

THE OCCURRENCE OF *Fusarium* spp. ON OAT (*Avena sativa* L.) AND SUSCEPTIBILITY OF SEEDLINGS OF SELECTED GENOTYPES TO INFECTION WITH *Fusarium graminearum* SCHWABE

Irena Kiecana, Elżbieta Mielniczuk,
Małgorzata Cegiełko, Alina Pastucha

Department of Phytopathology and Mycology, University of Life Sciences in Lublin
Leszczyńskiego 7, 20-069 Lublin, Poland
e-mail: irena.kiecana@up.lublin.pl

Received: 08.11.2013

Abstract

The present study was carried out in the years 2010–2012 in the fields of the Strzelce Plant Breeding Company Ltd., belonging to the Plant Breeding and Acclimatization Institute in Radzików, and it included 39 oat genotypes. At the six-week seedling stage, the percentage of plants with root and leaf sheath necrosis symptoms was evaluated. In 2010 the percentage of seedlings with disease symptoms ranged from 6.5% to 25%, in 2011 it ranged from 17% to 34.5%, whereas in 2012 from 10% to 25%. In 2010 the disease index ranged from 1.4 to 5.7, in 2011 from 4.5 to 8.8, while in 2012 it was between 2.0 and 5.4.

Mycological analysis showed that large numbers of *Fusarium* spp. colonies were obtained both from the roots and leaf sheaths. Isolates of these fungi accounted for 63.48% of the total fungi isolated from seedlings. Seedlings grown under the conditions of central Poland were damaged by the species *F. culmorum*, *F. avenaceum*, and *F. solani*. The investigation of the susceptibility of 15 oat genotypes to infection with two *Fusarium graminearum* strains – Tz 56 and Tk 235 – was carried out under growth chamber conditions at a temperature of 22–23°C and relative air humidity of 85%. The *F. graminearum* strain Tz 56 proved to be the most pathogenic to seedlings of the breeding lines STH 0.9403 and POB 1316/08, for which the disease index was 80.5 and 75.5, respectively. The lowest pathogenicity of the a.m. strain was recorded in the case of the genotype DC 1832/05, for which the disease index was 26.5. The *F. graminearum* strain Tk 235 proved to be the most pathogenic to the genotypes STH 0.9403 and STH 0.9423, for which the disease index was 70.5 and 70.0, respectively, whereas this strain was least pathogenic to the breeding line DC 2112/05, in the case of which the disease index was 25.5.

Key words: *Avena sativa*, genotypes, seedling damping-off, susceptibility, *Fusarium graminearum*

INTRODUCTION

Due to the introduction of reduced tillage systems, the problem of infection of seedlings as well as of the roots and stem base of cereals with fungi of the genus *Fusarium* is gaining special importance [1, 2, 3, 4, 5]. The oat is considered to be a species that produces good yields in crop rotations with a large proportion of cereals, but it is infected by fungi of the genus *Fusarium*, one of the components causing root and stem rot diseases [6, 1]. *F. culmorum* and *F. avenaceum* have been recognized as the main cause of these diseases in oat crops [7, 1, 2]. These fungi are distinguished by abundant conidial sporulation. Macroconidia of these fungi are formed in sporodochia developing in crop residue on the soil surface [8]. The above-mentioned *Fusarium* species are also characterized by high tolerance to temperature and humidity [1, 9]. Moreover, seedlings and the stem base of cereals are damaged by the species *F. graminearum*; seed material is thought to be the main source of primary infection of cereal seedlings with this fungus [10, 11, 12, 13]. The species *F. avenaceum* (teleomorph: *Gibberella avenacea*) and *F. graminearum* (teleomorph: *G. zeae*) form on cereal crop residue and on the lower nodes of the stem base perithecia containing ascospores, which are carried by the wind for long distances [12, 14, 15].

The aim of the present study was to determine the contribution of fungi of the genus *Fusarium* to the infection of oat seedlings grown under the conditions of central Poland as well as to determine the pathogenicity of *F. graminearum* to seedlings of several oat genotypes under controlled temperature and humidity conditions.

MATERIALS AND METHODS

The study on the occurrence of *Fusarium* spp. on oat was conducted in the years 2010–2012 in the fields of the Strzelce Plant Breeding Company Ltd., belonging to the Plant Breeding and Acclimatization Institute in Radzików. 15 oat genotypes were included in the study each year and a total of 39 genotypes were investigated (a list of cultivars and breeding lines is shown in Table 1). At the six-week seedling stage, 200 (4x50) seedlings of each genotype were sampled. The sampling method was the same as the one used in the study of oat conducted by Kiecana et al. [1].

The percentage of plants with root and leaf sheath necrosis symptoms was evaluated in a laboratory using a five-point scale and the disease index was calculated in the same way as in the study of oat conducted by Kiecana et al. [2]. The obtained results were statistically analyzed using T-Tukey's confidence half-intervals [16].

Subsequently, a mycological analysis of the diseased plants was performed. The number of pieces of seedlings collected for analysis and the analysis method were the same as in the study by Kiecana and Mielniczuk [7]. The cultures of *Fusarium* spp. were identified to the species level using monographs and keys by Kwaśna et al. [17] and Leslie and Summerell [18].

The infection experiment to evaluate the susceptibility of seedlings of 15 oat genotypes to infection by *Fusarium graminearum* No. Tz 56 obtained from kernels and with *F. graminearum* No. Tk 235 isolated from the roots was conducted in a growth chamber at a temperature of 23–24°C and relative air humidity of 85%. The strains whose pathogenicity had been earlier tested in the laboratory by the method of Mishra and Behr [19] were used in the investigation. 14-day-old cultures of *F. graminearum* grown on PDA medium in Petri dishes at 22°C were used as fungal inoculum.

Kernels of the analyzed oat genotypes whose sprouts had reached a length of 10 mm and were normally developed were selected for the investigation of susceptibility. The selected material was placed on a PDA layer with the studied fungal strains, which were in plastic pots with a diameter of 10 cm and a height of 15 cm filled with all-purpose growing medium with an addition of sand at a 2:1 ratio and a pH 6.5, previously sterilized twice in an autoclave for two hours at 121°C and then covered with the medium [20]. Pots in which sprouted kernels were placed on layers of PDA medium without fungus were the control. The experiment was established on 4 May 2012 and was performed in four replicates with 25 plants per replicate. Plants grew for 24 days and then the degree of infection of seedlings was determined using a four-point scale and

the disease index was calculated in the same way as in the study by Kiecana and Kocyłak [21]. The results were statistically analyzed in the same way as in the case of plants collected from plantations.

Ten seedlings with disease symptoms from each treatment of the growth chamber experiment were designated for mycological analysis, which was carried out in the same way as in the case of plants grown under field conditions. The cultures of fungi were identified to the species using monographs and keys [22, 23, 24, 25, 26, 27].

RESULTS

Spring observations revealed the occurrence of oat seedlings with root and leaf sheath necrosis. In 2010 the percentage of plants with disease symptoms varied from 6.5% (POB 648/06) to 25% (CHD 2316/03) (Fig. 1), in 2011 from 17% (STH 9110) to 34.5% (STH 9410) (Fig. 2), whereas in 2012 from 10% (POB 4129-4416/11) to 25% (DC 3674/02) (Fig. 3). In 2010 the significantly highest disease index was observed in the case of the breeding line CHD 2316/03 (5.7), whereas the significantly lowest value was found in the breeding line POB 648/06 (1.4) (Table 1). In 2011 the significantly highest disease index was recorded in the case of the genotype DC 1329/05 (8.8), while the significantly lowest index was found for the genotypes Nagus (4.5) and POB 1882/09 (4.7). In 2012 the significantly highest disease index was found in the case of the following genotypes: DC 3674/02 (5.7), DC 2112/05 (5.4), STH 0.8124 (5.3), and POB 2885/08 (5.3), whereas the significantly lowest value was found in the breeding line POB 4129-4416/11 (2.0) (Table 1).

The mycological analysis showed that large numbers of *Fusarium* species were obtained both from the roots and leaf sheaths. On average for the three-year study period, isolates of these fungi accounted for 63.48% of total isolates from oat seedlings. Among pathogenic species, the following were isolated from seedlings: *F. culmorum*, whose isolates accounted for 19.84% of total isolations, *F. solani* (6.70% of all fungi obtained from seedlings), *F. avenaceum* (7.04%), *F. sporotrichioides* (2.22%), *F. equiseti* (2.30%), *F. graminearum* (1.39%), and *F. crookwellense* (0.08%). Moreover, *F. oxysporum* (21.76% of all fungi) and *F. poae* (2.15 % of total isolates) were isolated from oat seedlings (Table 2).

The study in the growth chamber showed that plants with disease symptoms occurred in the experimental treatments with oat kernels infected with both *Fusarium graminearum* Tz 56 and *F. graminearum* Tk 235. Plant losses caused by pre- and post-emergence damping-off were recorded in the case of both

investigated *F. graminearum* strains. Infected plants were characterized by root and hypocotyl necrosis and quite often by hypocotyl contraction and root reduction. Furthermore, infected oat seedlings showed inhi-

bited growth and yellowing leaves. Control seedlings of the tested genotypes did not exhibit distinct disease symptoms; they had a well-developed root system and root necrosis was observed sporadically.

Table 1
Mean values of the disease index for seedlings of the tested oat genotypes grown under field conditions in 2010–2012

No.	Oat genotypes	2010	2011	2012
1.	ARDEN	2.8 ab	8.4cde	
2.	NAGUS n	3.5 bc	4.5a	
3.	DC 1193/04	3.0 b	5.7abcd	
4.	DC 1329/05		8.8e	
5.	CHD 2316/03	5.7 d		
6.	CHD 3047/03	3.5 bc		
7.	DC 1776/04		7.9bcde	
8.	DC 1832/05			3,6ab
9.	DC 2112/05			5,4b
10.	DC 2359/03			4,4ab
11.	DC 239/06			2,8ab
12.	DC 3674/02 n			5,7b
13.	POB 648/06	1.4 a		
14.	POB 1316/08	3.5 bc		3,5ab
15.	POB 1813/08	2.9 ab		
16.	POB 2053/06	3.4 bc		
17.	POB 4703-99/08	1.7 ab		
18.	POB 342/09		7.9bcde	
19.	POB 1882/09		4.7a	
20.	POB 2085/09		5.5abc	
21.	POB 2149/09		8.2bcde	
22.	POB 158/09		6.5abcde	
23.	POB 2842/08			2,9ab
24.	POB 2885/08			5,3b
25.	POB 3034 /08			3,1ab
26.	POB 4129-4416/11			2,0a
27.	BINGO	3.7 bc	8.6de	
28.	MACZO n	3.2 bc	5.2ab	
29.	Haker	4.4 cd		
30.	STH 8608	3.8 bc		
31.	STH 8809	2.8 ab		
32.	STH 9110		5.3ab	
33.	STH 9210		7.1abcde	
34.	STH 9410 n		8.6de	
35.	STH 0.8124			5,3b
36.	STH 9322			3,2ab
37.	STH 0.9403 n			4,1ab
38.	STH 0.9423 n			2,9ab
39.	STH 0.9770			4,8ab
	Mean	3.29	8.86	3.93

Means in columns followed by the same letter are not significantly different at $P \leq 0.05$

n – naked oat

Table 2
Fungi isolated from oat seedlings grown under field conditions in 2010–2012

Fungal species	Number of isolates (%)											Total number of isolates
	2010			2011			2012			Total		
	r	ls	r+ls	r	ls	r+ls	r	ls	r+ls	r	ls	
<i>Fusarium avenaceum</i> (Fr.) Sacc.	-	2	2 (0.38)	39	86	125 (11.92)	7	53	60 (5.57)	46	141	187 (7.04)
<i>Fusarium crookwellense</i> Burgess, Nelson & Tousson	-	-	-	-	-	-	-	2	2 (0.19)	-	2	2 (0.08)
<i>Fusarium culmorum</i> (W. G. Sm.) Sacc.	4	9	13 (2.45)	128	129	257 (24.5)	76	181	257 (23.86)	208	319	527 (19.84)
<i>Fusarium equiseti</i> (Corda) Sacc.	4	21	25 (4.72)	7	24	31 (2.96)	-	5	5 (0.46)	11	50	61 (2.30)
<i>Fusarium graminearum</i> Schwabe	-	1	1 (0.19)	15	17	32 (3.05)	1	3	4 (0.37)	16	21	37 (1.39)
<i>Fusarium oxysporum</i> Schl.	46	129	175 (33.02)	102	73	175 (16.68)	95	133	228 (21.17)	243	335	578 (21.76)
<i>Fusarium poae</i> (Peck.) Wollenw.	1	-	1(0.19)	-	13	13 (1.24)	15	28	43 (3.99)	16	41	57 (2.15)
<i>Fusarium solani</i> (Mart.) Sacc.	2	2	4 (0.75)	76	29	105 (10.01)	42	27	69 (6.41)	120	58	178 (6.70)
<i>Fusarium sporotrichioides</i> Sherb.	9	12	21 (3.96)	8	-	8 (0.76)	11	19	30 (2.79)	28	31	59 (2.22)
Other colonies	174	114	288 (54.34)	117	186	303 (28.88)	188	191	379 (35.19)	479	491	970 (36.52)
Total	240	290	530	492	557	1049	435	642	1077	1167	1489	2656

r-roots

ls-leaf sheaths

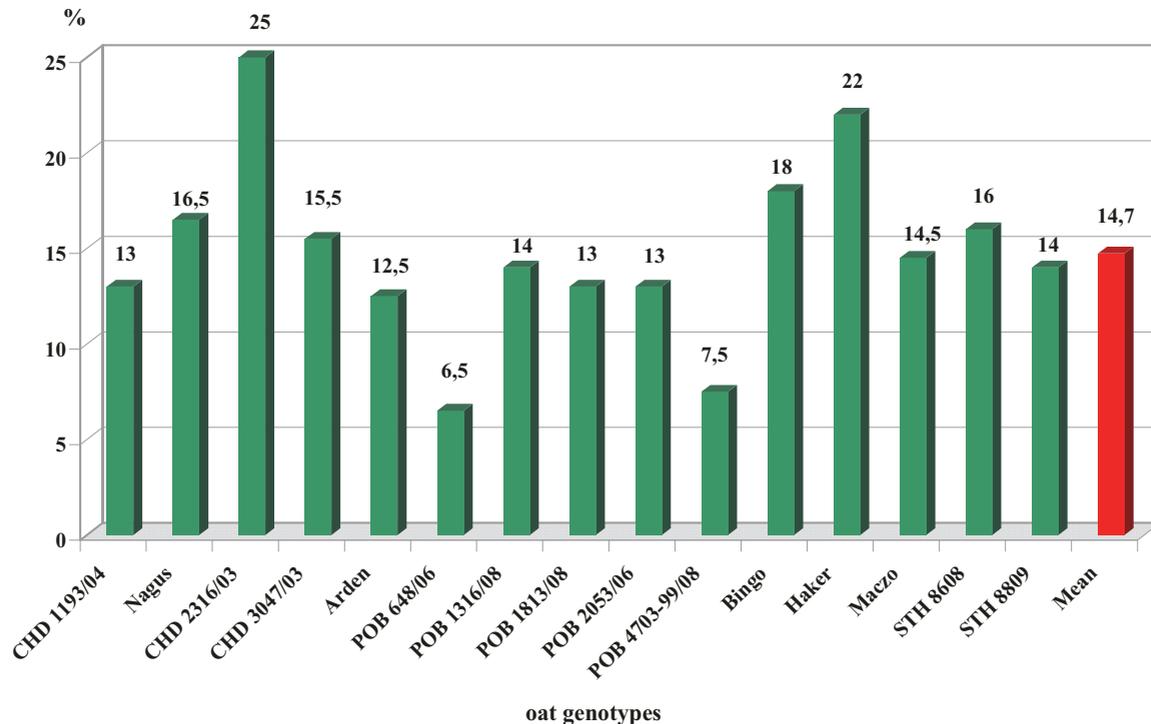


Fig. 1. Percentage of plants with root and leaf sheath necrosis symptoms in 2010.

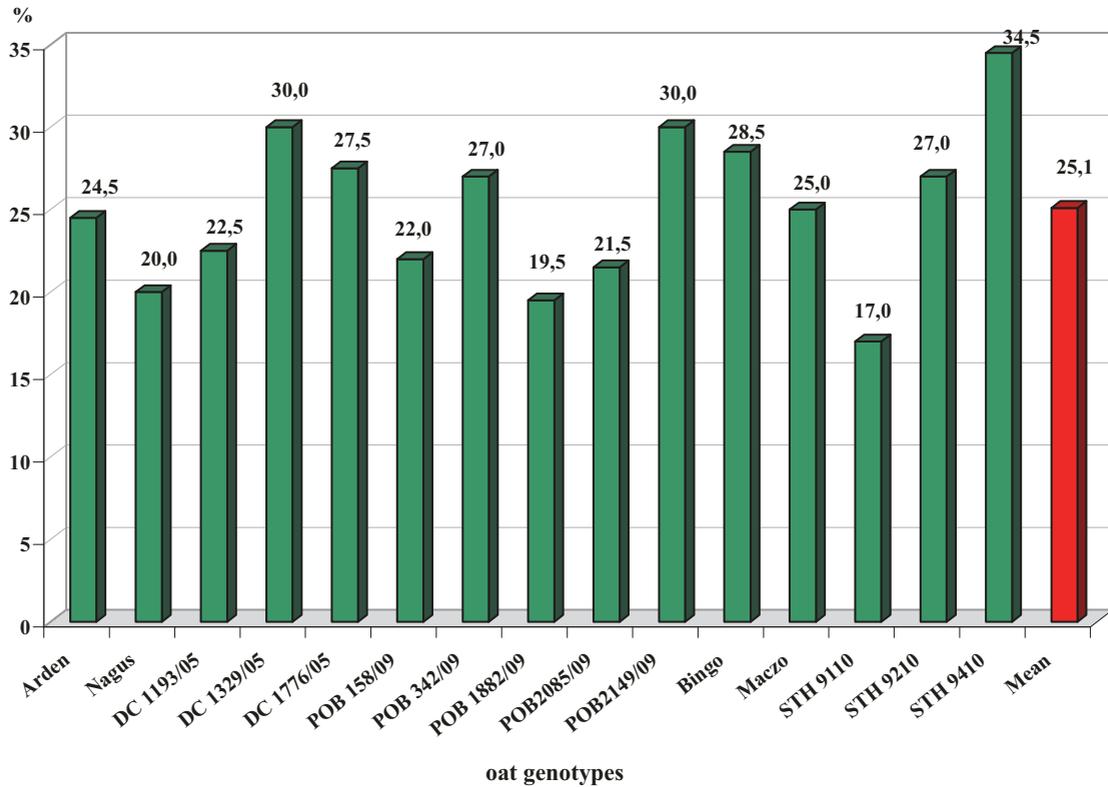


Fig. 2. Percentage of plants with root and leaf sheath necrosis symptoms in 2011.

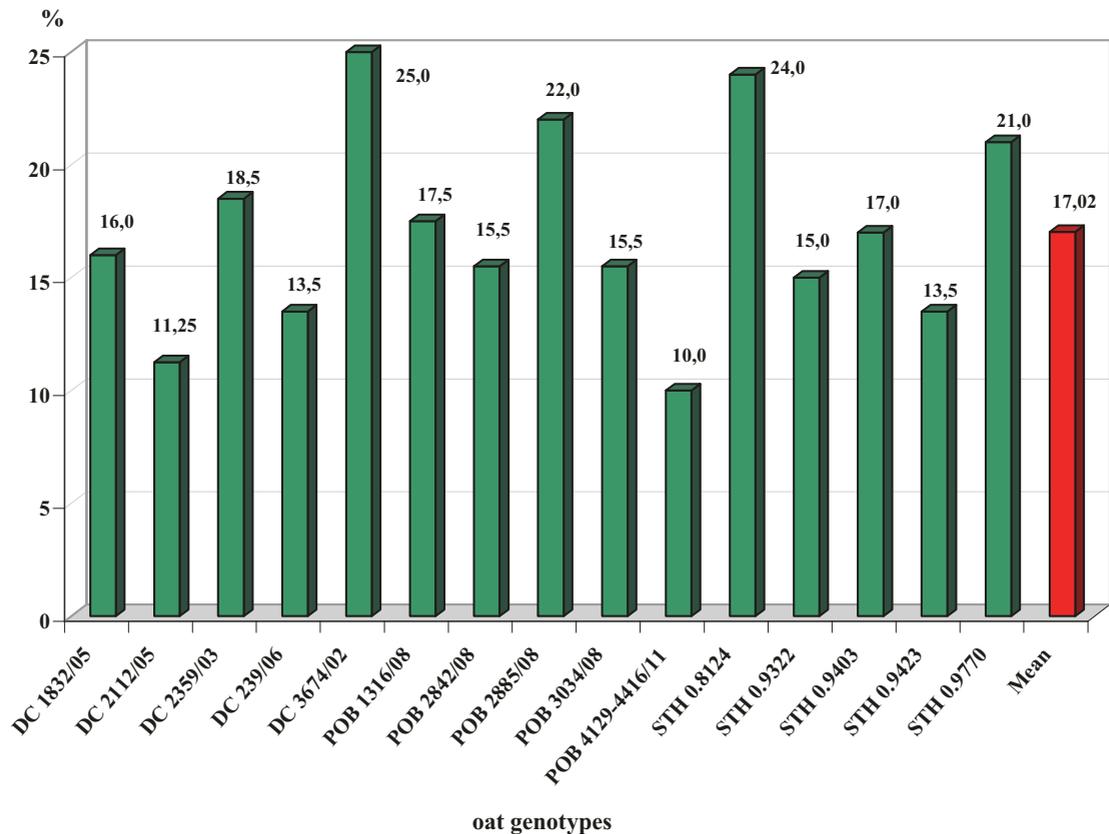


Fig. 3. Percentage of plants with root and leaf sheath necrosis symptoms in 2012.

On the basis of the statistical analysis of the disease indices, seedlings of the oat genotypes in question were found to show varying susceptibility to infection by the studied strains of *Fusarium graminearum*. The statistical analysis of the disease indices demonstrated that artificial inoculation of the medium with the *F. graminearum* strain Tz 56 had a significant effect on the health of investigated seedlings compared to the control treatment in the case of all studied oat genotypes (Table 3).

The *F. graminearum* strain Tz 56 proved to be most pathogenic to seedlings of the breeding lines STH 0.9403 and POB 1316/08, for which the disease index was 80.5 and 75.5, respectively. The lowest pathogenicity of the a.m. strain was found in the case of the genotypes DC 1832/05 and STH 0.9322, for which the disease index was 26.5 and 32.0, respectively (Table 3).

The present study showed that inoculation of the medium with the *F. graminearum* strain Tk 235 also had a significant effect on the infection of oat seedlings in all tested genotypes compared to the control (Table 3).

The *F. graminearum* strain Tk 235 proved to be most pathogenic to the genotypes STH 0.9403 and STH 0.9423, for which the disease index was 70.5 and 70.0, respectively, while it was least pathogenic to the breeding lines DC 2112/05, POB 4129-4416/11, DC 1832/05 and DC 239/06, in the case of which the disease index was, respectively, 25.5, 26.75, 28.25, and 29.25 (Table 3).

The disease index for seedlings in the control treatment ranged from 0.5 to 2.25 (Table 3).

The mycological analysis of infected oat seedlings indicates that the *F. graminearum* was the cause of pre- and post-emergence damping-off (Table 4).

Table 3
Mean values of the disease index for seedlings of the tested oat genotypes in the experimental treatment with *Fusarium graminearum* No. Tz 56, *F. graminearum*, No. Tk 235 and in the control treatment under growth chamber conditions.

Oat genotypes	Experimental treatment		
	<i>Fusarium graminearum</i> No. Tz 56	<i>Fusarium graminearum</i> No. Tk 235	Control
DC 1832/05	26.5*a	28.25*a	1.25
DC 2112/05	55.75*bcd	25.5*a	1.25
DC 2359/03	49.0*bc	51.0*cd	1.75
DC 239/06	54.75*bcd	29.25*a	0.5
DC 3674/02n	65.25*de	68.5*ef	1.25
POB 1316/08	75.5*ef	59.25*def	1.25
POB 2842/08	47.25*b	56.75*cde	1.0
POB 2885/08	57.0*bcd	50.75*cd	1.5
POB 3034 /08	49.0*bc	35.0*ab	1.0
POB 4129-4416/11	49.0*bc	26.75*a	1.75
STH 0.8124	54.5*bcd	48.0*c	2.25
STH 0.9322	32.0*a	46.0*bc	1.5
STH 0.9403n	80.5*f	70.5*f	0.5
STH 0.9423n	60.0*cd	70.0*f	1.0
STH 0.9770	51.25*bc	53.75*cd	0.75
Mean	54.32	47.95	1.23

*Means in lines are significantly different compared to the control at $0 \leq 0.05$

– Means in columns followed by the same letter are not significantly different at $0 \leq 0.05$

n – naked oat

Table 4
Fungi isolated from oat seedlings grown in the growth chamber experiment with inoculation of the medium with the *Fusarium graminearum* strain No. Tz 56 and the *F. graminearum* strain No. Tk 235 and in the control treatment.

Fungal species	Experimental treatment			Total number of isolates
	<i>F. graminearum</i> No. Tz 56	<i>F. graminearum</i> No. Tk 235	Control	
<i>Acremonium roseum</i> (Oud.) W. Gams	-	-	19	19
<i>Alternaria alternata</i> (Fr.) Keissler	6	8	43	57
<i>Aspergillus niger</i> van Tiegh.	3	-	-	3
<i>Bipolaris sorokiniana</i> (Sacc.) Shoemaker	3	-	-	3
<i>Epicoccum nigrum</i> Link	27	2	41	70
<i>Fusarium culmorum</i> (W.G. Sm.) Sacc.	-	2	15	17
<i>Fusarium equiseti</i> (Corda) Sacc.	-	3	6	9
<i>Fusarium graminearum</i> Schwabe	738	786	7	1531
<i>Fusarium oxysporum</i> Schl.	6	6	20	32
<i>Fusarium poae</i> (Peck.) Wollenw.	-	2	25	27
<i>Fusarium sporotrichioides</i> Sherb.	-	-	24	24
<i>Mucor hiemalis</i> Wehmer	42	3	10	55
<i>Mucor mucedo</i> Fres.	9	6	-	15
<i>Penicillium puberulum</i> Bainier	6	-	29	35
<i>Penicillium verrucosum</i> Dierckx var. <i>cyclopium</i> (Westling.) Samson et al.	18	4	35	57
<i>Rhizopus nigricans</i> Ehrenberg	6	-	20	26
<i>Trichoderma aureoviride</i> Rifai	-	-	11	11
<i>Trichoderma harzianum</i> Rifai	-	-	23	23
<i>Trichoderma koningii</i> Oud.	15	8	47	70
<i>Trichoderma viride</i> Persoon ex S.F. Gray	23	3	-	26
Total	902	833	375	2110

DISCUSSION

Field observations carried out in central Poland revealed that oat plants with root and leaf sheath necrosis symptoms occurred at a higher percentage than in the case of oat grown in the eastern area of the Lublin region [1].

The species *Fusarium culmorum*, *F. avenaceum* and *F. solani* proved to be the cause of damage of oat seedlings in the spring. The pathogenicity of *F. avenaceum* and *F. culmorum* to oat seedlings has also been confirmed by the research of Kiećana and Kocylak [21] as well as of Mańka [20]. *Fusarium culmorum* has been found to show high pathogenicity to seedlings of various cereal species [20, 28, 29].

Secondary metabolites, including deoxynivalenol (DON), participate in the disease process caused by *Fusarium culmorum*. This metabolite exhibits phytotoxic activity in particular against the coleoptile, shoots and callus tissue of wheat [30]. Moreover, DON reduces the germination capacity of seeds and

causes a reduction in the number and length of radicles of wheat seedlings. Reduced germination capacity and growth inhibition in seedlings occur already at a DON concentration of 4 µg/ml, whereas the use of a concentration of 25 µg/ml completely inhibits root growth and mitotic activity in the cells of the apical meristems of DON-treated roots. This mycotoxin leads to disturbances in the mitotic division of the meristematic cells of wheat roots [31]. In wheat roots subjected to the effect of DON, these authors observed the occurrence of chromatin bridges in the chromosomes and they also found the occurrence of lagging chromosomes in the anaphase. According to Paćka [32], DON leads to excessive condensation of chromosomes in the metaphase and anaphase and disturbs the synthesis of proteins making up the microtubules of the karyokinetic spindle. Other secondary metabolites can also participate in the pathogenesis of diseases caused by *F. culmorum*, including culmorin, which can inhibit wheat coleoptile elongation at a concentration from 100 µM to 1 mM [33].

Fusarium avenaceum is considered to be a serious pathogen of cereals, including oat [1, 34, 35]. Moniliformin participates in the pathogenesis of diseases caused by *F. avenaceum*; this mycotoxin affects the permeability of the cytoplasmic membranes and causes disturbances in mitotic cell division [32].

Fusarium solani has contributed to root infection in single cropped oat and in oat mixtures with spring barley and spring triticale as well as in triticale grown with oat. This fungus was the main cause of root damage in winter wheat grown after previous crops such as single cropped oat and oat grown in mixtures [35]. This species is considered to be an important producer of fusaric acid (5-(butyl)-2-pyridinecarboxylic acid) [36], a phytotoxin that destroys the vascular bundles in plants. The strong phytotoxicity of this metabolite to rice seedlings has been proved [33].

Fusarium equiseti belonged to the species isolated from infected oat roots and leaf sheaths. In the study by Majchrzak et al. [37], this fungus proved to be the main causal agent of 190 infections of wheat roots and had a significant contribution to stem base damage in this plant grown in the conditions of the north-eastern regions of Poland. *Fusarium oxysporum*, which is not considered to be a pathogen of cereal plants [38], was a species that was isolated in each year of the study.

The results obtained in the growth chamber experiment confirmed the high phytotoxicity of *Fusarium graminearum* to oat seedlings. Under the studied conditions in which 14-day cultures of the tested *F. graminearum* strains grown on dextrose-potato medium were used as an inoculum, this method proved effective, since there were seedlings with pre- and post-emergence damping-off in the studied oat genotypes in the experimental treatments with both *F. graminearum* Tz 56 obtained from grass seed and with *F. graminearum* Tk 235 obtained from grass roots. The present growth chamber study and previous information in the literature indicate the contribution of *F. graminearum* to the damage of cereal seedlings [10, 11].

CONCLUSIONS

1. The species *F. culmorum* and *F. avenaceum* are a threat to oat seedlings grown in central Poland.
2. Due to the high pathogenicity of these species, these pathogens should be taken into account in the breeding of new oat cultivars.
3. The breeding line DC 1832/05 seems to be useful in breeding oat for resistance to *F. graminearum*.

Acknowledgements

Research conducted in the period 2010–2012 supported by the Ministry of Agriculture and Rural

Development, projects No. HOR hn-078 dec-27/10, HOR hn 801-8/11, and HOR hn 801-1/12.

Author's contributions

Concept of the study: IK; writing of the manuscript: IK, EM, MC, AP; statistical analysis: MC, EM; mycological analysis: IK, AP, MC, EM; analysis of research results: IK, MC, EM, field research: IK, EM, MC, AP; experiment in the growth chamber: AP, MC, EM.

REFERENCES

1. Kiecana I, Mielniczuk E, Cegiełko M, Pszczołkowski P. 2003. Badania nad chorobami podsuszkowymi owsa (*Avena sativa* L.) z uwzględnieniem temperatury i opadów. / Investigations on root and stem rot diseases of oat (*Avena sativa* L.) with a special regard to temperature and rainfalls. Acta Agrobot. 56, 1–2: 95–107. (in Polish)
2. Kiecana I, Mielniczuk E, Cegiełko M. 2008. Grzyby porażające korzenie i podstawę źdźbła owsa (*Avena sativa* L.). / Fungi infecting roots and stem bases in oat (*Avena sativa* L.). Biul. IHAR 247: 73–79. (in Polish)
3. Kraska P, Mielniczuk E. 2012. The occurrence of fungi on the stem base and roots of spring wheat (*Triticum aestivum* L.) grown in monoculture depending on tillage systems and catch crops. Acta Agrobot. 65 (1): 79–90.
4. Kurowski TP, Marks M, Orzech K, Kowalska E. 2008. Stan sanitarny i plonowanie pszenicy ozimej w zależności od sposobu uprawy roli. / Sanitary state and yielding of winter wheat as dependent on soil tillage system. Zesz. Prob. Post. Nauk Rol. 531: 95–103. (in Polish)
5. Lemańczyk G, Sadowski Cz. 2000. The effect of different forecrops on the occurrence of *Fusarium* spp. in winter wheat rhizosphere. Phytopathol. Pol. 20: 131–138.
6. Adamiak J, Adamiak E. 1999. Plonotwórcza i plonochronna rola owsa w płodozmianach zbożowych. / Yield-forming and crop protecting role of oats in cereal crop rotations. Pamiętnik Puławski – Mat. Konf. 114: 16–21. (in Polish)
7. Kiecana I, Mielniczuk E. 2001. Występowanie *Fusarium culmorum* (W.G.Sm.) Sacc., *Fusarium avenaceum* (Fr.) Sacc. oraz *Fusarium crookwellense* Burgess, Nelson & Toussoun na rodach hodowlanych owsa (*Avena sativa* L.). / The occurrence of *Fusarium culmorum* (W.G.Sm.) Sacc., *Fusarium avenaceum* (Fr.) Sacc. and *Fusarium crookwellense* Burgess, Nelson & Toussoun on oats lines (*Avena sativa* L.). Acta Agrobot. 64, 1: 83–93. (in Polish)
8. Cook RJ. 1981. *Fusarium* diseases of wheat and other small grains in North America. [In:] Nelson P.E., Toussoun T.A. and Cook R.J. (ed.), Diseases, Biology and Taxonomy. The Pennsylvania State University Park and London: 39–52.
9. Łacicowa B, Pięta D. 1998. Wpływ temperatury i opadów na udział grzybów w powodowaniu chorób podsuszkowych jęczmienia jarego (*Hordeum vulgare* L.). / The

- effect of temperature and rainfall on the contribution of fungi to causing root and stem rot diseases in spring barley (*Hordeum vulgare* L.). Acta Agrobot. 51 (1–2): 51–61. (in Polish)
10. Asran MR, Eraky Amal MI. 2011. Aggressiveness of certain *Fusarium graminearum* isolates on wheat seedlings and relation with their trichothecene production. Plant. Pathol. J. 10 (1): 36–41. <http://dx.doi.org/10.39.23/ppi.2011.36.41>
 11. Bacon ChW, Hinton DM. 2007. Potential for control of seedling blight of wheat caused by *Fusarium graminearum* and related species using the bacterial endophyte *Bacillus mojavensis*. Biocontrol Science and Technology 17 (1/2): 81–94. <http://dx.doi.org/10.1080/09583150600937006>
 12. Bateman GL, Gutteridge RJ, Gherbawy Y, Thomsett MA, Nicholson P. 2007. Infection of stem bases and grains of winter wheat by *Fusarium culmorum* and *F. graminearum* and effects of tillage method and maize-stalk residues. Plant Pathology 56: 604–615. <http://dx.doi.org/10.1111/J.1365-3059.2007.01577.x>
 13. Kiecana I, Rachoń L, Mielniczuk E, Szumiło G. 2011. The occurrence of fungi on roots and stem bases of common wheat (*Triticum aestivum* ssp. *vulgare* L.) and durum wheat (*Triticum durum* Desf.) grown under two levels of chemical protection. Acta Agrobot. 64 (3): 93–102.
 14. Gjaerum HB, Tjamos EC, Viranyi F. 1988. European Handbook of Plant Diseases. Smith I.M. (ed.). Blackwell Sci. Publ. Oxford London, Edinburgh, Boston, Palo Alto, Melbourne.
 15. Sutton JC. 1982. Epidemiology of wheat blight and maize ear rot caused by *Fusarium graminearum*. Can. J. Plant Pathol. 4: 195–209.
 16. Żuk B. 1989. Biometria stosowana. Państwowe Wydawnictwo Naukowe, Warszawa. (in Polish)
 17. Kwaśna H, Chełkowski J, Zajkowski P. 1991. Grzyby (*Mycota*) Tom XXII., Grzyby niedoskonałe (*Deuteromycetes*), Strzępczakowe (*Hyphomycetales*), Gruzelkowate (*Tuberculariaceae*), Sierpik (*Fusarium*). Inst. Botaniki PAN Kraków. (in Polish)
 18. Leslie JF, Summerell BA. 2006. The *Fusarium* Laboratory Manual. Blackwell Publishing.
 19. Mishra CBP, Behr L. 1976. Der Einfluss von Kulturfiltraten von *Fusarium culmorum* (W. G. Sm.) Sacc., *Fusarium avenaceum* (Fr.) Sacc. und *Fusarium nivale* (Fr.) Ces., *Griphosphaeria nivalis* Müller et v. Arx auf die Keimung des Weizen. Arch. Phytopathol. Pflanzenschutz 12, 6: 373–377. (in German)
 20. Mańka M. 1989. Patogeniczność wybranych gatunków z rodzaju *Fusarium* dla siewek zbóż. / Pathogenicity of selected species from the genus *Fusarium* to cereal seedlings. Roczn. AR Poznań, Rozp. Nauk. 201: 1–64. (in Polish)
 21. Kiecana I, Kocyłak E. 1999. Pathogenicity of *Fusarium* spp. on oats seedlings (*Avena sativa* L.). Plant Breed. Seed Sci. 43, 1: 91–99.
 22. Ellis MB. 1971. Dematiaceous Hyphomycetes. Commonwealth Mycological Institute, Kew Surrey, England.
 23. Gams W. 1971. *Cephalosporium* – artige Schimmelpilze (*Hyphomycetes*). Gustav Fischer Verlag, Stuttgart.
 24. Ramirez C. 1982. Manual and atlas of the Penicillia. Elsevier Biomedical Press., Oxford.
 25. Rifai MA. 1969. A revision of the genus *Trichoderma*. Kew, Surrey, England.
 26. Skirgiełło A, Zadara M, Ławrynowicz M. 1979. Grzyby (*Mycota*). Tom X. Glonowce (*Phycomycetales*), Pleśniakowe (*Mucorales*), Klebiantkowe (*Endogonales*). Warszawa-Kraków. (in Polish)
 27. Thom Ch, Raper KB. 1945. A manual of the Aspergilli Baltimore. The Williams & Wilkins Company.
 28. Kiecana I, Cegiełko M, Mielniczuk E. 2009b. Występowanie *Fusarium* spp. na życie ozimym (*Secale cereale* L.) i podatność różnych genotypów na porażenie przez *F. avenaceum* (Fr.) Sacc. i *F. culmorum* (W.G.Sm.) Sacc. / The occurrence of *Fusarium* spp. on winter rye (*Secale cereale* L.) and susceptibility of different genotypes to infection with *F. avenaceum* (Fr.) Sacc. and *F. culmorum* (W.G.Sm.) Sacc. Biul. IHAR. 252: 151–161. (in Polish)
 29. Strausbaugh CA, Bradley CA, Koehn AC, Forster RL. 2004. Survey of root of wheat and barley in southeastern Idaho. Can J Plant Pathol. 26: 167–176. <http://dx.doi.org/10.1080/07060660409507128>
 30. Eudes F, Zhou W, Badea A, Laroche A. 2004. Toward the development of *Fusarium* head blight resistance and reduced levels of mycotoxin in wheat and barley. Recent Res. Develop. Plant Pathol. 3: 17–49.
 31. Wiśniewska H, Buśko M. 2005. Evaluation of spring wheat resistance to *Fusarium* seedling blight. Biologia, Bratislava 60 (3): 287–293.
 32. Packa D. 1997. Cytogenetic effect of *Fusarium* mycotoxins on tip cells of rye (*Secale cereale* L.), wheat (*Triticum aestivum* L.) and field bean (*Vicia faba* L. Var. Minor). J. Appl. Genet. 38, (3): 259–272.
 33. Desjardins AE. 2006. *Fusarium* mycotoxins, chemistry, genetics, and biology. The American Phytopathological Society St. Paul, Minnesota, U.S.A.
 34. Kiecana I, Mielniczuk E, Cegiełko M. 2009a. Fungi colonizing grains of oat (*Avena sativa* L.) grown in the conditions of infection by *Fusarium* spp. In: “Seed – source of infection”. 4th International Seed Health Conference of Seed Pathology Section. Wrocław, Poland 7–9 September 2009. The Polish Phytopathological Society – Section of Seed Pathology, Wrocław University of Environmental and Life Sciences. Book of Abstracts. Wrocław: 25.
 35. Lemańczyk G, Wenda-Piesik A, Wasilewski P. 2001. Wpływ uprawy owsa w siewie czystym oraz w mieszkankach na jego zdrowotność i wartość przedplonową dla pszenicy ozimej. / Impact of the cultivation of oats in mixtures on the health status and yielding and evaluation of its forecrop value for winter wheat. Fragmenta Agronomica 4 (72): 65–77. (in Polish)
 36. Bryden WL, Logrieco A, Hamed K, Abbas HK, Porter JK, Vesonder RF, Richard JL, Cole RJ. 2002. Other significant *Fusarium* mycotoxins. [In:] Summerell B.A., Leslie J.F., Backhouse D., Bryden W.L., Bur-

- gess L.W. *Fusarium* Paul E. Nelson Memorial Symposium. APS Press The American Phytopathological Society St. Paul, Minnesota: 360–392.
37. Majchrzak B, Kurowski TP, Okorski A. 2008. Fungi isolated from the roots and stem bases of spring wheat grown after different cruciferous plants as forecrops. *Pol. J. Natur. Sc.* 23 (2): 299–309.
38. Truszkowska W, Chmurzyńska I, Czyrek A, Dorenda M. 1983. Zagadnienie zgorzeli podstawy źdźbła owsa (*Avena sativa* L.) w świetle doświadczeń agrotechnicznych. / Problem of foot rot in oats (*Avena sativa* L.) in the light of field crop production experiments. *Rocz. Nauk Rol. ser. E* 13, 1–2: 73–82. (in Polish)

**Występowanie *Fusarium* spp.
na owsie (*Avena sativa* L.) i podatność siewek
wybranych genotypów na porażenie
przez *Fusarium graminearum* Schwabe**

Streszczenie

Badania przeprowadzono w latach 2010–2012 na polach Hodowli Roślin Strzelce Sp. z o. o., Grupa IHAR, objęto nimi 39 genotypów owsa. W fazie 6-tygodniowych siewek oceniano udział roślin z objawami nekrozy korzeni oraz pochw liściowych. Udział sie-

wek z objawami chorobowymi wynosił w 2010 roku od 6,5% do 25%, w 2011 roku od 17% do 34,5%, zaś w 2012 roku od 10,0% do 25,0%. Wartości wskaźnika chorobowego wahały się w 2010 roku od 1,4 do 5,7, w 2011 roku od 4,5 do 8,8, zaś w 2012 roku od 2,0 do 5,4. Analiza mykologiczna wykazała, że zarówno z korzeni jak i z pochw liściowych licznie uzyskiwano kolonie *Fusarium* spp. Izolaty tych grzybów stanowiły 63,48% ogółu grzybów wyosobnionych z siewek. Przyczyną uszkodzenia siewek owsa uprawianego w warunkach centralnej Polski były gatunki: *F. culmorum*, *F. avenaceum* i *F. solani*. Badania podatności siewek 15 genotypów owsa na porażenie przez dwa szczepy *Fusarium graminearum* - Tz 56 i Tk 235 przeprowadzono w warunkach fitotronowych, w temperaturze 22–23°C i wilgotności względnej powietrza 85%. Szczep *F. graminearum* Tz 56 okazał się najbardziej patogeniczny w stosunku do siewek rodów hodowlanych STH 0.9403 oraz POB 1316/08, dla których wartości wskaźnika chorobowego wynosiły odpowiednio 80,5 oraz 75,5. Najmniejszą szkodliwość w/w szczepu zanotowano w przypadku genotypu DC 1832/05, dla którego wartość wskaźnika chorobowego wynosiła 26,5. Szczep *F. graminearum* Tk 235 okazał się najbardziej patogeniczny dla genotypów STH 0.9403 i STH 0.9423, dla których wartości wskaźnika chorobowego wynosiły odpowiednio: 70,5 i 70,0, zaś w/w szczep był najmniej patogeniczny dla rodu hodowlanego DC 2112/05, w przypadku którego wartość wskaźnika chorobowego wynosiła 25,5.

Handling Editor: Elżbieta Weryszko-Chmielewska

This is an Open Access digital version of the article distributed under the terms of the Creative Commons Attribution 3.0 License (creativecommons.org/licenses/by/3.0/), which permits redistribution, commercial and non-commercial, provided that the article is properly cited.

©The Author(s) 2014 Published by Polish Botanical Society