and nitric oxide. *Tissue Reaction* 1993; in press:
- 45. Bult H, Fret HR, Herman AG. Interaction between SIN-1 and prostacyclin in inhibiting platelet aggregation. *J Cardiovasc Pharmacol* 1989; 14 Suppl 11: S120-S123.
- 46. Gryglewski RJ, Korbut R, Trąbka Janik E, Zembowicz A, Trybulec M. Interaction between NO donors and iloprost in human vascular smooth muscle, platelets, and leukocytes. *J Cardiovasc Pharmacol* 1989; 14 Suppl 11: S124-S128.
- 47. Gryglewski RJ, Palmer RM, Moncada S. Superoxide anion is involved in the breakdown of endothelium-derived vascular relaxing factor. *Nature* 1986; 320: 454-456.
- 48. Martin W, Villani GM, Jothianandan D, Furchgott RF. Selective blockade of endothelium-dependent and glyceryl trinitrate-induced relaxation by hemoglobin and by methylene blue in the rabbit aorta. *J Pharmacol Exp Ther* 1985; 232: 708-716.

- 49. Ding AH, Nathan CF, Stuehr DJ. Release of reactive nitrogen intermediates and reactive oxygen intermediates from mouse peritoneal macrophages. Comparison of activating cytokines and evidence for independent production. *J Immunol* 1988; 141: 2407-2412.
- 50. Stuehr DJ, Marletta MA. Induction of nitrite/nitrate synthesis in murine macrophages by BCG infection, lymophokines, or interferon-gamma. *J Immunol* 1987; 139: 518-525.
- 51. Stuehr DJ, Cho HJ, Kwon NS, Weise MF, Nathan CF. Purification and characterization of the cytokine-induced macrophage nitric oxide synthase: an FAD-and FMN-containing flavoprotein. *Proc Natl Acad Sci USA* 1991; 88: 7773-7777.
- 52. McCall TB, Palmer RM, Moncada S. Interleukin-8 inhibits the induction of nitric oxide synthase in rat peritoneal neutrophils. *Biochem Biophys Res Commun* 1992; 186: 680-685.
- 53. Cunha FQ, Moncada S, Liew FY. Interleukin-10 (IL-10) inhibits the induction of nitric oxide synthase by interferon-gamma in murine macrophages. *Biochem Biophys Res Commun* 1992; 182: 1155-1159.
- 54. Stuehr DJ, Nathan CF. Nitric oxide. A macrophage product responsible for cytostasis and respiratory inhibition in tumor target cells. J Exp Med 1989; 169: 1543-1555.
- 55. Hibbs JBJ, Taintor RR, Vavrin Z. Macrophage cytotoxicity: role for L-arginine deiminase and imino nitrogen oxidation to nitrite. *Science* 1987; 235: 473-476.
- 56. Hibbs JBJ, Vavrin Z, Taintor RR. L-arginine is required for expression of the activated macrophage effector mechanism causing selective metabolic inhibition in target cells. J Immunol 1987; 138: 550-565.
- 57. Wisseman CLJ, Waddell A. Interferonlike factors from antigen- and mitogen-stimulated human leukocytes with antirickettsial and cytolytic actions on Rickettsia prowazekii. Infected human endothelial cells, fibroblasts, and macrophages. J Exp Med 1983; 157: 1780-1793.
- 58. Nava E, Palmer RM, Moncada S. Inhibition of nitric oxide synthesis in septic shock: how much is beneficial? [see comments]. Lancet 1991; 338: 1555-1557.
- 59. Thiemermann C, Vane J. Inhibition of nitric oxide sytnesis reduces the hypotension induced by bacterial lipopolysaccharides in the rat in vivo. Eur J Pharmacol 1990; 182: 591-595.
- 61. Hibbs JBJ, Westenfelder C, Taintor R, et al. Evidence for cytokine-inducible nitric oxide synthesis from L-arginine in patients receiving interleukin-2 therapy [published erratum appears in J Clin Invest 1992; Jul; 90(1): 295]. J Clin Invest 1992; 89: 867-877.
- 62. Biliar TR, Ochoa JB, Curti B, et al. Increased circulating nitrogen oxides after human tummor immunotherapy correlated with toxic hemodynamic changes. In Biology of Nitric Oxide S. Moncada, MA. Marletta, JB Hibbs. Jr, (eds). London: Second International Meeting, 1991; pp.
- 63. Ochoa JB, Udekwu AO, Billiar TR, et al. Nitrogen oxide levels in patients after trauma and during sepsis. Ann Surg 1991; 214: 621-626.
- 64. Hibbs JBJr, Taintor RR, Vavrin V, et al. Synthesis of nitric oxide from a guanidino nitrogen of L-arginine: a molecular mechanism that targets interacellular iron. In: Nitric Oxide from L-Arginine: A Bioregulatory System. S. Moncada, EA. Higgs, (eds.) Amsterdam: Elsevier, 1990: 189-223.
- 65. Reif DW, Simmons RD. Nitric oxide mediates iron release from ferritin. Arch Biochem Biophys 1990; 283: 537-541.
- 66. Myers PR, Minor RLJ, Guerra RJ, Bates JN, Harrison DG. Vasorelaxant properties of the endothelium-derived relaxing factor more closely resemble S-nitrosocysteine than nitric oxide. *Nature* 1990; 345: 161-163.
- 67. Marletta MA. Mammalian synthesis of nitrite, nitrate, nitric oxide, and N-nitrosating agents. Chem Res Toxical 1988; 1: 249-257.
- 68. Gross SS, Stuehr DJ, Aisaka K, Jaffe EA, Levi R, Griffith OW. Macrophage and endothelial cell nitric oxide synthesis: cell-type selective inhibition by NG-aminoarginine, NG-nitroarginine and NG-methylarginine. Biochem Biophys Res Commun 1990; 170: 96-103.

- 69. Bredt DS, Snyder SH. Isolation of nitric oxide synthetase, a calmodulin-requiring enzyme. Proc Natl Acad Sci USA 1990; 87: 682-685.
- 70. Berde CB. Nitric oxide: a role in neurotransmission of neuronal toxity? IASP Newsletter 1992; 2-4.
- 71. Lambert LE, Whitten JP, Baron BM, Cheng HC, Doherty NS, McDonald IA. Nitric oxide synthesis in the CNS, endothelium and macrophages differs in its sensitivity to inhibition by arginine analogues. *Life Sci* 1991, 48, 69-75.
- 72. Ferrendelli JA, Chang MM, Kinscherf DA. Elevation of cyclic GMP levels in central nervous system by excitatory and inhibitory amino acids. *J Neurochem* 1974; 22: 535-540.
- 73. Deguchi T, Yoshioka M. L-Arginine identified as an endogenous activator for soluble guanylate cyclase from neuroblastoma cells. *J Biol Chem* 1982; 257: 10147-10151.
- 74. Knowles RG, Palacios M, Palmer RM, Moncada S. Formation of nitric oxide from L-arginine in the central nervous system: a transduction mechanism for stimulation of the soluble guanylate cyclase. *Proc Natl Acad Sci USA* 1989; 86: 5159-5162.
- 75. Klatt P, Heinzel B, John M, Kastner M, Bohme E, Mayer B. Ca2+/calmodulin-dependent cytochrome c reductase activity of brain nitric oxide synthase. *J Biol Chem* 1992; 267: 11374-11378.
- 76. Knowles RG, Palacios M, Palmer RM, Moncada S. Kinetic characteristic of nitric oxide synthase from rat brain [see comments]. *Biochem J* 1990; 269: 207-210.
- 77. Forstermann U, Gorsky LD, Pollock JS, Schmidt HH, Heller M, Murad F. Regional distribution of EDRF/NO-synthesizing enzyme(s) in rat brain. *Biochem Biophys Res Commun* 1990; 168: 727-732.
- 78. Bredt DS, Snyder SH. Nitric oxide mediates glutamate-linked enhancement of cGMP levels in the cerebellum. *Proc Natl Acad Sci USA* 1989; 86: 9030-9033.
- 79. Garthwaite J. Cellular uptake disguises action of L-glutanate on N-methyl-D-aspartate receptors. With an appendix: diffusion of transported amino acids into brain slices. Br J Pharmacol 1985; 85: 297-307.
- 80. Wood PL, Steel D, McPherson SE, Cheney DL, Lehmann J. Antagonism of N-methyl-D-aspartate (NMDA) evoked increases in cerebellar cGMP and striatal ACh release by phencyclidine (PCP) receptor agonists: evidence for possible allosteric coupling of NMDA and PCP receptors. Can J Physiol Pharmacol 1987; 65: 1923-1927.
- 81. Garthwaite J, Southam E, Anderton M. A kainate receptor linked to nitric oxide synthesis from arginine. J Neurochem 1989; 53: 1952-1954.
- 82. Garthwaite J, Garthwaite G. Cellular origins of cyclic GMP responses to excitatory amino acid receptor agonists in rat cerebellum in vitro. J Neurochem 1987; 48: 29-39.
- 83. East SJ, Garthwaite J. Nanomolar N(G)-nitroarginine inhibits NMDA-induced cyclic GMP formation in rat cerebellum. Eur J Pharmacol 1990; 184: 311-313.
- 84. Komuru H, Rakic P. Modulation of Neuronal Migration by NMDA Receptors. Science 1993; 260: 95-97.
- 85. de Vente J, Manshanden CG, Sikking RA. A functional parameter to study heterogeneity of glial cells in rat brain slices: cyclic guanosine monophosphate production in artial natriuretic factor (ANF)-responsive cells. Glia 1990; 3; 1: 43-54.
- 86. Collingride GL, Singer W. Excitatory amino acid receptors and synaptic plasticity. *Trends Pharmacol Sci* 1990; 11: 290-296.
- 87. Muller CM, Best J. Ocular dominance plasticity in adult cat visual cortex after transplantation of cultured astrocytes. *Nature* 1989; 342: 427-430.
- 88. Kotani K, Ueno S, Sano A, Kakimoto Y. Isolation and identification of methylarginines from bovine brain. J Neurochem 1992; 58: 1127-1129.
- 89. Rothman SM, Olney JW. Excitotoxicity and the NMDA receptor. *Trends Neurosci* 1987; 10: 299-302.

- 90. Boje KM, Skolnick P. Nitric oxide does not mediate the neurotrophic effects of excitatory amino acids in cultured cerebellar granulate neurons. Eur J Pharmacol 1992; 212: 151-158.
- 91. Aruffo C, Ferszt R, Hildebrandt AG, Cervos Navarro J. Low doses of L-monosodium glutamate promote neuronal growth and differentiation in vitro. *Dev Neurosci* 1987; 9: 228-239.
- 92. Pearce IA, Cambray Deakin MA, Burgoyne RD. Glutamate acting on NMDA receptors stimulates neurite outgrowth from cerebellar granule cells. FEDS Lett 1987; 223: 143-147.
- 93. Udin SB, Scherer WJ. Restoration of the plasticity of binocular maps by NMDA after the critical period in Xenopus. Science 1990; 249: 669-672.
- 94. Wood PL, Emmett MR, Rao TS, Cler J, Mick S, Iyengar S. Inhibition of nitric oxide synthase blocks N-methyl-D-aspartate, quisqualate-, kainate-, harmaline-, and pentylenetetrazole-dependent increases in cerebellar cyclic GMP in vivo. *J Neurochem* 1990; 55: 346-348.
- 95. Balazs R, Jorgensen OS, Hack N. N-methyl-D-asparate promotes the survival of cerebellar granule cells in culture. *Neuroscience* 1988; 27: 437-451.
- 96. Balazs R, Hack N, Jorgensen OS. Selective stimulation of excitatory amino acid receptor subtypes and the survival of cerebellar granule cells in culture: effect of kainic acid. *Neuroscience* 1990; 37: 251-258.
- 97. Dawson VL, Dawson TM, London ED, Bredt DS, Snyder SH. Nitric oxide mediates glutamate neurotoxicity in primary cortical cultures. *Proc Natl Acad Sci USA* 1991; 88: 6338-6371.
- 98. Park CK, Nehls DG, Graham DI, Teasdale GM, McCulloch J. The glutamate antagonist MK-801 reduces focal ischemic brain damage in the rat. *Ann Neurol* 1988; 24: 543-551.
- 99. Buisson A, Platkine M, Boulu RG. The neuroprotective effect of a nitric oxide inhibitor in a rat model of focal cerebral ischaemia. *Br J Pharmacol* 1992; 106: 766-767.
- 100. Nowicki JP, Duval D, Poignet H, Scatton B. Nitric oxide mediates neuronal death after focal cerebral ischemia in the mouse. Eur J Pharmacol 1991; 204: 339-340.
- 101. Pape HC, Mager R. Nitric oxide controls oscillatory activity in thalamocortical neurons. Neuron 1992; 9: 441-448.
- 102. Mollace V, Bagetta G, Nistico G. Evidence that L-arginine possesses proconvulsant effects mediated through nitric oxide. *Neuroreport* 1991; 2: 269-272.
- 103. Zhang J, Su Y, Oury TD, Piantadosi CA. Cerebral amino acid, norepinephrine and nitric oxide metabolism in CNS oxygen toxicity. *Brain Res* 1993; 606: 56-62.
- 104. Oury TD, Ho YS, Piantadosi CA, Crapo JD. Extracellular superoxide dismutase, nitric oxide, and central nervous system O₂ toxicity. *Proc Natl Acad Sci USA* 1992; 89: 9715-9719.
- 105. Hartung HP, Jung S, Stoll G, et al. Inflammatory mediators in demyelinating disorders of the CNS and PNS. *J-Neuroimmunol* 1992; 40(2-3): 197-210.
- 106. Banati RB, Gehrmann J, Schubert P, Kreutzberg GW. Cytotoxicity of microglia. Glia 1993; 7: 111-118.
- 107. Corradin SB, Mauel J, Donini SD, Quattrocchi E, Ricciardi Castagnoli P. Inducible nitric oxide synthase activity of cloned murine microglial cells. Glia 1993; 7: 255-262.
- 108. Gillespie JS, Liu X, Martin W. The neurotransmitter of the nonadrenergic non-cholinergic inhibitory nerves to smooth muscle of the genital system. In: S. Moncada, EA. Higgs, eds. Oxide from L-Arginine: A Bioregulatory Nitric System. Amsterdam: Elsevier, 1990 pp. 147-164.
- 109. Barnes PJ. Non-adrenergic non cholinergic neural control of human airways. Arch Int Pharmacodyn 1986; 280: 208-229.
- 110. Desai KM, Zembowicz A, Sessa WC, Vane JR. Nitroxergic nerves mediate vagally induced relaxation in the isolated stomach of the guinea pig. *Proc Natl Acad Sci USA* 1991; 88: 11490-11494.

- 111. Snyder SH, Bredt DS. Nitric oxide as a neuronal messenger. *Trends Pharmacol Sci* 1991; 12: 125-128.
- 112. Dembińska-Kieć A, Żmuda A, Miotla J, Gryglewski RJ. Involvement of nicotinic receptors in the nitric oxide mediated non-adrenergic non-cholinergic relaxations of guinea-pig. *J Physiol Pharmacol* 1992; 43: 1-6.
- 113. Liu SF, Crawely DE, Rohde JAL, Evans TW, Barnes PJ. Role of nitric oxide and guanosine 3', 5'-cyclic monophosphate in mediating nonadrenergic, noncholinergic relaxation in guinea pig pulmonary arteries. *Br J Pharmacol* 1992; 107: 861-866.
- 114. Mawe GM, Schemann M, Wood JD, Gershon MD. Immunocytochemical analysis of potential neurotransmitters present in the myenteric plexus and muscular layers of the corpus of the guinea pig stomach. *Anat Rec* 1989; 224: 431-442.
- 115. Kaushik MD, Zembowicz A, Sessa WC, Vane JR. Nitroxergic nerves mediate vagally induced relaxation in the isolated stomach of the guinea pig. *Proc Natl Acad* 1991; 88: 11490-11494.
- 116. Juan H. Nicotinic nociceptors on perivascular sensory nerve endings. Pain 1982; 12: 259 264.
- 117. Westerman RA, Widdop RE, Hogan C, Zimmet P. Non-invasive tests of neurovascular function: reduced responses in diabets mellitus. *Neurosci Lett* 1987; 81: 177-182.
- 118. Parkhouse N, Le Quesne PM. Quantitative objective assessment of peripheral nociceptive C fibre function. J Neurol Neurosurg Psychiatry 1988; 51: 28-34.
- 119. Lynn B, Shakhanbeh J. Substance P content of the skin, neurogenic inflammation and numbers of C-fibres following capsaicin application to a cutaneous nerve in the rabbit. Neuroscience 1988; 24: 769-775.
- 120. Lembeck F, Holzer P. Substance P as neurogenic mediator of antidromic vasodilatation and neurogenic plasma extravasation. *Naunyn-Schmiedebergs Arch Phrmacol* 1979; 310: 175-183.
- 121. Naggy JI, Iversen LL, Goedert M, Chapman D, Hunt SP. Dose-dependent effects of capsaicin on primary sensory neurons in the neonatal rat. *J Neurosci* 1983; 3: 399-406.
- 122. Gamse R, Holzer P, Lembeck F. Decrease of substance P in primary afferent neurones and impairment of neurogenic plasma extravasation by capsaicin. *Br J Pharmacol* 1980; 68: 207-213.
- 123. Holzer P. Capsaicin: cellular targets, mechanisms of action, and selectivity for thin sensory neurons. *Pharmacol Rev* 1991; 43: 143-201.
- 124. Szolcsanyi J. Capsaicin, irritation and desensitization: neurophysiological basis and future perspectives. In Chemical Senses. BG Green, JR Mason, MR Kare, eds. New York: Marcel Dekker Inc., 1990: pp. 141-168.
- 125. Dalsgaard CJ, Haegerstrand A, Theodorsson Norheim E, Brodin E, Hokfelt T. Neurokinin A-like immunoreactivity in rat primary sensory neurons; coexistance with substance P. Histochemistry 1985; 83: 37-39.
- 126. Ralevic V, Khalil Z, Dusting GJ, Helme RD. Nitric oxide contributes to substance P-induced inflammation in rat blisters. In: Prog. Microcirc. Res. Proceedings of the Sixth Australian & New Zeland Symposium Printing Unit. The University of South Wales, 1991; 99-100.
- 127. Ralevic V, Khalil Z, Dusting GJ, Helme RD. Nitric oxide and sensory nerves are involved in the vasodilator response to acetylcholine but not calcitonin gene-related peptide in rat skin microvasculature. Br J Pharmacol 1992; 106: 650-655.
- 128. Gray DW, Marshall I. Nitric oxide synthesis inhibitors attenuate calcitonin gene-related peptide endothelium-dependent vasorelaxation in rat aorta. Eur J Pharmacol 1992; 212: 37-42.
- 129. Geppetti P, Del Bianco E, Tramontana M, et al. Arachidonic acid and bradykinin share a common pathway to release neuropeptide from capsaicin-sensitive sensory nerve fibers of the guinea pig heart. J Pharmacol Exp Ther 1991; 259: 759-765.

- 130. Withrington PG. The actions of two sensory neuropeptides, substance P and calcitonin generelated peptide, on the canine hepatic arterial and portal vascular beds. *Br J Pharmacol* 1992; 107: 296-302.
- 131. Lundberg JM, Saria A. Capsaicin-induced desensitization of airway mucosa to cigarette smoke, mechanical and chemical irritants. *Nature* 1983; 302: 251-253.
- 132. Petersson G, Malm L, Ekman R, Hakanson R. Capsaicin evokes secretion of nasal fluid and depletes substance P and calcitonin gene-related peptide from the nasal mucosa in the rat. Br J Pharmacol 1989; 98; 930-936.
- 133. Moskowitz MA. Reinhard JF, Romero J, Melamed E, Pettibone DJ. Neurotransmitters and fifth cranial nerve; is there a relation with the headache phase of migraine. *Lancet* 1979; ii: 883-884.
- 134. Bill A, Stjernschantz J, Mandahl A, Brodin E, Nilsson G. Substance P: release on trigeminal nerve stimulation, effects in the eye. *Acta Physiol Scand* 1979; 106: 371-373.
- 135. Lam FY, Ferrell WR. Specific neurokinin receptors mediate plasma extravasation in the rat knee joint. Br J Pharmacol 1991; 103: 1263-1267.
- 136. Pernow B. Role of tachykinins in neurogenic inflammation. *J Immunol* 1985; 135: 812s-815s.
- 137. Saria A, Lundberg JM, Skofitsch G, Lembeck F. Vascular protein linkage in various tissue induced by substance P, capsaicin, bradykinin, serotonin, histamine and by antigen challenge. Naunyn Schmiedebergs Arch Pharmacol 1983; 324: 212-218.
- 138. Evangelista S, Renzi D, Guzzi P, Surrenti C, Santicioli P, Maggi CA. Capsaicin-like activity of N-ethylmaleimide in rat stomach. *Gen Pharmacol* 1992; 23: 39-41.
- 139. Stark ME, Szurszewski JH. Role of nitric oxide in gastrointestinal and hepatic function and disease. Gastroenterology 1992; 103: 1928-1949.
- 140. Chlopicki S, Gryglewski RJ. The endothelium-dependent and the endothelium-independent vasodilators in the isolated, perfused guinea pig heart. *J Physiol Pharmacol* 1992; 43: 353-366.
- 142. Ferreira SH, Duarte ID, Lorenzetti BB. The molecular mechanism of action of peripheral morphine analgesia: stimulation of the cGMP system via nitric oxide release. Eur J Pharmacol 1991; 201: 121-122.
- 143. Duarte IDG, dos Santos IR, Lorenzetti BB, Ferreira SH. Analgesia by direct antagonism of nociceptor sensitization involves the arginine-nitric oxide-cGMP pathway. Eur J Pharmacol 1992; 217: 225-227.
- 144. Ferreira SH, Lorenzetti BB, Faccioli LH. Blockade of hyperalgesia and neurogenic oedema by topical application of nitroglycerin. Eur J Pharmacol 1992; 217: 207-209.
- 145. Tseng LF, Xu JY, Pieper GM. Increase of nitric oxide production by L-arginine potentiates i.c.v. administered beta-endorphin-induced antinociception in the mouse. *Eur J Pharmacol* 1992; 212: 301-303.
- 146. Ialenti A, Ianaro A, Moncada S, Di Rosa M. Modulation of acute inflammation by endogenous nitric oxide. Eur J Pharmacol 1992; 211: 177-182.
- 147. Przewlocki R, Machelska H, Przewlocka B. Inhibition of nitric oxide synthase enhances morphine antinociception in rat spinal cord. Life Sciences 1993; 53: 1-5.
- 148. Morris R, Southam E, Braid DJ, Garthwaite J. Nitric oxide may act as a messenger between dorsal root ganglion neurones and their satellite cells. *Neurosci Lett* 192; 137: 29-32.
- 149. Kawabata A, Umeda N, Takagi H. L-arginine exerts a dual role in nociceptive processing in the brain: involvement of the kyotorphin-Met-enkephalin pathway and NO-cyclic GMP pathway. Br J Pharmacol 1993; 109: 73-79.
- 150. Kawabata A, Nishimura Y, Takagi H. L-Leucyl-L-arginine, naltrindole and D-arginine block antinociception elicited by L-arginine in mice with carrageenin-induced hyperalgesia. *Br J Pharmacol* 1992; 107: 1096-1101.

- 151. Takagi H, Shiomi H, Ueda H, Amano H. Morphine-like analgesia by a new dipeptyde, L-tyrosyl-L-arginine (kyotorphin) and its analogue. Eur J Pharmacol 1979; 55: 109-111.
- 152. Takagi H, Harima A, Shimizu H. A novel clinical treatment of persistent pain with L-arginine. Eur J Pharmacol 1990; 183: 1443.
- 153. Harima A, Shimizu H, Takagi H. Analgesic effect of L-arginine in patients with persistent pain. Eur Neuropsychopharmacol 1991; 1: 529-533.
- 154. Toda N. Okamura T. Modification by L-NG-monomethyl arginine (L-NMMA) of the response to nerve stimulation in isolated dog mesenteric and cerebral arteries. *Jpn J Pharmacol* 1990; 52: 170-173.
- 155. Toda N, Minami Y, Okamura T. Inhibitory effects of L-NG-nitro-arginine on the synthesis of EDRF and the cerebroarterial response to vasodilator nerve stimulation. *Life Sci* 1990; 47: 345-351.
- 156. Pearce WJ, Reynier Rebuffel AM et al. Effects of methylene blue on hypoxic cerebral vasodilatation in the rabbit. J Pharmacol Exp Ther 1990; 254: 616-625.
- 157. Morikawa E., Rosenblatt S., Moskowitz M. L-Arginine dilates rat pial arterioles by nitric oxide-dependent mechanisms and increases blood flow during focal cerebral ischaemia. *Br J Pharmacol* 1992; 107: 905-907.
- 158. Inoue A, Yanagisawa M, Kimura S, et al. The human endothelin family: three structurally and pharmacologically distinct isopeptides predicted by three separate genes. *Proc Natl Acad Sci USA* 1989; 86: 2863-2867.
- 159. McMahon EG, Palomo MA, Moore WM, McDonald JF, Stern MK. Phosphoramidon blocks the pressor activity of porcine big endothelin-1-(1-39) in vivo and conversion of big endothelin-1-(1-39) to endothelin-1-(1-21) in vitro. *Proc Natl Acad Sci USA* 1991; 88: 703-707.
- 160. Saida K, Mitsui Y, Ishida N. A novel peptide, vasoactive intestinal contractor, of a new (endothelin) peptide family. Molecular cloning, expression, and biological activity. *J Biol Chem* 1989; 264: 14613-14616.
- 161. Kloog Y, Ambar I, Sokolovsky M, Kochva E, Wollberg Z, Bdolah A. Sarafotoxin, a novel vasoconstrictor peptide: phosphoinositide hydrolysis in rat heart and brain. *Science* 1988; 242: 268-270.
- 162. Lee ME, de la Monte SM, Ng SC, Bloch KD, Quertermous T. Expression of the potent vasoconstrictor endothelin in the human nervous system. J Clin Invest 1990; 86: 141-147.
- 163. Giaid A, Gibson SJ, Ibrahim BN, et al. Endothelin 1, an endothelium-derived peptide, is expressed in neurons of the human spinal cord and dorsal root ganglia. *Proc Natl Acad Sci USA* 1989; 86: 7634-7638.
- 164. Yanagisawa M, Inoue A, Takuwa Y, Mitsui Y, Kobayashi M, Masaki T. The human preproendothelin-1 gene: possible regulation by endothelial phosphoinositide turnover signaling. *J Cardiovasc Pharmacol* 1989; 13 Suppl 5: S13-S17.
- 165. Masaki T. The discovery, the present state, and the future prospects of endothelin. J Cardiovasc Pharmacol 1989; 13 Suppl 5: S1-S4.
- 166. Han SP, Trapani AJ, Fok KF, Westfall TC, Knuepfer MM. Effects of endothelin on regional hemodynamics in conscious rats. Eur J Pharmacol 1989; 159: 303-305.
- 167. Minkes RK, MacMillan LA, Bellan JA, Kerstein MD, McNamara DB, Kadowitz PJ. Analysis of regional responses to endothelin in hindquarters vascular bed of cats. Am J Physiol 1989; 256: H598-H602.
- 168. Minkes RK, Coy DH, Murphy WA, McNamara DB, Kadowitz PJ. Effects of porcine and rat endothelin and an analog on blood pressure in the anesthetized cat. *Eur J Pharmacol* 1989; 164: 571-575.
- 169. Rodman DM, McMurtry IF, Peach JL, O'Brien RF. Comparative pharmacology of rat and porcine endothelin in rat aorta and pulmonary artery. Eur J Pharmacol 1989; 165: 297-300.

- 170. Takuwa N, Takuwa Y, Yanagisawa M, Yamashita K, Masaki T. A novel vasoactive peptide endothelin stimulates mitogenesis through inositol lipid turnover in Swiss 3T3 fibroblasts. J Biol Chem 1989; 264: 7856-7861.
- 171. Advenier C, Sarria B, Naline E, Puybasset L, Lagente V. Contractile activity of three endothelins (ET-1, ET-2 and ET-3) on the human isolated bronchus. *B J Pharmacol* 1990; 100: 168-172.
- 172. de Nucci G, Thomas R, D'Orleans Juste P, et al. Pressor effects of circulating endothelin are limited by its removal in the pulmonary circulation and by the release of prostacyclin and endothelium-derived relaxing factor. *Proc Natl Acad Sci USA* 1988; 85: 9797-9800.
- 173. Ishikawa T, Yanagisawa M, Kimura S, Goto K, Masaki T. Positive inotropic action of novel vasoconstrictor peptide endothelin on guinea pig atria. *Am J Physiol* 1988; 255: H970-H973.
- 174. Fukuda Y, Hirata Y, Yoshimi H, et al. Endothelin is a potent secretagogue for atrial natriuretic peptide in cultured rat atrial myocytes. *Biochem Biophys Res Commun* 1988; 155: 167-172.
- 175. Greenberg DA, Chan J. Sampson HA. Endothelins and the nervous system. *Neurology* 1992; 42: 25-31.
- 176. Wiklund NP, Ohlen A, Cederqvist B. Inhibition of adrenergic neuroeffector transmission by endothelin in the guinea-pig femoral artery. *Acta Physiol Scand* 1988; 134: 311-312.
- 177. Luscher TF, Yang Z, Tschudi M, et al. Interaction between endothelin-1 and endothelium-derived relaxing factor in human arteries and veins. *Circ Res* 1990; 66: 1088-1094.
- 178. Warner TD, de Nucci G, Vane JR. Rat endothelin is a vasodilator in the isolated perfused mesentery of the rat. Eur J Pharmacol 1989; 159: 325-326.
- 179. Korbut R, Lidbury P, Thomas GR, Vane JR. Fibrinolytic activity of endothelin-3. *Thromb Res* 1989; 55: 797-799.
- 180. Spooren PF, Vermes I, Kip L, Haanen C. Endothelin: a possible role in the occurrence of renal failure in thrombotic thrombocytopenic purpura (letter). *Thromb Haemost* 1993; 69: 401-432.
- 181. Morel DR, Lacroix JS, Hemsen A, Steinig DA, Pittet JF, Lundberg JM. Increased plasma and pulmonary lymph levels of endothelin during endotoxin shock. *Eur J Pharmacol* 1989; 167: 427-428.
- 182. Salminen K, Tikkanen I, Saijonmaa O, Nieminen M, Fyhrquist F, Frick MH. Modulation of coronary tone in acute myocardial infarction by endothelin (letter). Lancet 1989; 2: 747.
- 183. Cernacek P, Stewart DJ. Immunoreactive endothelin in human plasma: marked elevations in patients in cardiogenic shock. *Biochem Biophys Res Commun* 1989; 161: 562-567.
- 184. Koyama H, Tabata T, Nishzawa Y, Inoue T, Morii H, Yamaji T. Plasma endothelin levels in patients with uraemia. *Lancet* 1989; 1: 991-992.
- 185. Nomura A, Uchida Y, Kameyana M, Saotome M, Oki K, Hasegawa S. Endothelin and bronchial asthma (letter). Lancet 1989; 2: 747-748.
- 186. Arai H. Hori S, Aramori I, Ohkubo H, Nakanishi S. Cloning and expression of a cDNA encoding an endothelin receptor (see comments). *Nature* 1990; 348: 730-732.
- 187. Sakurai T, Yanagisawa M, Takuwa Y, et al. Cloning of a cDNA encoding a non-isopeptide-selective subtype of the endothelin receptor (see comments). *Nature* 1990; 348: 732-735.
- 188. Lin HY, Kaji EH, Winkel GK, Ives HE, Lodish HF. Cloning and functional expression of a vascular smooth muscle endothelin 1 receptor. *Proc Natl Acad Sci USA* 1992; 88: 3185-3189.
- 189. Simonson MS, Dunn MJ. Cellular signaling by peptides of the endothelin gene family. FASEB J 1990; 4: 2989-3000.

- 190. Supattapone S, Simpson AW, Ashley CC. Free calcium rise and mitogenesis in glial cells caused by endothelin. *Biochem Biophys Res Commun* 1989; 165: 1115-1122.
- 191. Marsault R, Vigne P, Breittmayer JP, Frelin C. Astrocytes are target cells for endothelins and sarafotoxin. J Neurochem 1990; 54: 2142-2144.
- 192. Marin P, Delumeau JC, Durieu Trautmann O, et al. Are several G proteins involved in the different effects of endothelin-1 in mouse striatal astrocytes? *J Neurochem* 1991; 56: 1270-1275.
- 193. Hamilton MG, Frew R, Lundy PM. Effect of endothelin on Ca²⁺ influx, intracellular free Ca²⁺ levels and ligand binding to N and L type Ca²⁺ channels in rat brain. *Biochem Biophys Res Commun* 1989; 162: 1332-1338.
- 194. Chan J, Greenberg DA. SK&F 96365, a receptor-mediated calcium entry inhibits calcium responses to endothelin-1 in NG108-15 cells. *Biochem Biophys Res Commun* 1991; 177: 1141-1146.
- 195. Ferguson AV, Smith P. Cardiovascular responses induced by endothelin microinjection into area postrema. Regul Pept 1990; 27: 75-85.
- 196. Wiklund NP, Ohlen A, Cederqvist B. Adrenergic neuromodulation by endothelin in guinea pig pulmonary artery. *Neurosci* Lett 1989; 101: 269-273.
- 197. Wiklund NP, Wiklund CU, Ohlen A, Gustafsson LE. Cholinergic neuromodulation by endothelin in guinea pig ileum. *Neurosci Lett* 1989; 101: 342-346.
- 198. Shichrii M, Hirata Y, Kanno K, Ohta K, Emori T, Marumo F. Effect of endothelin-1 on release of arginine-vasopressin from perifused rat hypothalamus. *Biochem Biophys Res Commun* 1989; 163: 1332-1337.
- 199. Lin WW, Lee CY, Chuang DM. Endothelin-1 stimulates the release of preloaded [3H]D-aspartate from cultured cerebellar granule cells. *Biochem Biophys Res Commun* 1990; 167: 593-599.
- 200. Asano T, Ikegaki I, Suzuki Y, Satoh S, Shibuya M. Endothelin and the production of cerebral vasospasm in dogs. *Biochem Biophys Res Commun* 1989; 159: 1345-1351.
- 201. Ide K, Yamakawa K, Nakagomi T, et al. The role of endothelin in the pathogenesis of vasospasm following subarachnoid haemorrhage. *Neurol Res* 1989; 11: 101-104.
- 202. Edwards R, Trizna W. Response of isolated intracerebral arterioles to endothelins. *Pharmacology* 1990; 41: 149-152.
- 203. Faraci FM. Effects of endothelin and vasopressin on cerebral blood vessels. Am J Physiol 1989; 257: H799-H803.
- 204. Mima T, Yanagisawa M, Shigeno T, et al. Endothelin acts in feline and canine cerebral arteries from the adventitial side. Stroke 1989; 20: 1553-1556.
- 205. Kadel KA, Heistad DD, Faraci FM. Effects of endothelin on blood vessels of the brain and choroid plexus. *Brain Res* 1990; 518: 78-82.
- 206. Hieda HS, Gomez Sanchez CE. Hypoxia increases endothelin release in bovine endothelial cells in culture, but epinephrine, norepinephrine, serotonin, histamine and angiotensin II do not. *Life Sci* 1990; 47: 247-251.
- 207. Kohno M, Yasunari K, Murakawa K, et al. Plasma immunoreactive endothelin in essential hypertension. Am J Med 1990; 88: 614-618.
- 208. Larkin SW, Clarke JG, Keogh BE, et al. Intracoronary endothelin induces myocardial ischemia by small vessel constriction in the dog. Am J Cardiol 1989; 64: 956-958.
- 209. Miyauchi T, Yanagisawa M, Tomizawa T, et al. Increased plasma concentrations of endothelin-1 and big endothelin-1 in acute myocardial infarction (letter). *Lancet* 1989; 2: 53-54.
- 210. Watanabe T, Suzuki N, Shimamoto N, Fujino M, Imada A. Endothelin in myocardial infarction (letter; comment). Nature 1990; 344: 114.

- 211. Kon V, Yoshioka T, Fogo A, Ichikawa I. Glomerular actions of endothelin in vivo. *J Clin Invest* 1989; 83: 1762-1767.
- 212. Yoshimoto S, Ishizaki Y, Sasaki T, Murota S. Effect of carbon dioxide and oxygen on endothelin production by cultured porcine cerebral endothelial cells. *Stroke* 1991; 22: 378-383.
- 213. Suzuki H, Sato S, Suzuki Y, Takekoshi K, Ishihara N, Shimoda S. Increased endothelin concentration in CSF from patients with subarachnoid hemorrahage. *Acta Neurol Scand* 1990; 81: 553-554.
- 214. Suzuki H, Sato S, Suzuki Y, Oka M, et al. Endothelin immunoreactivity in cerebrospinal fluid of patients with subarachnoid haemorrhage. *Ann Med* 1990; 22: 233-236.
- 215. Masaoka H, Suzuki R, Hirata Y, Emori T, Marumo F, Hirakawa K. Raised plasma endothelin in aneurysmal subarachnoid haemorrhage (letter) (see comments). *Lancet* 1989; 2: 1402.
- 216. Fujimori A, Yanagisawa M, Saito A, et al. Endothelin in plasma and cerebrospinal fluid of patients with subarachnoid haemorrhage (letter). *Lancet* 1990; 336: 633.
- 217. Kiwak KJ, Heros RC. Cerebral vasospasm after subarachnoid hemorrhage. *Trends Neurosci* 1987; 10: 89-92.
- 218. Alafaci C, Jansen I, Arbab MA, Shiokawa Y, Svendgaard NA, Edvinsson L. Enhanced vasoconstrictor effect of endothelin in cerebral arteries from rats with subarachnoid haemorrhage. *Acta Physiol Scand* 1990; 138: 317-319.
- 219. Reiser G. Endothelin and a Ca²⁺ ionophore raise cyclic GMP levels in a neuronal cell line via formation of nitric oxide. *Br J Pharmacol* 1990; 101: 722-726.
- 220. Zoja C, Benigni A, Renzi D, Piccinelli A, Perico N, Remuzzi G. Endothelin and eicosanoid synthesis in cultured mesangial cells. *Kidney Int* 1990; 37: 927-933.
- 221. Serhan CN, Hamberg M, Samuelsson B. Lipoxins: novel series of biologically active compounds formed from arachidonic acid in human leukocytes. *Proc Natl Acad Sci USA* 1984; 81: 5335-5339.
- 222. Wolfe LS., Pappius HM. Arachidonic acid metabolism in cerebral ischemia and brain injury. Cereb Ischemia 1984; 10: 223-231.
- 223. Petroni A, Socini A, Blasevich M, Borghi A, Galli C. Differential effects of various vasoactive drugs on basal and stimulated levels of TXB2 and 6-keto-PGF1 alpha in rat brain. *Prostaglandins* 1985; 29: 579-587.
- 224. Wolfe LS, Pappius HM, Pokrupa R, Hakim A. Involvement of arachidonic acid metabolites in experimental brain injury. Identification of lipoxygenase products in brain. Clinical studies on prostacyclin infusion in acute cerebral ischemia. Adv Prostaglandin Thromboxane Leukot Res 1985; 15: 585-588.
- 225. Katz B, Sofonio M, Lyden PD, Mitchell MD. Prostaglandin concentrations in cerebrospinal fluid of rabbits under normal and ischemic conditions. *Stroke* 1988; 19: 349-351.
- 226. Hsu P, Zuckerman S, Mirro R, Armstead WM, Leffer CW. Effects of ischemia/reperfusion on brain tissue prostanoids and leukotrienes in newborn pigs. *Prostaglandins* 1991; 42: 557-569.
- 227. Gaudet RJ, Alam I, Levine L. Accumulation of cyclooxygenase products of arachidonic acid metabolism in gerbil brain during reperfusion after bilateral common carotid artery occlusion. *J Neurochem* 1980; 35: 653-658.
- 228. Abdel-Halim MS, Anggard E. Regional and species differences in endogenous prostaglandin biosynthesis by brain homogenates. *Prostaglandins* 1979; 17: 411-418.
- 229. Abdel Halim MS, Lunden I, Cseh G, Anggard E. Prostaglandin profiles in nervous tissue and blood vessels of the brain of various animals. *Prostaglandins* 1980; 19: 249-258.
- 230. Dembinska Kiec A, Simmet T, Peskar BA. Formation of leukotriene C4-like material by rat brain tissue. Eur J Pharmacol 1984; 99: 57-62.

- 231. Lindgren JA, Hokfelt T, Dahlen SE, Patrono C, Samuelsson B. Leukotrienes in the rat central nervous system. *Proc Natl Acad Sci USA* 1984; 81: 6212-6216.
- 232. Moskowitz MA, Kiwak KJ, Hekimian K, Levine L. Synthesis of compounds with properties of leukotrienes C4 and D4 in gerbil brains after ischemia and reperfusion. *Science* 1984; 224: 886-889.
- 233. Simmet T, Seregi A, Hertting G. Formation of sulphidopeptide-leukotrienes in brain tissue of spontaneously convulsing gerbils. *Neuropharmacology* 1987; 26: 107-110.
- 234. Gu M, Elliott DA, Ong BY, Bose D. Possible role of leukotrienes in hypoxic contraction of canine isolated basilar artery. Br J Pharmacol 1991; 103: 1629-1632.
- 235. Bhakoo KK, Crockard HA, Lascelles PC, Avery SF. Prostaglandin synthesis and oedema formation during reperfusion following experimental brain ischaemia in the gerbil. Stroke 1984; 15: 891-895.
- 236. Kempski O, Shohami E, von Lubitz D, Hallenbeck JM, Feuerstein G. Postischemic production of eicosanoids in gerbil brain. Stroke 1987; 18: 111-119.
- 237. Egg D., Herold M, Rumpl E. Prostaglandin $F2_{\alpha}$ in cerebrospinal fluid after stroke. Lancet 1978; 1: 990.
- 238. Egg D, Herold M, Rumpl E, Gunther R. Prostaglandin F2 alpha levels in human cerebrospinal fluid in normal and pathological conditions. J Neurol 1980; 222: 239-248.
- 239. Rodriguez B, Gaetani P, Folco G, Branzoli U, Paoletti P. Cisternal and lumbar CSF concentration of arachidonate metabolites in vasospasm following subarachnoid haemorrhage from ruptured aneurysm: biochemical and clinical considerations. Surg Neurol 1985; 24: 428-432.
- 240. Rodriguez B, Gaetani P. Silvani V, Vigano T, Crivellari MT, Paoletti P. Cisternal and lumbar CSF levels of arachidonate metabolites after subarachnoid haemorrhage: an assessment of the biochemical hypothesis of vasospasm. *Acta Neurochir Wien* 1987; 84: 129-135.
- 241. Fagan SC, Castellani D, Gengo FM. Prostanoid concentrations in human CSF following acute ischaemic brain infarction. Clin Exp Pharmacol Physiol 1986; 13: 629-632.
- 242. Toda N, Miyazaki M. Responses of isolated dog cerebral and peripheral arteries to prostaglandins after application of aspirin and polyphloretin phosphate. *Stroke* 1978; 12: 490-498.
- 243. Pettigrew C, Papp A, Wu KK. Dose-related stimulation of platelet cyclic adenosine monophosphate by prostacyclin in thrombotic stroke. *Thromb Res* 1987: 45: 669-674.
- 244. Sadoshima S, Ooboshi H, Okada Y, Yao H, Ishitsuoka T, Fudjishima M. Effect of thromboxane synthetase inhibitor on cerebral circulation and metabolism during experimental cerebral ischemia in spontaneously hypertensive rats. *Eur J Pharmacol* 1989; 169: 75-83.
- 245. Chyatte D. Prevention of chronic cerebral vasospasm in dogs with ibuprofen and high-dose methylprednisolone. Stroke 1989; 20: 1021-1026.
- 246. Chemtob S, Beharry K, Rex J, Varma DR, Aranda JV. Prostanoids determine the range of cerebral blood flow autoregulation of newborn piglets. Stroke 1990; 21: 777-784.
- 247. Hallenbeck JM, Furlow TW. Jr. Prostaglandin I2 and indomethacin prevent impairment of postischemic brain reperfusion in the dog. Stroke 1979; 10: 629-637.
- 248. Hallenbeck JM, Leitch DR, Dutka AJ, Greenbaum LJJ, McKee AE. Prostaglandin I2, indomethacin, and heparin promote postischemic neuronal recovery in dogs. *Ann Neurol* 1982; 12: 145-156.
- 249. Roy MW, Dempsey RJ, Cowen DE, Donaldson DL, Young AB. Thromboxane synthetase inhibition with imidazole increases blood flow in ischemic penumbra. *Neurosurgery* 1988; 22: 317-323.
- 250. Pluta R. Experimental treatment with prostacyclin of global cerebral ischemia in rabbit-new data. Neuropatol Pol 1990; 28: 205-215.

- 251. Mossakowski MJ, Gadamski R. [Effect of prostacyclin PGI2 and indomethacin on ischemic damage of sector CA1 of Ammon's horn in the Mongolian gerbil]. Neuropatol Pol 1987; 25: 21-34.
- 252. Nikolov R. Prostacyclin as a cerebroprotective agent against brain hypoxia. *Biomed Biochim Acta* 1989; 48: S183-S187.
- 253. Kistler JP, Ropper AH, Heros RC. Therapy of ischemic cerebral vascular disease due to atherothrombosis. (2). N Engl J Med 1984; 311: 100-105.
- 254. Goyan JE. The "trials" of a long-term clinical trial: the Ticlopidine Aspirin Stroke Study and the Canadian-American Ticlopidine Study. Controlled Clin Trials 1989; 10: 236S-244S.
- 255. ESPS Group European Prevention Study. Stroke 1990; 21: 1122-1130.
- 256. Barnett HJ. Aspirin in stroke prevention. An overview. Stroke 1990; 21: IV40-IV43.
- 257. Stachenko SJ, Bravo G, Cote R, Boucher J, Battista RN. Aspirin in transient ischemic attacks and minor stroke: a meta-analysis. Fam Pract Res J 1991; 11: 179-191.
- 258. Gryglewski RJ, Nowak S, Kostka-Trabka E, et al. Treatment of ischaemic stroke with prostacyclin. Stroke 1983; 14: 197-202.
- 259. Martin JF, Hamdy N, Nicholl J, et al. Double-blind controlled trial of prostacyclin in celebral infarction. *Stroke* 1985; 16: 386-390.
- 260. Hsu CY, Faught REJ, Furlan AJ, et al. Intravenous prostacyclin in acute nonhaemorrhagic stroke: a placebo-controlled double-blind trial. Stroke 1987; 18: 352-358.

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