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Seedling growth, root development and nutrient use efficiency of Cypress clones in response to calcium fertilizer

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Abstract: Cypress (Cupressus funebris Endl.) is an important tree species in the subtropical regions of China; it is also a major tree species for afforestation and forest land restoration under low-fertility soil conditions. Cypress is considered a calcicolous tree, and its growth and development can be promoted significantly by exchangeable calcium (Ca²⁺) in the soil. However, most of the subtropical regions have low-fertility acidic soils, in which Ca²⁺ gradually becomes a limiting element for Cypress growth. In this study, different concentrations of Ca^{2+} fertilizer were added under fertile soil (3 g kg⁻¹ NPK fertilizer added) and low-fertility soil (0 g kg⁻¹ NPK fertilizer added) conditions. Cypress clones responded differently to Ca²⁺ addition in different soil conditions. The seedling height and dry matter quality of Cypress in fertile soil were significantly greater than those in low-fertility soil, but plant height and dry biomass did not differ significantly among Ca²⁺ treatments. The accumulation efficiencies of nitrogen (N), phosphorous (P) and Ca all differed significantly among the Ca²⁺ treatments. In low-fertility soil, the addition of 3 g kg-1 Ca²⁺ significantly promoted development of roots 0.5-2 mm in diameter, and both the C1 and C2 clones achieved their highest N, P and Ca accumulation efficiencies. When the Ca^{2+} concentration increased to 6 g kg⁻¹, the seedling height, dry matter quality and root development were lower than those of the 3 g kg⁻¹ Ca²⁺ treatment. In the fertile soil, the addition of Ca²⁺ significantly inhibited development of roots 0.5-1 mm in diameter. The highest N accumulation efficiency was achieved under the 0 g kg^{-1} Ca²⁺ treatment, and the highest Ca accumulation efficiency was achieved under the 6 g kg⁻¹ Ca²⁺ treatment. Seedling height, root dry weight, roots 0–1.5 mm in diameter and Ca accumulation showed a significant interaction effect between NPK fertilizer and Ca²⁺. Therefore, Ca accumulation was more efficient in low-fertility soils. Under low-fertility soil conditions, the addition of $CaSO_4$ can promote the root development of seedlings and advance and prolong the fast growth period of seedling height. Cypress clones can be used as an important tree species for afforestation under low-fertility soil conditions, especially under calcareous soil conditions.

Keywords: Cupressus funebris, root development, nutrient accumulation efficiency, calcium response, fertile soil, low fertility soil

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Introduction

Calcium (Ca^{2+}) is an essential nutrient required for plant growth and development (Kudla et al., 2010; Rashid et al., 2020). Plants generally have a Ca^{2+} content ranging from 0.1–5.0%, and this element plays a role in maintaining the stability of plant cell walls, cell membranes and membrane-bound proteins, modulating inorganic ion transport and regulating various enzyme activities (Kinzel, 1989; White & Broadley, 2003; Hepler, 2005; Abbasi & Manzoor, 2013). Co-application of Ca^{2+} and other elements, such as nitrogen (N), phosphorous (P) and potassium (K), can modulate intracellular Ca^{2+} levels, promote seedling growth and root development and enhance plant stress resistance (Jammes et al., 2011; Liu et al., 2018). Applying an appropriate amount of Ca²⁺ fertilizer to forest trees, such as Chinese fir [Cunninghamia lanceolata (Lamb.) Hook.], can facilitate root development and increase N, P and K uptake by seedlings (Rashid et al., 2020). Despite the various benefits of Ca²⁺ for plant growth, high concentrations of Ca²⁺ often have an inhibitory effect on plant growth and development (Chan et al., 2008).

The root is an important organ of plants for resource acquisition, and the spatiotemporal distribution of plant roots determines the amounts of water and nutrients that are absorbed for photosynthesis and harvest products (Hodge, 2004; Mommer et al., 2012; Rogers & Benfey, 2015). In different growth conditions, functional attributes such as root number and morphology have differential responses to changes in underground resources. As the most dynamic and physiological component of the underground part of plants, fine roots have gradually become the focus of plant growth and foraging strategies. Usually, fine roots are defined as all roots ≤ 2 mm in diameter, and these roots are important organs that allow trees to absorb water and nutrients (Meinen et al., 2009). Due to the continuous formation, senescence, death and regrowth of fine roots, when the soil environment is altered, the length growth rate of fine roots becomes faster or slower, and the mean root diameter becomes thicker or thinner in a short amount of time; the duration of such changes varies (Strand et al., 2008; Kong et al., 2014). According to a functional classification system, roots ≤ 2 mm in diameter are classified as absorptive and transport fine roots. Absorptive fine roots represent the most distal roots, which are involved primarily in the acquisition and uptake of soil resources, whereas transport fine roots are more common in the branching hierarchy and serve primarily structural and transport functions with some additional capacity for storage. It is assumed that roots \leq 1.0 mm in diameter are absorptive roots and roots 1.0-2.0 mm in diameter are transport roots. Considerable differences in biomass exist between these two root types among different tree species (McCormack et al., 2015). Resource uptake and transport also vary consistently among fine root orders. In particular, uptake capacity declines with increasing root order, while transport capacity increases (Mc-Cormack et al., 2017). At present, the relationship between root functional traits and nutrient uptake of forest trees is not very clear, and this relationship directly determines the foraging strategies of different tree species and affects the ability of different genotypes of the same tree species to obtain nutrients under significantly different site conditions (Chen et al., 2016). Recent studies suggest that the diameter of absorptive roots, which affects the carbon costs of constructing root length, also influences root foraging strategies for mineral nutrients (Eissenstat et al., 2015; Liu et al., 2015).

Plants growing in acidic or low base-saturation soils are prone to Ca^{2+} deficiency (Goulding, 2016). Ca²⁺ deficiency has gradually emerged as a limiting factor that influences the growth of forest trees. In the subtropical regions of China, fast-growing plantations are primarily distributed in large areas of hills and mountains, and these areas are mostly covered by low-fertility acidic red soils. Due to heavy rainfall and strong weathering in southern China, soil acidification is serious. Acid deposition areas are continuously expanding along with a serious loss of base ions, especially Ca2+ in the soil. This condition results in a deficiency of soil nutrients and a decline in soil fertility, which in turn severely restricts the growth and development of forest trees and the productivity of forest stands (Azevedo et al., 2013).

Cypress (Cupressus funebris Endl.) is an important tree species in the subtropical regions of China, mainly occurring in the Yangtze River Basin and to the south. Cypress is highly adaptive and can grow in a variety of site conditions, especially on limestone mountains (including acidic or weakly alkaline soils); thus, it is also a major tree species for afforestation and forest land restoration under low-fertility soil conditions. Cypress is a calcicole, and its growth and development can be promoted significantly by exchangeable Ca2+ in the soil. However, most subtropical regions have low-fertility acidic soils, in which Ca²⁺ gradually becomes a limiting element for Cypress growth. Previous research has found substantial variation in the root length of Cypress when the soil environment is disturbed; this effect is long lasting and, in particular, has a great effect on the number, morphology and function of fine roots (Brunner et al., 2019). However, there are diverse types of soil with a wide variety of physicochemical properties in areas where Cypress is distributed. Soil types and physicochemical properties have

important effects on the growth, distribution and activity of plant roots. Soil fertility determines the ability to supply nutrients to plants. Soil types and physicochemical properties, soil particle composition and total porosity differ. Through the continuous movement of air, water and nutrients in the soil, the nutrient content changes, which further affects the growth and development of roots. For example, clay has high water content and poor soil permeability, which will lead to a shorter root system and decreased root dry weight. However, various plants have different needs for soil types and often show differing comprehensive reflections of soil physicochemical properties (Katerji & Mastrorilli, 2009). Few data have been reported on root development and nutrient use of Cypress in response to Ca2+ across different soil environments. The effect of Ca²⁺ availability on diversity of root traits and plant growth has been poorly studied. Research is therefore needed to identify the optimal Ca²⁺ availability/ accessibility for the seedling growth and root development of Cypress, as the results play major indicative roles in improving seedling quality and forest tree productivity (Tracy et al., 2019; Paez-Garcia et al., 2015).

Materials and methods

Experimental site and materials

The experiment was conducted in a greenhouse of Laoshan Forestry Farm in Zhejiang Province, China. The semi-lignified branches from 4-year-old Cypress seedlings were collected as the propagation materials from elite individual plants of clone C1 (fast height growth) and clone C2 (slow height growth) in the full-sib progeny. A 1:1 ratio of riversand to loess was used as the cutting medium, and shading measures were performed at the same time. The cutting seedbed temperature was 22 °C, and the air humidity was 90%. The seedlings were selected based on their plant height (5.15 \pm 0.05 cm) and ground diameter (0.17 \pm 0.01 cm) after surviving cuttage. These seedlings were planted in containers with a height of 30 cm and diameter of 20 cm. The potting soil was an acidic red soil collected from forestland, and the soil layer was 0-20 cm thick. The physicochemical properties of the soil are provided in Table 1. The controlled-release fertilizer used in the experiment was a nursery fertilizer (APEX, USA).

Experimental design

NPK fertilizer was added at 3 and 0 g per kg of soil to simulate fertile and low-fertility soils, respectively. For Ca^{2+} fertilizer, $CaSO_4$ was added at 0, 3 and 6 g per kg of soil. Both NPK fertilizer and CaSO were mixed with their respective soil, stirred uniformly and placed into the containers. The experiment involved six treatments. Treatment 1 (T1): 3 g kg⁻¹ NPK fertilizer + 0 g kg⁻¹ CaSO₄; treatment 2 (T2): 3 g kg⁻¹ NPK fertilizer + 3 g kg⁻¹ CaSO₄; treatment 3 (T3): 3 g kg⁻¹ NPK fertilizer + 6 g kg⁻¹ $CaSO_4$; treatment 4 (T4): 0 g kg⁻¹ NPK fertilizer + 0 $g kg^{-1} CaSO_4$; treatment 5 (T5): 0 $g kg^{-1} NPK$ fertilizer + 3 g kg⁻¹ CaSO₄; and treatment 6 (T6): 0 g kg⁻¹ NPK fertilizer + 6 g kg⁻¹ CaSO₄. The experiment used a completely randomized block design. During seedling cultivation, the plants were watered every 10 days, and weeding, shading (75%) and pest control were performed. Twenty cutting seedlings were planted per treatment per clone, with three replicates each; therefore, 720 potted seedlings were planted.

Cultivation, harvest and analysis

The experiment was started on April 2, 2018, and plant height and ground diameter were measured for all plants. Thereafter, plant height was measured for each plant once every 20 days. Measurements were completed in November 2018, and continuous data for plant height were used to analyse the plant height growth pattern of Cypress. Seedlings were harvested on November 23. Whole plants were collected and divided into roots, stems and leaves, with each organ harvested separately. First, the roots were separated from the soil, washed with deionized water and stored. Root diameter was classified as follows: class D1 (root diameter range: 0–0.5 mm), class D2 (0.5– 1.0 mm), class D3 (1.0–1.5 mm), class D4 (1.5–2.0 mm) and class D5 (> 2.0 mm) (Liu et al., 2018). The root length, surface area and root volume of each diameter class were measured using the image analysis software WinRHIZO Pro STD1600+ (Regent Instruments, Canada). Next, the roots, stems and leaves were deactivated in an oven at 105°C for 30 min and then dried at 80°C until a constant weight was achieved in order to obtain the dry biomass of each part. The N content of each organ was measured using a FOSS (Foss Sossanalytizal a-s., Ahlleroed, Denmark) nitrogen analyser (Anderson & Ingram, 1993). The P content was measured by molybdenum

Table 1. Physical and chemical properties of potted soil

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Nutrient	Total N	Total P	Hydrolytic N	Available K	Available P	Organic matter	Exchange Ca	Exchange Mg	pII valua
elements	(g•kg ⁻¹)	(g•kg ⁻¹)	(mg•kg ⁻¹)	(mg•kg ⁻¹)	(mg•kg ⁻¹)	(g•kg ⁻¹)	(mg•kg ⁻¹)	(mg•kg ⁻¹)	pii value
Average	0.75 ± 0.09	0.32 ± 0.05	535+470	185 ± 112	0.99 ± 0.14	15 8+1 89	128 ± 125	9 24 + 0 85	4.65 ± 0.21
content	0.73±0.09	0.52 ± 0.05	55.5-4.70	10.3 - 1.12	0.99±0.14	13.0 - 1.09	120-12.3	9.24±0.03	4.03±0.21

antimony anti-colorimetry (He et al., 2013). The Ca content was measured by atomic absorption spectrophotometry (Rashid et al., 2020). The N, P and Ca contents were multiplied by the dry biomass of the whole plant to obtain the N, P and Ca accumulation. N accumulation efficiency = dry biomass accumulation of the whole plant/N uptake of the whole plant (g mg⁻¹); P and Ca accumulation efficiencies were calculated following the same method as that used for N accumulation efficiency.

Data analysis

Logistic regression was used to fit the seedling height growth pattern of Cypress under different Ca²⁺ treatments; the fitting equation was $y = k/(1+a e^{-bt})$, where *y* is cumulative growth of seedling height, t is the growth time, *k* is the theoretical upper limit of height growth, and *a* and *b* are the undetermined coefficients. One-way analysis of variance (ANOVA) was used to assess significant differences in seedling growth, root morphological characteristics and nutrient accumulation efficiency under Ca²⁺ treatments in fertile and low-fertility soils. All statistical analyses were performed using IBM SPSS Statistics 22.0 (IBM Corp., Armonk, NY, USA).

Results

Cypress seedling height and height growth pattern

The seedling height of Cypress in fertile soil was significantly greater than that in low-fertility soil (P < 0.01). The seedling height of Cypress did not differ significantly among the Ca²⁺ treatments (Table 2), but did differ significantly among the clones; the C1 clone was significantly taller than the C2 clone (Fig. 1A).

In both soil conditions, the seedling height exhibited a "slow-fast-slow" growth pattern (Fig. 2). In the fertile soil, the fast growth period with regard to seedling height began at 61 and 69 days under the 3 g kg⁻¹ (T2) and 6 g kg⁻¹ Ca²⁺ (T3) treatments,

respectively; the beginning date was delayed by 5 and 13 days, while the duration of the fast growth period was shortened by 9 and 14 days, respectively, compared with that of the 0 g kg⁻¹ Ca²⁺ (T1) treatment. In the low-fertility soil, the fast growth period for seedling height began at 76 days after the addition of 3 g kg⁻¹ Ca²⁺ (T5); the beginning date was advanced by 5 days, while the duration of the fast growth period was prolonged by 6 days compared to that of the 0 g kg⁻¹ Ca²⁺ (T4) treatment. In contrast, the fast growth period for seedling height began at 82 days after the addition of 6 g kg⁻¹ Ca²⁺ (T6); the beginning date was delayed by 1 day, while the duration of the fast-growth period was prolonged by 4 days compared with that of the 0 g kg⁻¹ Ca²⁺ treatment (T4).

Dry biomass

 Ca^{2+} addition had no significant effect on dry matter accumulation in the roots, stems or leaves of the Cypress C1 and C2 clones. The root, stem and leaf dry matter quality were significantly different among clones (Table 2). In the fertile soil, the mean dry biomasses of roots, stems and leaves of the C1 clone were 5.53, 5.97 and 10.46 g, respectively; these mean values were 30%, 87% and 86% higher than those of the C2 clone, respectively. In the low-fertility soil, the mean dry biomasses of roots, stems and leaves of the C1 clone were 3.58, 2.23 and 4.50 g, respectively; these mean values were 15.71%, 10.05% and 3.12% greater than those of the C2 clone (Fig. 1).

Root growth and development

The root length, root surface area and root volume of the D2–D4 diameter classes differed significantly among the Ca²⁺ treatments (Table 3). In the fertile soil, the addition of Ca²⁺ significantly inhibited the root length and root surface area of the D2 class. The sum of the root lengths of classes D1–D2 accounted for 90.9%, 90% and 89.1% of total root length across the three Ca²⁺ treatments (T1, T2 and T3, respectively); among these, the root length of the D2 class accounted for 32.0%, 35.2% and 36.8% of total root length, respectively. In the low-fertility soil,

Table 2. Variance analysis of seedling height, dry matter and nutrient absorption of two *C. funebres* clones under different NPK fertilizer and CaSO₄ treatments.

Traits	NPK	Ca ²⁺	Clones	NPK×Ca ²⁺	Clones imes NPK	$Clones \times Ca^{2+}$	$Clones \times NPK \times Ca^{2+}$
Seedling height	425.71**	0.50	6.74**	4.77**	7.34**	3.01**	1.53
Root dry matter	11.67**	0.43	12.08**	4.20*	11.19**	1.34	1.91
Stem dry matter	180.60**	1.65	8.96**	1.82	8.77**	2.63**	1.39
Leaf dry matter	191.75**	0.15	10.72**	0.69	7.56**	1.73	1.52
Nitrogen accumulation efficiencies	268.94**	4.03*	10.38**	0.87	6.29**	2.02*	2.32*
Phosphorus accumulation efficiencies	2.08	4.71**	1.61	1.69	1.09	2.73*	1.63
Calcium accumulation efficiencies	12.34**	39.14**	7.26**	4.62*	5.27**	2.71*	1.76

* stands for significant difference level (p<0.05), ** stands for extremely significant level (p<0.01).

the addition of 3 g kg⁻¹ Ca²⁺ (T5) promoted root development of classes D2–D4, and the resulting root lengths were 1.17-, 1.15- and 1.30-fold that under the 0 g kg⁻¹ Ca²⁺ treatment (T4), respectively. The sum of the root length of classes D1–D2 accounted for 90.7%, 93.2% and 92.6% of the total root length across the three Ca²⁺ treatments (T4, T5 and T6,

respectively). The corresponding root surface area and root volume also showed similar variation, with a 1.05–1.35-fold increase (Fig. 3).

The root length, root surface area and root volume differed significantly among Cypress clones (Table 3). In the fertile soil, the total root lengths of the C1 clone were 26.5%, 108.0% and 67.4% longer than



Fig. 1. Differential seeding height and dry mass of *C. funebris* clones in different solid environment and calcium treatments. Different small letters above the bars indicate the significant differences (p<0.05)



 $y_{(T1)}=31.61/(1+14.04 \times e^{-0.024 \times 120}), R^{2}=0.9814$ $y_{(T2)}=30.84/(1+16.33 \times e^{-0.024 \times 111}), R^{2}=0.9834$ $y_{(T3)}=33.51/(1+21.17 \times e^{-0.025 \times 106}), R^{2}=0.9892$ $y_{(T4)}=15.32/(1+35.61 \times e^{-0.027 \times 96}), R^{2}=0.9846$ $y_{(T5)}=19.40/(1+27.05 \times e^{-0.026 \times 102}), R^{2}=0.9854$ $y_{(T6)}=14.12/(1+32.85 \times e^{-0.026 \times 100}), R^{2}=0.9848$

Fig. 2. Seedling height growth curve of *C. funebris* under different calcium treatments. The dotted line in the picture indicates low fertile soil and the solid line indicates fertile soil. y is cumulative growth of seedling height of each treatment

those of the C2 clone across the three Ca^{2+} treatments (T1, T2 and T3, respectively). The root surface area and root volume of the C1 clone were also significantly greater than those of the C2 clone. In the low-fertility soil, the total root lengths of in the C1 clone were 11.5%, 7.0% and 25.1% greater than those of the C2 clone across the three Ca^{2+} treatments (T4, T5 and T6, respectively). Similar trends were observed for root surface area and root volume (Supplementary Fig. 1).

N, P and Ca accumulation efficiencies

The utilization efficiency of N in fertile soil was significantly greater than that in low-fertility soil, while the utilization efficiency of Ca^{2+} in low-fertility soil was significantly greater than that in fertile soil.

No significant difference was observed in P use efficiency between the two soil fertility conditions.

The accumulation efficiencies of N, P and Ca all significantly differed among the Ca^{2+} treatments (Table 3). In fertile soil, N accumulation efficiency for both clones exhibited a downward trend with increasing Ca^{2+} concentration. Both clones achieved their highest Ca accumulation efficiency under the 6 g kg⁻¹ Ca²⁺ treatment (T3) in the fertile soil. In the low-fertility soil, both the C1 and C2 clones achieved their highest N, P and Ca accumulation efficiencies under the 3 g kg⁻¹ Ca²⁺ treatment (Fig. 4).

The N and Ca accumulation efficiencies significantly differed between clones, but no significant difference was detected in P accumulation efficiency between clones. In the fertile soil, the N accumulation efficiencies for C1 were 10.64%, 13.52% and



Fig. 3. Effects of calcium supply levels on root morphology of *C. funebris* clones under two soil fertility levels. Different small letters above the bars indicate the significant differences (p<0.05)

3.04% higher than those for C2 across the three Ca²⁺ treatments (T1, T2 and T3, respectively). The Ca accumulation efficiencies for C1 were 16.94%, 10.39% and 6.84% higher than those for the C2 clone (Supplementary Fig. 2). In the low-fertility soil, the N accumulation efficiencies for C1 were 5.16%, 12.69% and 11.09% higher than those for C2 across the three Ca²⁺ treatments (T4, T5 and T6, respectively). The Ca accumulation efficiencies for C1 were 22.93%, 15.54% and 12.78% higher than those for C2.

The interaction effect

There was significant interaction effect between NPK fertilizer and Ca²⁺ for seedling height, dry matter mass and root development of the D1–D3 diameter stages of Cypress (Table 2).

A significant interaction was found between clones and Ca²⁺ fertilizer for seedling height, root length, root surface area and root volume of the D3–D5 diameter stages of Cypress (Table 2). In the low-fertility soil, treatment with 3 g kg⁻¹ Ca²⁺ significantly promoted seedling height growth, resulting in 34.22% and 32.19% increases in seedling height in the C1 and C2 clones, respectively, compared to the 0 g kg⁻¹ Ca²⁺ treatment. In contrast, seedling height showed a decrease following the addition of 6 g kg⁻¹ Ca²⁺ (Supplementary Fig. 3). In the fertile soil, the root length, root surface area and root volume of the D3-D5 diameter classes all significantly decreased in the C2 clone following Ca²⁺ addition. In contrast, Ca²⁺ addition had no significant effect on root length, root surface area or root volume in the C1 clone (Supplementary Fig. 1).

Table 3. Variance analysis of root length, surface area and volume of each diameter class of *C. funebres* clones under different NPK fertilizer and CaSO₄ treatments

Diameter cla	NPK	Ca ²⁺	Clones	NPK×Ca ²⁺	Clones×NPK	$Clones \times Ca^{2+}$	$Clones \times NPK \times Ca^{2+}$	
Root length (cm) 0–0.5 (D1)		1.84	0.60	7.03**	4.62*	3.57**	0.89	1.68
	0.5–1.0 (D2)	125.78**	12.93**	13.21**	5.72**	5.88**	0.94	1.99*
	1.0–1.5 (D3)	173.76**	15.01**	9.22**	4.79**	6.34**	2.05*	2.30*
	1.5–2.0 (D4)	170.69**	4.58*	15.57**	2.23	10.13**	3.79**	2.5*
	≥2.0 (D5)	48.68**	0.62	15.02**	0.13	8.53**	3.37**	0.94
Root Surface area (cm ²)	0–0.5 (D1)	3.49*	0.82	8.15**	3.94*	3.88**	0.88	1.79
	0.5–1.0 (D2)	132.24**	13.56**	12.81**	6.00**	5.86**	0.93	1.98*
	1.0–1.5 (D3)	174.98**	14.69**	9.35**	4.57*	6.46**	2.12*	2.34*
	1.5–2.0 (D4)	169.13**	4.38*	15.64**	2.23	9.99**	3.75**	2.45*
	≥2.0 (D5)	54.97**	0.98	13.52**	0.28	8.01**	4.02**	0.87
Root volume (cm ³)	0–0.5 (D1)	6.90**	1.28	9.03**	3.18*	4.08**	0.89	1.84
	0.5–1.0 (D2)	138.44**	14.17**	12.38**	6.24**	5.81**	0.92	1.98*
	1.0–1.5 (D3)	176.23**	14.33**	9.51**	4.35*	6.58**	2.19*	2.38*
	1.5–2.0 (D4)	167.72**	4.19*	15.68**	2.25	9.82**	3.70**	2.40*
	≥2.0 (D5)	61.20**	1.18	12.09**	0.90	6.65**	4.18**	0.86

* - significant difference level (p<0.05), ** - extremely significant level (p<0.01).



Fig. 4. Effects of calcium supply levels on absorption efficiency of nitrogen, phosphorus and calcium of *C. funebris* clones under two soil fertility levels. Different small letters above the line indicate the significant differences (p<0.05)

Discussion

It is generally thought that a plant is not deficient in Ca^{2+} when the exchangeable Ca^{2+} content in soil is greater than 400 mg kg⁻¹. In the present study, the experimental soil contained an exchangeable Ca2+ content of 128 mg kg⁻¹, which was lower than the cutoff value without Ca²⁺ application, thus resulting in Ca²⁺-deficient soil (Ji et al., 2009). Compared to that of the low-fertility soil, the seedling height of Cypress growing in fertile soil showed a reduced response to Ca^{2+} . The addition of Ca^{2+} fertilizer reduced the N accumulation efficiency in Cypress. These observations indicate that the N and P nutrients were sufficient in the fertile soil, exchangeable Ca²⁺ was exchanged with a large amount of NH_4^+ and K⁺, which facilitated the desorption of exchangeable Ca²⁺, and N accumulation efficiency decreased with increasing Ca²⁺ concentration (Li et al., 2017). Moreover, exchangeable Ca²⁺ and PO₄³⁻ underwent irreversible ion exchange, and the increase in P promoted the conversion of water-soluble and exchangeable Ca²⁺ to unavailable non-acid-soluble Ca, which also resulted in a decrease in the exchangeable Ca²⁺ in soil (White & Broadley, 2003; Hirschi, 2004). This may be why Ca use efficiency was higher under a high Ca²⁺ concentration; however, the exact process still requires further investigation. In the low-fertility soil, the seedling height of Cypress was increased when an appropriate amount of Ca^{2+} (3 g kg⁻¹) was added, and the highest N, P and Ca accumulation efficiencies were all achieved the under the 3 g kg⁻¹ Ca²⁺ treatment, with a synergy between Ca²⁺ fertilizer versus N and P accumulation efficiencies. However, when the Ca^{2+} concentration was increased (6 g kg⁻¹ Ca²⁺ treatment), seedling growth of Cypress clones decreased. These results indicate that the synergy had a range of adaptation to the amount of Ca^{2+} applied. That is, an appropriate amount of Ca^{2+} promoted plant N and P uptake, while an excessively high concentration of Ca2+ fertilizer resulted in an inhibitory effect (Borchert, 1986; Franceschi, 1989). In a study conducted on coniferous species such as pine (Pinus massoniana Lamb.), good adaptation was also observed in the soil environment with Ca²⁺ supplied at $1-2 \text{ mmol } L^{-1}$, while the plant height growth of pine seedlings decreased after the Ca²⁺ supply exceeded this concentration (Li et al., 2017). Therefore, full consideration should be given to the tolerance of tree species when applying Ca^{2+} to promote seedling growth.

Root architecture refers to the spatial distribution of plant roots in the soil, and it reflects the specific root characteristics of plants that form during evolution for adaptation to the environment (Eissenstat & Yanni, 2002; Mou et al., 2013). In the low-fertility soil, the addition of 3 g kg⁻¹ Ca²⁺ promoted root development of the D2-D4 classes (diameter 0.5-2.0 mm) in the C1 clone; when the Ca²⁺ concentration was increased to 6 g kg⁻¹, root length, root surface area and root volume increases of classes D2-D4 were inhibited in this clone. Moreover, in the fertile soil, Ca²⁺ addition inhibited root length, root surface area and root volume increases of classes D1-D5 of the C2 clone. These results demonstrate that the roots of different diameter classes have distinct reactions to Ca^{2+} (Mou et al., 2013; Kanno et al., 2016). McCormack's method is used for functional classification of fine roots; fine roots with a diameter of 0-1.0 mm are absorptive roots, and fine roots with a diameter of 1.0–2.0 mm are transport roots (Mc-Cormack et al., 2015). The root length of absorptive roots accounted for 91.1% of the total root length, the root surface area of absorptive roots accounted for 74.8% of the total root surface, and the root volume of absorptive roots accounted for 45.2% of total root volume; the proportions of transport roots were 8.0%, 19.6% and 27.2%, respectively. Absorptive roots are the most active components of fine roots of trees; their lignification degree is relatively low and closely related to the apical region, which is sensitive to soil nutrient changes. The root length and root surface area of absorptive fine roots enable plants to easily respond to changes in the soil environment (Eissenstat & Yanni, 2002; Guo et al., 2008; Mou et al., 2013; Band & Bennett, 2013). The proportion of fine roots with a diameter of 0-1.0 mm in low-fertility soil was higher than that in fertile soil. In the fertile soil, lateral root growth was inhibited with increasing Ca²⁺ concentration, which indicates that Cypress roots adapted to the Ca²⁺ environment by reducing lateral root growth and soil contact area under Ca²⁺ stress. Under low-fertility soil conditions, Cypress could adjust the morphology of fine roots to adapt to different Ca²⁺ environments, increasing the fine root diameter to ≤ 1.0 mm to improve their ability to obtain nutrients (Hodge, 2004).

Conclusions and implications for forest practice

In conclusion, adding calcium fertilizer under two soil conditions affected the growth of fine roots of Cypress clonal seedlings and the element accumulation of N, P and Ca, thus influencing seedling growth. Under fertile soil conditions, addition of Ca fertilizer inhibited effective N accumulation of Cypress and significantly inhibited the development of root length in roots 0.5–1.0 mm in diameter. Therefore, for Cypress clones grown under fertile soil conditions, addition of Ca²⁺ fertilizer should not be considered. Under low-fertility soil conditions, the addition of CaSO₄ can promote the root development of seedlings to some extent and advance and prolong the fast growth period of seedling height. Therefore, Cypress clones can be used as a major tree species for afforestation under low-fertility soil conditions and especially under calcareous soil conditions.

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