

Flowering biology and pollen production of four species of the genus *Rosa* L.

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Abstract

Wild growing rose species are of great importance as a source of pollen for insects. Oil extracted from the petals of various *Rosa* species is used in perfumery, cosmetic industry, and therapeutics. In our study, we compared the flowering duration and flower lifespan, the number of stamens and pistils, the mass and size of pollen grains as well as the anatomical features of the petals of four *Rosa* species: *R. canina*, *R. ×damascena*, *R. gallica*, and *R. rugosa*. Moreover, we examined the pollen loads collected by bumblebees foraging on rose flowers in order to determine the attractiveness of pollen of this genus to insects. We showed the flower lifespan to vary (3.5–8 days) in the roses studied and revealed high variation in the number of stamens (82–260) and pistils (17–65) as well as in the mass of pollen produced. The flowers of *R. rugosa* produced the highest amount of pollen (26.7 mg per flower), while the flowers of *R. canina* the least (3.3 mg per flower), which is associated with differences in the number of stamens developed in the flowers between these species. The largest pollen grains were found in *R. ×damascena* and *R. gallica*. We demonstrated that *R. ×damascena* produces the thickest petals and that scent-emitting papillae found on the adaxial surface of the petals differ in size and shape in the rose species investigated.

Keywords: *Rosa*; flowers; functional morphology; petal structure; scent emission; pollen mass; pollen loads

Introduction

The genus *Rosa* includes about 200 species growing only in the northern hemisphere. According to some authors, 16 species of this genus are found in the wild in Poland [1], while some other ones mention 32 species [2]. Due to their androecium of numerous stamens, wild rose species provide food resources for insects as a rich source of pollen [3]. *Rosa canina* L. belongs to the most common species found in Poland, whereas *Rosa gallica* L. is a very rarely encountered species that is fully protected [2,4].

In the world, rose oil is currently obtained for industrial purposes from several rose species. *Rosa gallica* L., *R. ×damascena* Mill., and *R. rugosa* Thunb., inter alia, are mentioned among them [5]. The two latter species belong to introduced plants in Poland, while *R. rugosa* has the status of invasive plant in Europe [6–8]. Rose oil is used in perfumery, cosmetic industry, and therapeutics. It exhibits antiseptic and anti-inflammatory activity, and accelerates healing of wounds [5,9,10]. Rose oil is also used in aromatherapy, since it has a sedative and mentally stabilizing effect [11]. The scent of the flowers of individual rose species may significantly vary.

Rosa species with rose, fruity, spicy, musky and violet scent are mentioned [5].

Dobson et al. [12] showed the scent of the *R. rugosa* flower to be determined by qualitatively different oils emitted by the sepals, petals, stamens, and pollen. Bergougnoux et al. [13] found that in *Rosa ×hybrida* both surfaces of the petals emit odorous substances. Similar results were obtained for *R. rugosa* by Sulborska et al. [14]. The above-mentioned authors demonstrated the presence of essential oils both on the adaxial surface of the petals, covered with papillae, and on the abaxial surface on which flat epidermal cells were found.

The aim of our study was to evaluate the apicultural value of 4 *Rosa* species, including three species grown for oil. The floral morphology, including the number of stamens, and the mass of pollen produced by the stamens was compared. The petal structure was analyzed, with special attention to the micromorphology of papillae emitting odorous substances which are found on the adaxial surface of the petals, to show whether an intense emission of scent is associated with some special structural features of the epidermis. Microscopic analysis of pollen loads collected by bumblebees visiting the flowers of *R. rugosa* and *R. ×damascena* was performed in order to determine the share of *Rosa* pollen in them.

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Material and methods

The study was carried out in the Botanical Garden of the Maria Curie-Skłodowska University in Lublin in 2013. Observations included the flowers of 4 *Rosa* species: *R. canina* L., *R. gallica* L., *R. rugosa* Thunb., and *R. ×damascena* Mill.

Flowering biology

The phenology of flowering was investigated using the method by Łukasiewicz [15]. The time when 10% of flowers on a shrub were open was considered to be the beginning of flowering for each species, full flowering was when about 25% of flowers were open, while the end of flowering when 75% of flowers were found to be senescent (the end of the effect of mass flowering). The date of senescence of the last flowers was considered to be the end of flowering. Flower diameter was measured in 30 flowers. The number of petals, stamens and pistils was estimated in 10 flowers. The stamens were excised from the flowers in order to determine pollen production.

Petal anatomy

Sections from newly open flowers (0.5 × 0.5 cm from the middle part of petal) were fixed in 2% glutaraldehyde with 2.5% paraformaldehyde in 0.75 M phosphate buffer with a pH of 6.8 at a temperature of 4°C for 12 h. Next, the samples were dehydrated in an ethanol series, dried at the critical point in liquid CO₂ and coated with gold using an EMITECH K 550x sputter coater. The preparations were observed under a TESCAN/VEGA LMU scanning electron microscope at an accelerating voltage of 30 kV.

Examination of pollen

Pollen viability was estimated using acetocarmine slides. For each species, viable (staining pink) and non-viable (colorless) pollen grains were counted under a microscope in 4 successive longitudinal transects of the slides, up to a total of 100 grains in each transect.

The size of pollen grains was determined based on measurement of the polar (P) and equatorial (E) axes in semi-permanent slides with glycerol-gelatin mounting medium. The size was estimated using light microscopy with an eyepiece micrometer in 30 replicates for each species.

Pollen production

Pollen production was estimated using the ether method by Warakomska [16] with the alcohol modifications of Szklanowska [17]. The study material consisted of buds picked at stamen maturity before pollen release (excluding the

first anthers beginning to shed pollen). A hundred anthers were placed in each of previously tared glass microcylinders. The microcylinders with fresh anthers were inserted in the drier at a temperature of about 30°C. After 14 days the dried anthers were immersed twice in dimethyl ether. A 96% alcohol was then instilled into the microcylinders to thoroughly wash out the remnants of pollen from the anthers and subsequently the remains of the anthers were removed. After the alcohol evaporated, the pollen samples were placed in the laboratory oven, dried to constant mass, and then weighed. Four samples of 100 anthers were collected for each species. The mass of pollen obtained was calculated per 100 stamens and per 10 flowers.

Analysis of pollen loads

To determine the mass of pollen loads formed by bumblebees, five individuals were captured. Under field conditions, pollen packages were removed from the baskets on the tibiae of the third pair of legs and the insects were released to the environment. After several days the pairs of air-dried pollen loads were weighed and their color was determined using *A dictionary of color* by Maerz and Paul [18]. Preparations were then prepared following the recommendations of Smaragdova [19]. For this purpose, each pollen load was ground with 1 ml of distilled water with glycerol at a ratio of 1:1 until fine. Subsequently, permanent microscope slides were prepared. The pollen spectrum of each sample was examined under a Nikon Eclipse E600 light microscope at a magnification of 40×15. At least 300 grains were counted in each slide, according to the recommendations of Moar [20]. These grains were identified and assigned to species, genus, type of structure, or family. During the microscopic pollen analysis, reference slides and available keys were used [21–25]. The standard deviation (SD) was calculated in an Excel spreadsheet.

Results

Flowering biology

The rose species studied flowered at similar times over the period from May 21 to July 3 (Tab. 1). *Rosa rugosa* bloomed first (May 21), followed by *R. canina* (June 2), while the other two species entered the flowering period latest (June 8). The flowering of *R. canina* and *R. gallica* lasted slightly less than 3 weeks, in the case of *R. ×damascena* the flowering period was almost 4 weeks, whereas for *R. rugosa* it was as much as 6 weeks. Among the taxa investigated, the flowers of *R. canina* had the shortest lifespan (3.5 days), the flowers of *R. gallica* and *R. rugosa* were characterized by a medium

Tab. 1 Flowering phenology and flower lifespan of the four *Rosa* species.

Species	Flowering time	Flowering duration (days)	Flower lifespan (days)
<i>Rosa canina</i>	June 2–19	18	3.4
<i>Rosa ×damascena</i>	June 8 – July 3	26	8.0
<i>Rosa gallica</i>	June 8–26	19	4.5
<i>Rosa rugosa</i>	May 21 – July 2	43	5.0

lifespan (4.5–5 days), while the flowers of *R. ×damascena* bloomed longest (8 days; Tab. 1).

Floral morphology

The flower diameter of the studied species ranged 5.5–11.0 cm (Tab. 2). The *R. rugosa* was characterized by the largest flowers (Fig. 1a, Fig. 2a, Fig. 3a, Fig. 4a).

In the case of three species studied, the number of petals was fixed and it was 5; only in the hybrid *Rosa ×damascena* the number of petals ranged between 22 and 35, with a mean number of 28 (Tab. 2, Fig. 2a). The mean number of stamens was in the range of 83–260, whereas the number of pistils in the range of 17–65 (Tab. 2). The flowers of *R. rugosa* had the most stamens and pistils. The flowers of *R. canina* produced the least generative parts. Compared to the *R. rugosa* flowers, they produced 3 times fewer stamens and almost 4 times fewer pistils. The measurements and photographs included in this paper show that the flowers of the species studied differ not only in size, but also in shape and petal color (Fig. 1a, Fig. 2a, Fig. 3a, Fig. 4a,b). We also found differences in the type of scent emitted by the petals. The *R. ×damascena* flowers were characterized by the strongest scent with a rosy-fruity note. The flowers of *R. gallica* and *R. rugosa* emitted a medium strong scent, while the flowers of *R. canina* had the weakest scent.

Petal anatomy

The petal thickness in the investigated *Rosa* species ranged from 120 to 374 μm (Tab. 3). The thickest petals were found in the flowers of *R. ×damascena*, followed by *R. rugosa*. *R. canina* and *R. gallica* had a similar petal thickness which was at the same time the smallest.

The height of the papillae found on the adaxial surface of the petals and the width of the cuticular striae on their surface were compared (Tab. 3). The largest papillae (28 μm) were found on the petals of *R. gallica* (Fig. 3c,e,f). In the case of *R. ×damascena* (Fig. 2d,e) and *R. rugosa* (Fig. 4e–h), the papilla height reached similar values. The width of the cuticular striae on the papillae also significantly differed among the species (Tab. 3). The widest striae formed on the papillae of *R. ×damascena* (Fig. 2b–d), while the striae of *R. canina* had an almost twice smaller width (Fig. 1b–d).

It can be concluded based on the SEM analysis that the papillae of *Rosa* species differ not only in size but also in shape. In *R. rugosa* they narrow down towards the apex

most (Fig. 4c–f,h), whereas in *R. canina* and *R. ×damascena* they are the most rounded at the tip (Fig. 1b–d, Fig. 2b–d). The papillae on the petals of *R. gallica* (Fig. 3b–f) show an intermediate form between the two above-mentioned species. Moreover, in *R. canina* high variation is observed in the size of the papillae located next to one another on the petals (Fig. 1b–d). The massive structure of the striae in *R. ×damascena* can also be seen in the SEM images (Fig. 2b–d). A comparison of the petal cross sections for the four *Rosa* species reveals that the thickest petals, with the most dense arrangement of cells, are produced by *R. ×damascena* (Fig. 2e).

Characteristics of pollen grains

Rose pollen grains are tricolporate and round in shape, and have a striate exine (Fig. 5). In the species studied, the length of the polar axis in equatorial view was 29.4–33.5 μm , whereas that of the equatorial axis in polar view ranged 29.2–34.2 μm (Tab. 4). Among the species studied, pollen grains of *R. ×damascena* and *R. gallica* were largest. Generally, pollen grains of the investigated species can be considered to be medium. The P/E ratio reached a value close to 1.0, which indicates the round shape of pollen grains. The pollen viability, as estimated using the acetocarmine method, ranged from 40% (*R. canina*) to 95% (*R. rugosa*; Tab. 5).

Pollen production

The pollen mass determined for 100 stamens ranged between 4.0 mg (*R. canina*) and 10.3 mg (*R. rugosa*; Tab. 5). Significant differences were found when the pollen mass was calculated per flower, which resulted from the varying numbers of stamens in the flowers. Three times more pollen was estimated for *R. rugosa* flowers (267 mg per 10 flowers) compared to *R. ×damascena* flowers (89 mg per 10 flowers). Much higher (eightfold) differences were found between the pollen content in the flowers of *R. rugosa* and *R. canina*.

Pollen loads

Bumblebees showed interest in pollen from the flowers of *Rosa* species and formed pollen loads. The mass of a pair of sampled and dried pollen loads ranged from 9.7 mg to 50.5 mg (on average 30.3 mg). The smallest pollen loads were formed on the *R. rugosa* flowers, while the largest ones on the *R. ×damascena* flowers. In both cases, the pollen load was sampled from *Bombus sylvarum* L.

Tab. 2 Characteristics of the flowers of the four *Rosa* species.

Species	Flower diameter (cm)	Number of petals	Number of stamens per flower			Number of pistils per flower		
			range	mean	SD	range	mean	SD
<i>Rosa canina</i>	5.7	5	73–91	83	±7	9–24	17	±7
<i>Rosa ×damascena</i>	7.0	5	88–116	103	±12	41–47	43	±3
<i>Rosa gallica</i>	5.5	22–35 (28)	146–179	161	±11	37–44	41	±3
<i>Rosa rugosa</i>	11.0	5	240–279	260	±14	46–82	65	±13
Mean				151.8			41.5	

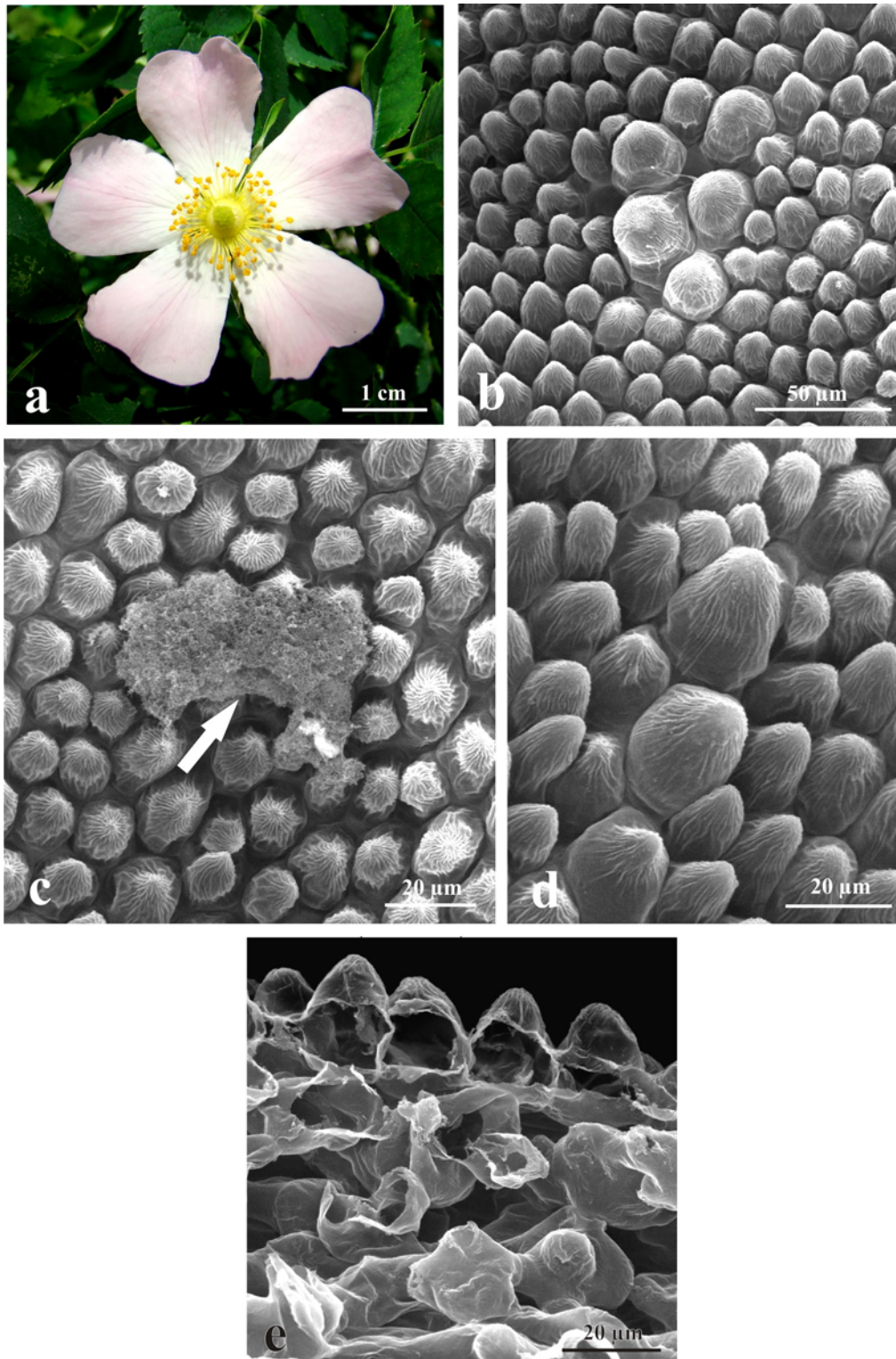


Fig. 1 Habit of the flower (a) and petals of *Rosa canina* (SEM). **b–d** Different sized papillae on the adaxial epidermis with a striated cuticle. Visible remnants of dried secretion (arrow; c). **e** Petal cross section showing a loose arrangement of mesophyll cells.

The color of the pollen loads sampled, as determined using *A dictionary of color* by Maerz and Paul [18], ranged between yellow amber and mustard green (T10/H-4 – T11/J-3).

The microscopic analysis of the samples of pollen loads revealed the presence of pollen of 16 taxa, including 10

belonging to nectariferous plants and 6 non-nectariferous plants (entomophilous or anemophilous). From 5 to 11 types of pollen were found in the individual samples. *Rosa* pollen grains were recorded in each pollen load sample and their participation was dominant, ranging from 88.61% to

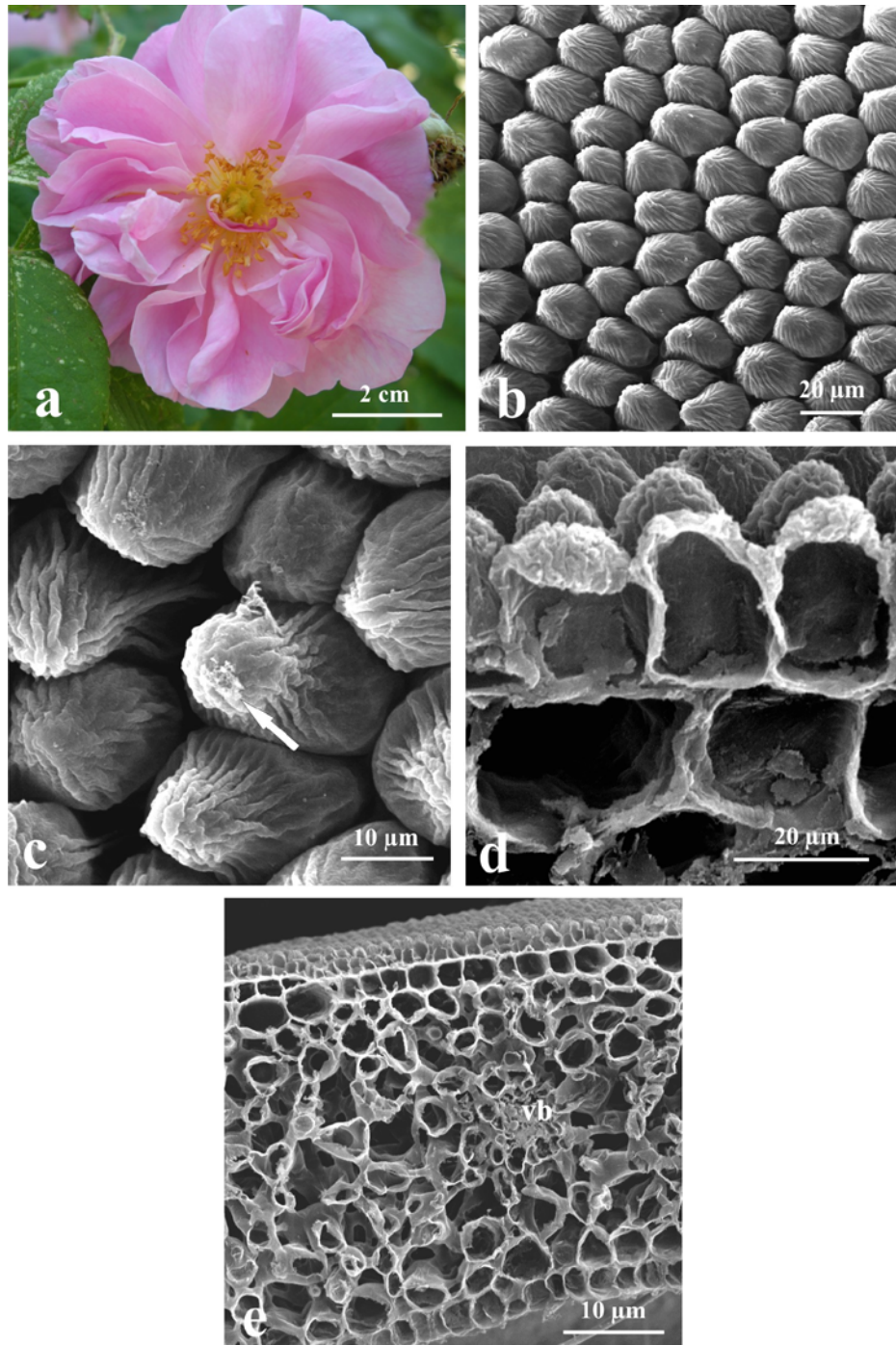


Fig. 2 Habit of the flower (a) and petal sections of *Rosa* \times *damascena* (SEM). **b,c** Papillae from the adaxial surface of the petals with massive cuticular striae on the outer wall surface and remnants of dried secretion (arrow). **d** Partial cross section of outer tissues of the petal. **e** Petal cross section with a visible vascular bundle (vb).

97.94% (Fig. 6, Fig. 7). The percentage of pollen belonging to the other taxa was low and corresponded to the groups of single pollen (3–16%) or sporadic pollen (below 3%).

Discussion

In the flowers of the genus *Rosa*, the perianth is situated at the apex of a green hypanthium (cup-shaped) that

surrounds the gynoecium. The hypanthium in *Rosa* has to be interpreted as “a fusion product of leaf structures” on the basis of the vascular bundle system [26]. Among the *Rosa* species investigated, three species have 5 petals (*R. canina*, *R. gallica*, and *R. rugosa*), whereas *R. x damascena* produces many more petals (28) in several whorls. In addition to the petals of the corolla in this species, numerous petaloids can be found, which are intermediate forms of the stamens. The number of stamens in the flowers of the *Rosa* species

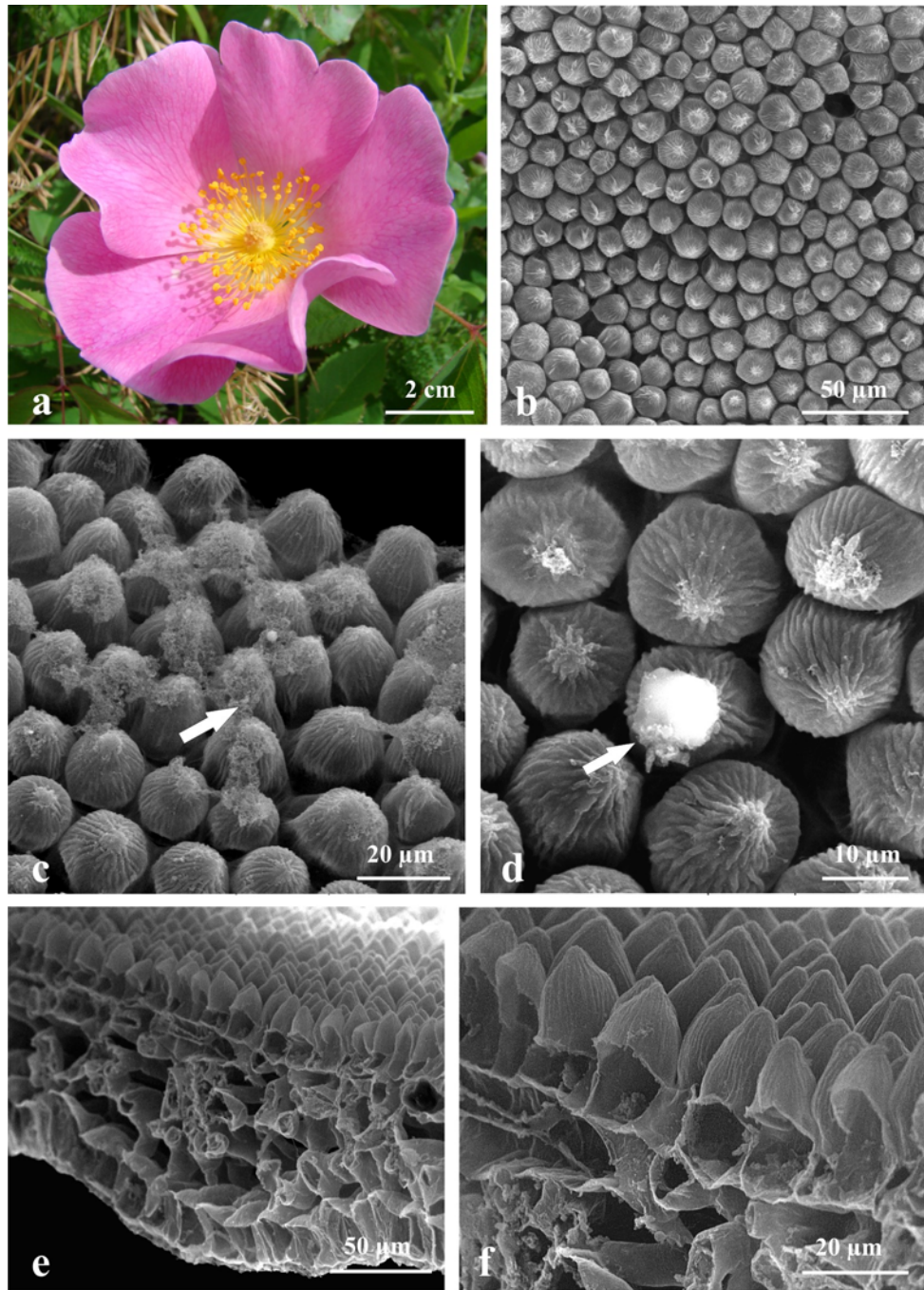


Fig. 3 Habit of the flower (a) and petal sections of *Rosa gallica* (SEM). **b–d** Papillae from the adaxial surface of the petals with a striated cuticle and remnants of dried secretion (arrows). **e, f** Petal cross sections.

investigated greatly varied, ranging 83–260 stamens. In terms of the increasing number of stamens, the species in question can be ranked as follows: *R. canina*, *R. ×damascena*, *R. gallica*, *R. rugosa*.

Flower scents are most frequently secreted diffusely by the corolla epidermis. However, scent can also be produced by other parts of the flower. In *R. rugosa* and *R. canina*, the scent emitted by the flowers is dominated by terpenoid and benzenoid alcohols produced by the petals. Other parts of the flowers of these species produce scents with a different chemical composition. Sesquiterpenes have a major

contribution to the scent of the sepals, while in the case of the anthers and pollen grains it has been found that there is a diversity of compounds, largely similar to those produced by the corolla [27,28].

Among the *Rosa* species investigated in our study, *R. ×damascena* was characterized by the most intense floral scent. It was classified as a rosy-fruity scent. Góra and Lis [5] report that among various floral scents of roses grown for essential oil, the scent of *R. ×damascena*, which is described as a beautiful floral scent with a warm tea/honey undertone, is most appreciated. The floral scent of *R. ×damascena* is

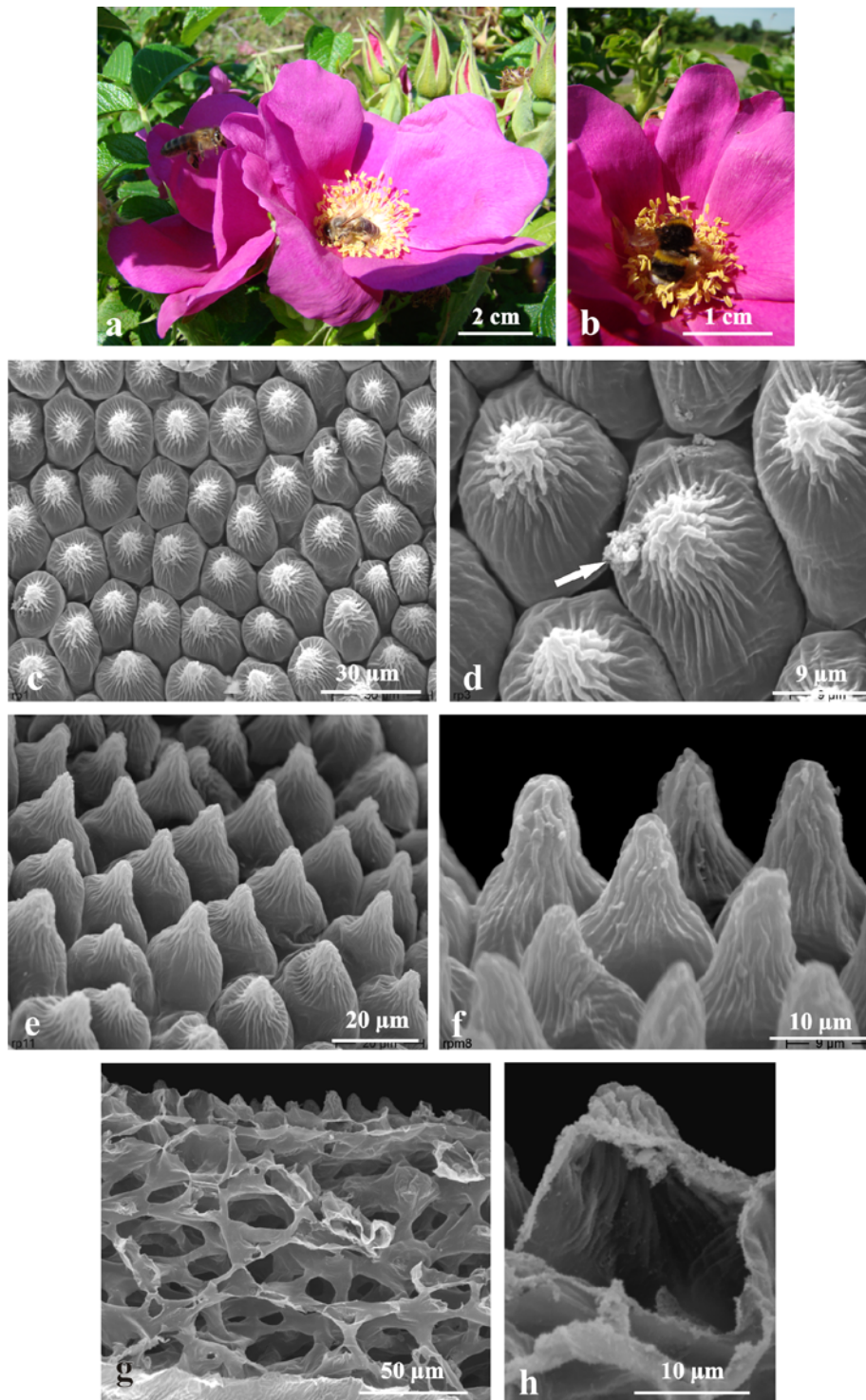


Fig. 4 Habit of the flowers visited by *Apis mellifera* and *Bombus* sp. (a,b) and petal sections of *Rosa rugosa* (SEM). c–f Papillae narrowing at the apex on the adaxial epidermis of the petals with visible striae and secretion remnants (arrow). g Petal cross section. h Enlargement of a longitudinal section of a papilla.

considered to be a standard for rose scent. According to these authors, in the petals of *R. damascena* var. *kazanlik* (Bulgaria) the average essential oil content is 0.035%, which is one of the highest values found for the genus *Rosa*. In the oil obtained from *R. ×damascena* flowers, the following

monoterpenes predominate: citronellol and geraniol [5]. The above data concerning the type of scent of *Rosa* flowers correspond to the findings of Proctor et al. [28] who claim that the flowers that are more specially adapted to bees have a sweet or honey-like scent. The scents of *R. gallica* and

Tab. 3 Anatomical features of the petals of the four *Rosa* species.

Species	Petal thickness (μm)			Papilla height (μm)			Width of cuticular striae on papillae (μm)		
	range	mean	SD	range	mean	SD	range	mean	SD
<i>Rosa canina</i>	101.0–136.9	119.9	± 18.0	21.3–25.3	23.4	± 1.7	0.43–0.50	0.46	± 0.03
<i>Rosa \times damascena</i>	364.3–384.4	374.4	± 14.2	22.4–29.5	25.8	± 2.4	0.67–1.14	0.94	± 0.21
<i>Rosa gallica</i>	116.7–126.7	121.7	± 7.1	25.8–29.7	27.9	± 1.5	0.66–0.91	0.78	± 0.11
<i>Rosa rugosa</i>	106.5–160.0	140.0	± 24.6	22.2–28.7	25.0	± 2.1	0.60–1.01	0.73	± 0.13

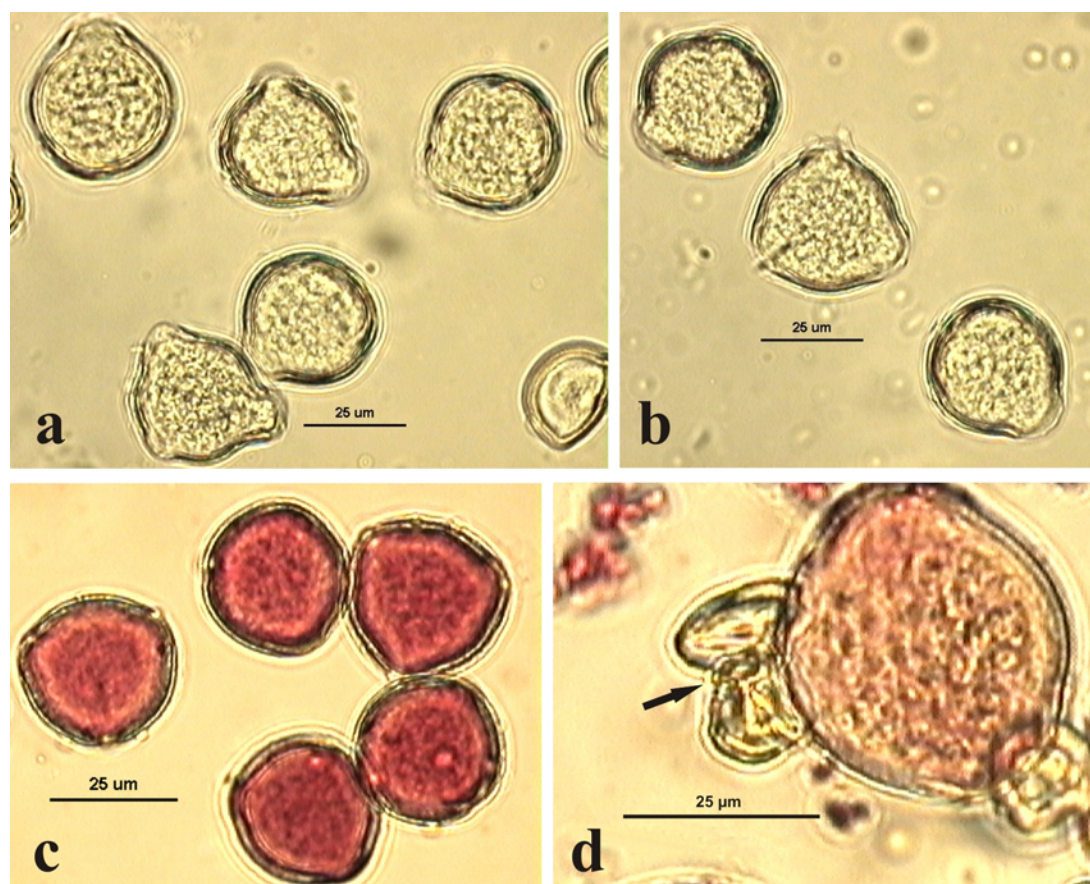


Fig. 5 Pollen grains. **a,b** *Rosa* sp. pollen grains in microscopic slides of different pollen loads sampled from *Bombus sylvarum* L. **c** Viable pollen grains of *R. rugosa* with a pink-stained protoplast after treating with acetocarmine. **d** Viable pollen grains of *R. canina* with a protoplast stained pink with acetocarmine as well as unstained and deformed non-viable pollen grains (arrow).

R. rugosa were determined in our research to be medium intense and this is also reflected in the data on the essential oil content in *R. gallica* petals (0.017%) [5], which is much lower than in the case of *R. \times damascena*. The *R. canina* flowers were characterized by the least intense scent. It is worth noting that this species is not mentioned among the roses used for the production of rose oil [5].

In the case of the *R. \times damascena* flowers, the large number of petals and their considerable thickness as well as their intense scent confirm their usefulness as oil material. It has been shown that the highest content of rose oil, located

mainly on the petal surface, is found in roses at the bud stage in early morning hours [5].

The present study found that each of the four *Rosa* species produces papillae of different shape and size. It is recognized that the role of the papillae produced in the petal epidermis is adaptation to entomophily as a foothold or as a light trap and petal reflexing [28,29].

The length of the polar axis of pollen grains in the rose species studied was in the range of 23–43 μm , which proved to be wider compared to that given by Kirk [30] for various rose varieties (24–36 μm). A wider size range was also

Tab. 4 Dimensions of pollen grains of the four *Rosa* species.

Species	Length of polar axis (P; μm)			Length of equatorial axis (E; μm)			P/E ratio
	range	mean	SD	range	mean	SD	
<i>Rosa canina</i>	23.2–33.5	29.4	± 3.3	28.4–33.5	29.7	± 1.9	0.99
<i>Rosa \times damascena</i>	31.0–43.2	33.5	± 1.2	31.0–38.7	34.2	± 2.2	0.98
<i>Rosa gallica</i>	25.8–31.0	33.5	± 2.2	25.8–33.5	33.5	± 3.1	1.00
<i>Rosa rugosa</i>	25.8–31.0	30.2	± 1.7	28.4–31.0	29.2	± 1.3	1.00
Mean		31.7			31.6		0.99

Tab. 5 Pollen value of the four *Rosa* species.

Species	Pollen mass per 100 stamens (mg)			Pollen mass per 10 flowers (mg)			Viability of pollen grains (%)		
	range	mean	SD	range	mean	SD	range	mean	SD
<i>Rosa canina</i>	3.30–4.63	4.00	± 0.53	27.39–38.18	33.20	± 4.45	26.7–48.8	40.0	± 9.6
<i>Rosa \times damascena</i>	7.89–9.29	8.65	± 0.58	81.27–95.69	89.12	± 6.01	56.9–61.3	59.3	± 1.7
<i>Rosa gallica</i>	4.92–8.88	7.36	± 0.62	79.22–127.19	131.54	± 9.99	34.1–51.1	42.0	± 7.1
<i>Rosa rugosa</i>	9.17–12.09	10.27	± 1.33	238.42–313.04	267.02	± 34.64	91.6–96.8	94.8	± 2.6
Mean		7.57			130.21			59.0	

obtained in the case of *R. canina* pollen grains (23.2–33.5 μm) compared to the data reported by Ricciardelli d'Albore [24] (31.4–32.6 μm), but it was smaller than that given by Beug [31] (30.5–40.5 μm). In our study, the dimensions of the polar axis in *R. gallica* were in the range of 25.8–31.0 μm , whereas Beug [31] found a range of 28.1–38.6 μm for the pollen of this species.

In this research, the shape of pollen grains was determined to be round or close to round, as indicated by the P/E ratio ranging from 0.99 to 1.00. The ratio of 0.99 obtained for *R. canina* was slightly higher compared to the value of 0.96 that is reported for this species by Ricciardelli d'Albore [24].

In terms of apicultural usefulness, the flowers of wild roses are treated as polleniferous flowers, since they do not produce nectar [3,32]. Dobson et al. [33] emphasise that in the case of *R. rugosa* the main stimulus that attracts bumblebees is the pollen odor and the visual attractiveness of the anthers, whereas the petal color and scent play a secondary role. Cultivated roses are not important as a source of food for insects due to numerous petals and a low number of stamens. Some authors report that the rose flowers produce small amounts of nectar [3,30,34].

A large number of stamens in the flower frequently determines an abundant mass of pollen [28,35]. We found that the *R. rugosa* flowers, which had the highest number of stamens (260) among the species investigated, produced the highest mass of pollen (26.7 mg per flower). Very diverse results concerning the abundance of pollen in the latter

mentioned species can be found in the literature. Jabłoński [36] reports that *R. rugosa* produces 19.1 mg of pollen per flower, whereas Lipiński [3] mentions a pollen mass of 45.9 mg per flower. The significant divergence in the results obtained by individual authors may arise from different soil nutrient availability and from the impact of weather conditions on the plants before and during flowering [36,37]. We found the lowest pollen mass in the flowers of *R. canina* (3.3 mg per flower). Rawski [34] also concluded that the flowers of *R. canina* are poor in pollen, giving them the lowest value on a three-level scale. A much lower mass of pollen was shown for *R. multiflora* (2.0 mg per flower) whose flowers had a similar number of stamens as the flowers of *R. canina*, but a much smaller corolla diameter [37].

The presence of *Rosa* pollen grains in samples of insect pollen loads [38,39] and in honey samples [40,41] is a confirmation of the attractiveness of pollen rewards to insects. The pollen loads formed by bumblebees which were sampled in this study had different shades of color, from yellow amber to mustard color. According to Hodges [21] and Kirk [30], bee loads of *Rosa* pollen are dark orange or even brownish. Ricciardelli d'Albore [24] reports that insects forage on *Rose* form slightly pink loads. Lipiński [3] finds that pollen loads collected from the same plant species can have a different color, depending whether the bee gathered pollen from open anthers or extracted it by gnawing through the anther walls. Their color can also be affected by the color of honey used by bees to moisten the pollen [42].

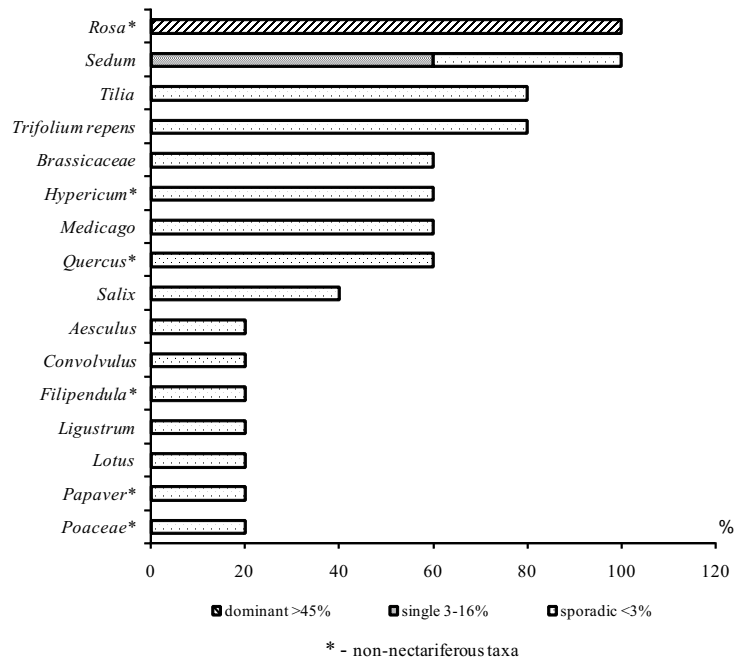


Fig. 6 Pollen frequency and percentages in pollen loads sampled from *Bombus sylvarum* L.

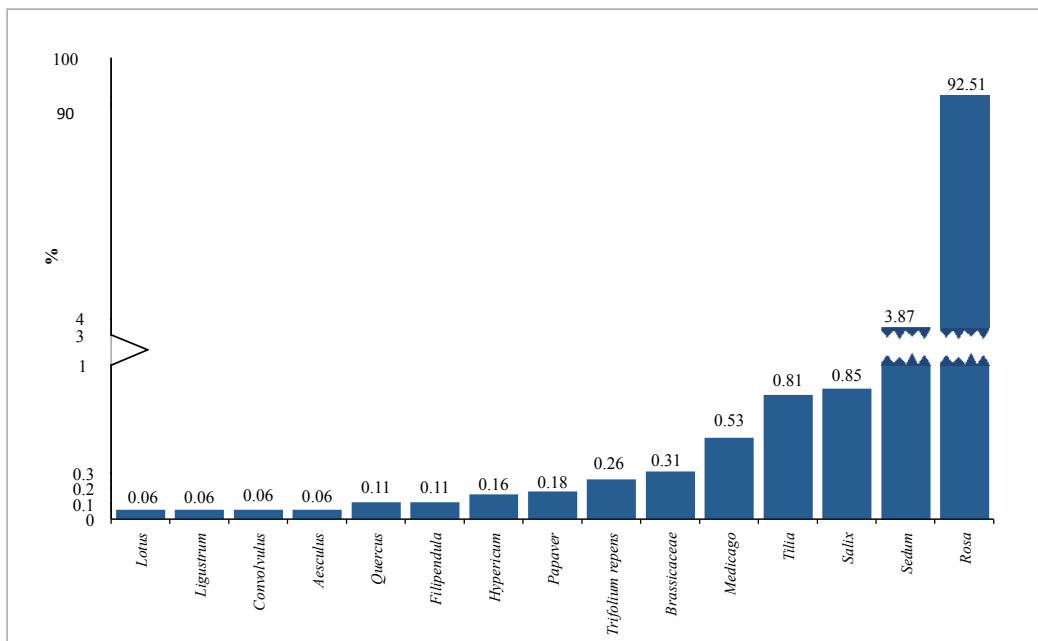


Fig. 7 Percentage participation of pollen of the individual taxa in pollen loads from *Bombus sylvarum* L.

Conclusions

1. Due to their largest biomass (the number and thickness of petals), the *R. ×damascena* flowers with an intense scent can be abundant raw material for the production of rose oil.
2. The scent-emitting papillae that are found on the ad-axial surface of the petals differ in size and shape in the species studied.
3. Among the *Rosa* species investigated, *R. rugosa* is characterized by the highest apicultural value due to the production of the highest mass of pollen, the longest flowering period as well as the greatest flower diameter.
4. *Rosa* pollen predominated (over 90%) in the pollen loads of bumblebees visiting two of the *Rosa* species investigated, which may be evidence of its attractiveness to insects.

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Authors' contributions

The following declarations about authors' contributions to the research have been made: idea of the study: EWC; microscopical analysis: AS, BŻ, ES, EWC; photographs: AS, ES; writing of the manuscript: AS, BŻ, ES, EWC.

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Biologia kwitnienia i produkcja pyłku czterech gatunków z rodzaju *Rosa* L.

Streszczenie

Dziko rosnące gatunki z rodzaju róża (*Rosa*) mają duże znaczenie jako źródło pyłku dla owadów. Uzyskiwany z płatków olejek znalazł zastosowanie w perfumerii, przemyśle kosmetycznym i lecznictwie. W pracy porównano długość okresów kwitnienia, długość życia kwiatów, liczbę pręcików i słupków, masę i wielkość ziaren pyłku oraz cechy anatomiczne płatków czterech gatunków z rodzaju *Rosa*: *R. canina*, *R. ×damascena*,

R. gallica i *R. rugosa*. Wykazano różną długość życia kwiatów badanych róż (3.5–8 dni) oraz duże zróżnicowanie w liczbie pręcików (82–260) i słupków (17–65), a także w masie produkowanego pyłku. Najwięcej pyłku wytwarzają kwiaty *R. rugosa* (26.7 mg/kwiat), najmniej kwiaty *R. canina* (3.3 mg/kwiat), co jest związane w dużej mierze z różną pomiędzy gatunkami liczbą pręcików w kwiatach. Największe ziarna pyłku stwierdzono u *R. ×damascena* i *R. gallica*. Wśród badanych gatunków róż najgrubsze płatki wytwarza *R. ×damascena*, a występujące na doosiowej powierzchni płatki papille emitujące zapach różnią się wielkością i kształtem. Udział pyłku *Rosa* w obnóżach pyłkowych zebranych przez trzmiele oblatujące kwiaty róż jest dominujący i wynosi >90%.