

D. WANG, J. C. MCGIFF, N. R. FERRERI<sup>1</sup>

## REGULATION OF CYCLOOXYGENASE ISOFORMS IN THE RENAL THICK ASCENDING LIMB: EFFECTS OF EXTRACELLULAR CALCIUM

Department of Pharmacology, New York Medical College, Valhalla, New York 10595, USA

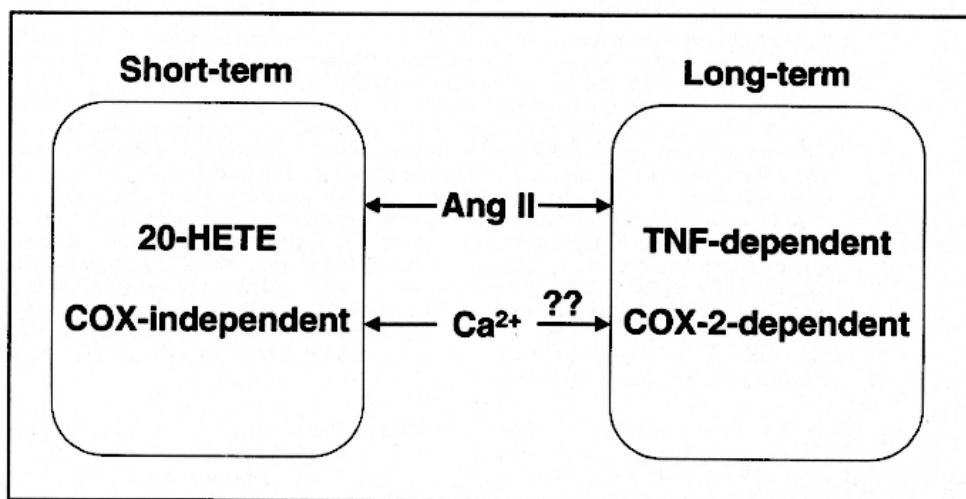
We previously showed that primary cultures of mTAL cells express cyclooxygenase 2 (COX-2) when challenged with tumor necrosis factor alpha (TNF $\alpha$ ) or phorbol myristate acetate (PMA). Moreover, expression of COX-2 was linked to decreases in TNF $\alpha$ -mediated <sup>86</sup>Rb uptake, an *in vitro* correlate of natriuresis. mTAL cells in primary culture express calcium sensing receptor (CaR), a G-protein coupled receptor that senses changes in extracellular calcium concentration and ultimately increases intracellular calcium concentration ([Ca<sup>2+</sup>]<sub>i</sub>) and protein kinase C (PKC) activity. PGE<sub>2</sub> synthesis by mTAL cells increases in a dose- and time-dependent manner after exposure of these cells to extracellular Ca<sup>2+</sup>. Similar effects were observed when cells were challenged with the CaR-selective agonist, poly-L-arginine. These data suggest that intracellular signaling mechanisms initiated *via* activation of CaR contribute to mTAL PGE<sub>2</sub> synthesis. As TNF production is calcium-sensitive in some cells types, we postulate that these effects involve the regulation of COX-2 expression *via* a TNF-dependent mechanism. The functional implications of these studies relate to a cytokine-mediated mechanism that contributes to salt and water balance, and suggests that small changes in Ca<sup>2+</sup> may contribute to the regulation of these events. The possibility that the effects of Ca<sup>2+</sup> involve activation of CaR suggests that novel calcimimetic molecules might be useful in conditions, such as hypertension or other conditions, in which manipulation of extracellular fluid volume provides beneficial effects.

**Key words:** cyclooxygenase-2, kidney, calcium-sensing receptor, tumor necrosis factor-alpha

## INTRODUCTION

There are two isoforms of prostaglandin H (PGH) synthase: commonly referred to as cyclooxygenase-1 (COX-1) and COX-2, respectively. These two enzymes catalyze the same reactions; i.e., arachidonic acid (AA) is converted to PGG<sub>2</sub> *via* the COX reaction, followed by a peroxidase reaction in which the 15-hydroperoxyl group of PGG<sub>2</sub> is reduced to the 15-hydroxyl group of PGH<sub>2</sub>. Expression of COX-2 appears to be differentially regulated in the kidney. For instance, recent studies have shown that COX isoforms are regulated in a zone-specific way by dietary salt (1) and suggest that COX-2 may

subserve different functions in the cortex (vascular resistance) and medulla (salt and water homeostasis). Recent work from our laboratory has shown that the TNF-mediated decrease in rubidium uptake, an *in vitro* correlate of natriuresis, by primary cultures of mTAL cells is COX-2 dependent (2). In addition, COX-2 expression by specific renal structures depends on the nature of the inducing stimulus. Macula densa cells expressed COX-2 when rats were placed on a low sodium diet for seven days (3), while adrenalectomy (AdrX) revealed a substantial recruitment of TAL cells containing COX-2 from the cortex towards the medulla (4). As AdrX has been associated with a decrease in sodium reabsorption by the loop of Henle (5), the preferential expression of COX-2 in the cortex rather than the medulla after AdrX may reflect a functional adaptation to conserve salt in the absence of aldosterone. COX-2 expression in the mTAL also may provide a mechanism that contributes to the regulation of renal function in a time-dependent manner. Thus, short-term regulation of ion transport in the mTAL by angiotensin II (Ang II) occurs in a COX-independent manner, while long-term regulation was TNF- and COX-2 dependent (*Fig. 1*) (2, 6). Interestingly, short-term incubations with AA



*Fig. 1.* Short- and long-term regulatory mechanisms in the mTAL. Ang II-mediated short-term (<15 min) regulatory mechanisms that affect ion transport in the mTAL are 20-HETE-dependent and COX-independent. In contrast, long-term (>3 hr) regulatory mechanisms do not appear to have a Cyp450 component, but are TNF- and COX-2-dependent. A similar paradigm may apply to the ability of extracellular calcium to affect ion transport pathways in the mTAL.

inhibited AVP-mediated increases in cAMP in the mTAL, by a pertussis toxin-sensitive mechanism that was independent of AA metabolism (7). This finding is consistent with our studies showing that short-term regulation of some renal ion transport mechanisms is COX-independent and in some

instances is mediated by cytochrome P450 metabolites such as 20-hydroxyeicosatetraenoic acid (2, 6, 8).

### *Hypercalcemia and loss of renal concentrating ability*

Disturbances in renal concentrating ability, water intake, and loop of Henle function have been associated with hypercalcemia in humans and experimental animals (9, 10). In particular, inhibition of NaCl reabsorption by the TAL in hypercalcemia has been linked to increased prostaglandin levels in the kidney (11). Moreover, in a model of hypercalcemia induced by feeding rats dihydrotachysterol, levels of phospholipase A<sub>2</sub> and COX-2, but not COX-1, were elevated in the renal cortex, outer medulla, and inner medulla. Other studies have shown that experimentally-induced hypercalcemia was associated with increased prostanoid formation in the urine and increased renal COX-2 expression (12). The locus and mechanism(s) of this effect is not fully understood. Moreover, the linkage between COX-2 expression and PGE<sub>2</sub> formation in this model was not established (12). However, immunohistochemical data indicated an increase in COX-2 expression in the outer medulla, suggesting that COX-2 expression in the mTAL may increase after challenge with Ca<sub>o</sub><sup>2+</sup>.

The total urinary Ca<sub>o</sub><sup>2+</sup> concentration has been estimated to be 2.3 ± 0.3 mM in a normal human ingesting a low calcium (10 mM) diet with unrestricted access to fluids (13). When a systemic Ca<sup>2+</sup> load is excreted, small alterations in peritubular Ca<sup>2+</sup> concentrations in the TAL attenuate its capacity for NaCl and Ca<sup>2+</sup> reabsorption (14). However, urinary Ca<sup>2+</sup> concentrations may increase substantially when water intake is restricted. Thus, during antidiuresis, AVP-induced osmotic water permeability in the terminal inner medullary collecting duct raises luminal Ca<sup>2+</sup> concentrations to greater than 5 mM (14). The renal concentrating defect associated with hypercalcemia is characterized by resistance to arginine vasopressin (AVP), and an important role of the AT1 receptor for Ang II was observed in experimentally-induced hypercalcemia, associated with an increased expression of renal COX-2 (12). Localization and functional coupling of AT1A receptors in the mTAL was demonstrated by RT-PCR and measuring changes in Ca<sub>i</sub><sup>2+</sup> by digital imaging microscopy (15). The Ang II-mediated increases in Ca<sub>i</sub><sup>2+</sup> were completely abolished by the AT1 receptor antagonist, losartan. As Ang II increases mTAL TNF production (6), which increases COX-2 expression in this nephron segment (2), changes in Ca<sub>i</sub><sup>2+</sup> may be part of a mechanism that promotes salt and water loss.

### *Ca<sub>o</sub><sup>2+</sup>-mediated mechanisms that affect ion and water homeostasis*

Ca<sub>o</sub><sup>2+</sup>-sensing receptor (CaR), originally cloned from bovine parathyroid (16), also has been found in several nephron segments including the rat mTAL

(17). CaR are expressed in tissues that are involved in  $\text{Ca}^{2+}$  homeostasis including the parathyroid (16), parafollicular cells of the thyroid (18), and the kidney (19). Heterozygous and homozygous CaR knockout mice exhibit mild and severe alterations in  $\text{Ca}^{2+}$  homeostasis, respectively, confirming the importance of CaR in  $\text{Ca}^{2+}$  homeostasis (20). Moreover, the physiological relevance of this receptor in humans has been demonstrated by identifying hyper- and hypocalcemic disorders resulting from CaR mutations: familial hypocalciuric hypercalcemia (FHH) and neonatal severe hyperparathyroidism result from inactivating CaR mutations, while an autosomal dominant form of hypocalcemia is caused by activating mutations (21). Interestingly, persons with FHH have alterations in their handling of water, suggesting that CaR also may contribute to the regulation of water metabolism.

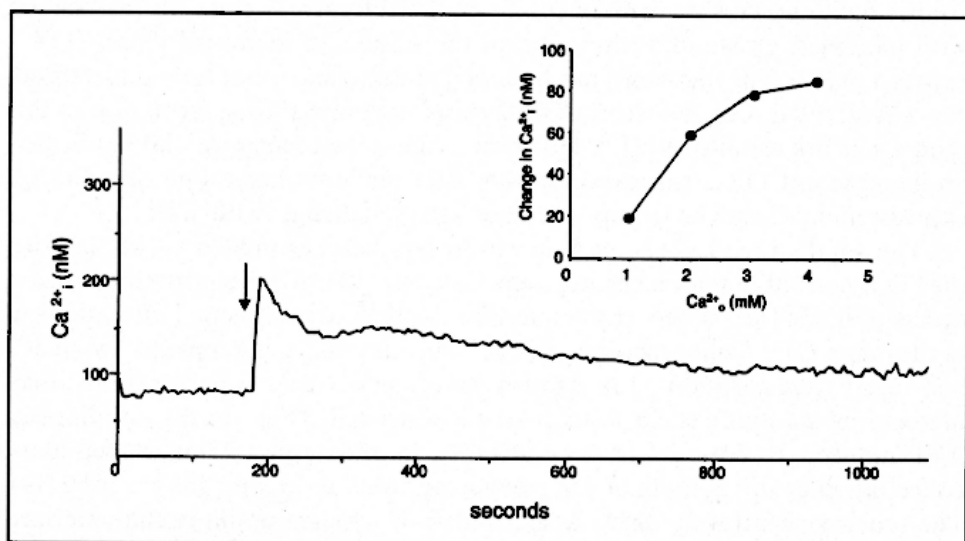


Fig. 2  $\text{Ca}_o^{2+}$  increases  $\text{Ca}_i^{2+}$  in mTAL cells. Cells grown on glass coverslips, were preincubated for 30 min in Hank's Balanced Salt Solution containing 1.23 mM  $\text{CaCl}_2$  then consecutively challenged with 1 mM  $\text{CaCl}_2$  (insert; representative figure,  $n = 2$ ) or 5 mM  $\text{CaCl}_2$ ; (representative figure,  $n = 3$ ).  $\text{Ca}_i^{2+}$  was determined using digital imaging microscopy in cells that were loaded with 2  $\mu\text{M}$  Fura-2 acetoxymethyl ester for 30 min at 37°C/5%  $\text{CO}_2$ .

CaR are G-protein-coupled receptors that transduce extracellular  $\text{Ca}^{2+}$  binding into several intracellular signals including stimulation of inositol triphosphate ( $\text{IP}_3$ ) production and release of  $\text{Ca}_i^{2+}$ . The release of  $\text{Ca}^{2+}$  from intracellular stores, in response to  $\text{Ca}_o^{2+}$ , implies that after stimulation of CaR,  $\text{IP}_3$  and diacylglycerol release subsequent to activation of phosphatidylinositol-phospholipase will increase PKC activity. The effects of  $\text{Ca}_o^{2+}$  on changes in  $\text{Ca}_i^{2+}$  levels in mTAL cells were determined using digital imaging microscopy. Successive additions of 1 mM  $\text{CaCl}_2$  increased  $\text{Ca}_i^{2+}$  in

a dose-dependent manner and showed no desensitization (Fig. 2), as previously shown in rabbit and mouse cTAL (22, 23). Basal levels of  $\text{Ca}_i^{2+}$  in cultured mTAL cells were approximately 100 nM and increased rapidly following additions of 5 mM  $\text{Ca}_o^{2+}$  (Fig. 2). The biphasic nature of the increase in  $\text{Ca}_i^{2+}$  may reflect an increase in  $\text{Ca}_i^{2+}$  by more than one mechanism. The rapid rise in  $\text{Ca}_i^{2+}$  is characteristic of release of  $\text{Ca}^{2+}$  from intracellular stores, while the sustained phase, in which the  $\text{Ca}_i^{2+}$  did not return to baseline levels as long as  $\text{Ca}_o^{2+}$  was present, may represent influx of  $\text{Ca}_o^{2+}$  via  $\text{Ca}^{2+}$  channels. We postulate that the rapid increase in  $\text{Ca}_i^{2+}$  after addition of  $\text{Ca}_o^{2+}$  is attributed to activation of the G-protein-coupled CaR, through which  $\text{IP}_3$  production and release of  $\text{Ca}^{2+}$  is stimulated.

#### *$\text{Ca}_o^{2+}$ and poly-L-arginine increase $\text{PGE}_2$ production by mTAL cells*

Calcium is a known activator of phospholipase  $\text{A}_2$  ( $\text{PLA}_2$ ), a major pathway by which AA is made available to various enzymes including COX, cytochrome P450 (CyP450), and lipoxygenases. Stimulation of CaR may, therefore, increase AA release for subsequent metabolism by COX-1 in mTAL cells. This mechanism is evident after short-term exposure (10 min) to  $\text{Ca}_i^{2+}$  or CaR agonists, as the effects of high (5 mM)  $\text{Ca}_i^{2+}$  and  $\text{Gd}^{3+}$  were abolished after inhibition of  $\text{PLA}_2$  activity (24) or addition of 17-octadecynoic acid, a specific inhibitor of CyP450 monooxygenase. However, recent work from our laboratory has demonstrated that different eicosanoid mediators regulate the short- and long-term effects of Ang II (6). Moreover, the coupling of CaR to intracellular signaling pathways may vary according to the magnitude of the stimulus and the parameter that is assessed (25). Thus, mTAL cells were challenged for various times with 1.7 or 2.0 mM  $\text{Ca}_o^{2+}$  and  $\text{PGE}_2$  levels were determined by ELISA. Control levels of  $\text{PGE}_2$  increased slightly over the time course studied, as did  $\text{Ca}_o^{2+}$ -stimulated  $\text{PGE}_2$  formation after a 1 hr challenge (Fig. 3A). In contrast, substantial increases in  $\text{PGE}_2$  production were observed when cells were challenged with either 1.7 or 2.0 mM  $\text{Ca}_o^{2+}$  for 3 or 9 hr (Fig. 3A). The magnitude of the responses at 1 hr vs. 3 and 9 hr may reflect differences in the mechanisms by which  $\text{PGE}_2$  was produced (i.e.; a COX-1-dependent mechanism at 1 hr and a COX-2-dependent mechanism at 3 and 9 hr). Control experiments in which extracellular  $\text{Cl}^-$  concentrations were increased, (by adding NaCl), without changing  $\text{Ca}_o^{2+}$  had no effect on  $\text{PGE}_2$  production (data not shown). Cells challenged for 9 hr with the CaR agonist, poly-L-arginine, also produced significantly more  $\text{PGE}_2$  than control cells (Fig. 3B). As PMA, a direct activator of PKC, also increases COX-2 expression in mTAL cells (2), it is possible that activation of PKC may be one mechanism by which COX-2 expression increases after challenge with  $\text{Ca}_o^{2+}$  and CaR agonists.

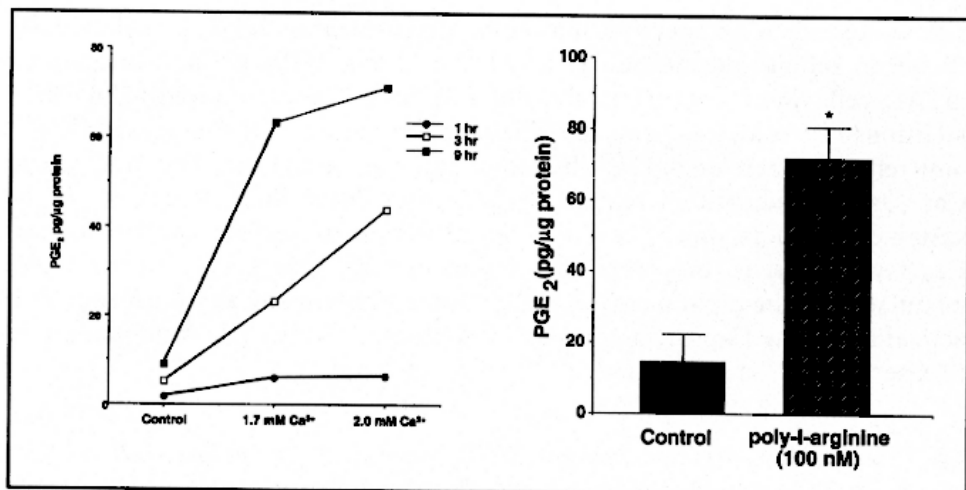


Fig. 3. Effects of  $\text{Ca}_0^{2+}$  and poly-L-arginine on  $\text{PGE}_2$  formation by mTAL cells. Primary cultures of mTAL cells were quiesced in RPMI 1640 containing 0.42 mM  $\text{CaCl}_2$  and 0.5% FCS for 18 hr then exposed for (A) various times to  $\text{Ca}_0^{2+}$ , (representative experiment,  $n = 2$ ) or (B) 9 hr to poly-L-arginine; (\*):  $p < 0.01$ ,  $n = 3$ .  $\text{PGE}_2$  was determined by ELISA. "Control" indicates that cells were incubated in media containing 0.42 mM  $\text{Ca}^{2+}$ ; this amount should be added to the amounts used to challenge the cells to obtain total  $\text{Ca}_0^{2+}$  present.

### $\text{Ca}_0^{2+}$ -mediated mechanisms that affect ion and water homeostasis

Stimulation of CaR by  $\text{Ca}_0^{2+}$  was recently shown to acutely increase production of 20-HETE, which inhibits activity of the apical 70 pS  $\text{K}^+$  channel in the mTAL (26). This channel is important for  $\text{K}^+$  recycling and maintaining activity of the apical  $\text{Na}^+-\text{K}^+-2\text{Cl}^-$  cotransporter, a major determinant of salt and water regulation by the mTAL (27).  $\text{PGE}_2$  also inhibits the  $\text{Na}^+-\text{K}^+-2\text{Cl}^-$  cotransporter, as well as the  $\text{Na}^+$  pump and apical  $\text{K}^+$  channel (28–30). However, the short-term effect of  $\text{Ca}_0^{2+}$  on the apical  $\text{K}^+$  channel activity occurred by a cyclooxygenase-independent mechanism. Previous work from our laboratory has shown that a temporal dependency determines the involvement of either cytochrome P450- or COX-derived AA metabolites on mTAL ion transport mechanisms. For instance, the short-term (15 min) effects of Ang II were mediated *via* a cytochrome P450-dependent mechanism, while long-term effects (>3–4 hr) were indomethacin-sensitive and involved a mechanism that also was dependent on the expression of TNF production (6) (Fig. 1). It is possible that regulation of ion transport mechanisms mediated by AA metabolites in the mTAL have different characteristics that are determined by the duration of the stimulus. Thus, while the short-term effects of  $\text{Ca}_0^{2+}$  are mediated by 20-HETE and appear to involve activation of the CaR, we propose that the ability of  $\text{Ca}_0^{2+}$  and CaR agonists to increase  $\text{PGE}_2$

production include a TNF-dependent mechanism that facilitates COX-2 expression.

Our previous finding that Ang II increases TNF production in the mTAL suggests that, under certain circumstances, this cytokine is critical to the activity of Ang II on ion transport function in this nephron segment (6). As these effects were attributed to an increase in COX-2 mRNA accumulation, protein expression and activity, induction of TNF may be essential to the regulation of COX-2 expression in the mTAL. Thus, other molecules that affect TNF production by these cells also may influence transport mechanisms *via* prostanoids derived from COX-2. The functional implications of such a mechanism are those predicted by increased levels of PGE<sub>2</sub>; i.e. inhibition of the Na<sup>+</sup> pump, cotransporter, and apical 70 pS K<sup>+</sup> channel are all mechanisms that promote salt and water excretion. We hypothesize that calcium, *via* stimulation of the CaR, may be a regulator of TNF production and COX-2 expression in the mTAL. Such an interaction may provide a mechanism by which eicosanoids derived from distinct biochemical pathways (CyP450 vs. COX) and subject to different regulatory influences might reinforce each other's actions. For instance, while calcium rapidly increases 20-HETE formation by the mTAL *via* a putative CaR-dependent mechanism, stimulation of PGE<sub>2</sub> synthesis *via* a COX-2-dependent mechanism would require considerably more time. The net effect of these temporally dissociated events would be the synthesis of two different eicosanoids, 20-HETE and PGE<sub>2</sub>, with similar effects on mTAL ion transport pathways. The role of COX-2 in this setting also may include an important mechanism for the conversion of 20-HETE to prostanoid-like compounds.

Recently, compounds termed "calcimimetics", which mimic or potentiate the effects of Ca<sup>2+</sup> at the level of the CaR have been discovered (31). These agents were designed as a means of providing pharmacological manipulation of plasma levels of parathyroid hormone (PTH), and the first-generation calcimimetic, NPS R-568, has exhibited promising results in clinical trials for primary hyperthyroidism and hyperparathyroidism secondary to chronic renal insufficiency (32). Our studies suggest that these molecules also may offer a novel means of affecting salt and water reabsorption. This mechanism may provide the rationale for the use of these molecules in conditions, such as hypertension and congestive heart failure, in which manipulation of extracellular fluid volume has a beneficial effect.

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Author's address: Dr. Nicholas R. Ferreri, Dept of Pharmacology, New York Medical College, Valhalla, USA.

E-mail: nick.ferreri@nymc.edu