

FLORA OF TOXIC DEPOTS IN SELECTED INDUSTRIAL ZONES

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(Received: December 18, 2008. Accepted: July 21, 2009)

ABSTRACT

Floristic composition in three industrial areas with soils contaminated by heavy metals (As, Cd, Cu, Hg, Pb, Zn) and organic pollutants (polychlorinated biphenyls) was studied. The content of Pb was only significantly correlated with the floristic composition and explained 13.8% of its variability considering spatial dependency of the sites. No correlation was found for PCBs. Altogether, 237 plant vascular species were found at three study sites (117, 133 and 105, respectively). The three study areas differed in their species composition represented by their own characteristic species. The gradient in the content of natives/non-natives, species number, prevailing life forms and indicator values for plant species investigated was revealed. Based on our results, for phytoremediation purposes we can select productive plant species with high biomass and ability to accumulate large amounts of heavy metals or organic compounds and surviving on soils with low mineral content.

KEY WORDS: bioindication, Ellenberg indicator values, heavy metals, phytoremediation, polychlorinated biphenyls.

INTRODUCTION

Importance of environmental pollutants has been recognized internationally (see e.g. EC regulation No 1907/2006 Registration, Evaluation, Authorisation and Restriction of Chemical substances) and, based on precautionary principle, their occurrence is intensively monitored in all developed countries.

Nevertheless, many wastes and noxious residua (heavy metals and polychlorinated biphenyls PCBs, particularly) are buried in soil usually in the vicinity of industrial areas or within them as remnants of the previous companies' management, which had totally ignored basic principles of environment-friendly waste treatment (Rebele et al. 1993). Fortunately, after political changes in most post-communistic countries, the industry was structurally transformed, contributing less to environmental pollution and natural habitat degradation. In the Czech Republic, Ministry of the Environment has recently adopted methodology for environmental management accounting and companies ought to measure and monitor identified pollutant emissions following the operation permits of Integrated Prevention and Pollution Control (IPPC). Most industrial companies have

contingency plans and training for extraordinary situations that may occur – in temperate regions it is the leaching of toxics into water drainage and eolian erosion in arid regions are the most frequent (see Mendez and Maier 2008).

High concentrations of heavy metals or organic pollutants limit the colonization process or plant growth (see Kučerová et al. 2001) but it could be a selection factor for naturally stress-tolerant species. Based on knowledge of natural conditions with noxious substrate (e.g. arsenic in serpentine bedrock), we have the opportunity to find some natural biological accumulators with high bioavailability to heavy metals (so called hyperaccumulators, see Brey and Fabiszewski 2006) in order to restore sites degraded by human activity (e.g. mine tailings). Using accumulation and utilisation of the pollutants in plant tissues is a promising way for phytoremediation activities (see e.g. Adriano et al. 2004; Soudek et al. 2007). Thus, to help with environmental detoxication, the studies on vascular plant species indicating the harmful pollutants in the substrates polluted by heavy metals or PCBs from industrial activities are urgently needed.

The knowledge on plant cover could be useful both for phytoextraction (bioaccumulation) and for phytostabilization (immobilization of specific elements) (Mendez and

Maier 2008). Based on knowledge on species composition of naturally recovered toxic sites, we are able to select resistant species either to accumulate or stabilize toxic elements in a substrate. However, such selection should be experimentally tested utilizing modern approaches such as interactions with mycorrhiza and considering possible threats (e.g. potential invasiveness of the species if using non-invasive plants or gene flow when using genetically modified plants).

The goal of phytoremediation (plant-based remediation) processes is to eliminate the impact of contaminants on living organisms using plants to extract or to stabilize the poisonous conditions. Prior to the selection of appropriate plants, we have to find a correlation between toxic conditions (critical content of arsenic, copper, zinc, mercury, lead, cadmium and PCBs in soil) and the plant cover. This paper is therefore aimed at geobotanical survey description of two sites with deposits of heavy metals in the Czech Republic and another one in the Slovak Republic (containing old residua of PCBs).

METHODS

Study areas (for description see Fig. 1)

Kovohutě Příbram

The locality Kovohutě Příbram in the Czech Republic (49°42'26.514"N, 13°58'58.589"E) is situated in the area of Kovohutě Příbram nástupnická, a.s. The whole history of metallurgy in Příbram is inseparably connected with mining activities in this city. The mining and production of silver then subsequently accelerated the production of lead, using domestic lead concentrates until 1973, and thereafter the lead waste entirely. The lead wastes consist mainly of used vehicle accumulators, processed for raw lead, lead alloys and lead products. Three sampling and experimental areas with high concentrations of toxic metals were chosen on the basis of soil analysis.

Spolana Neratovice

The history of chemical factory in Neratovice, Czech Republic (50°15'50.053"N, 14°31'28.427"E) starts in 19th century. The first products were oil, stearine, soap and candles. For a few decades, the food production (chocolate,

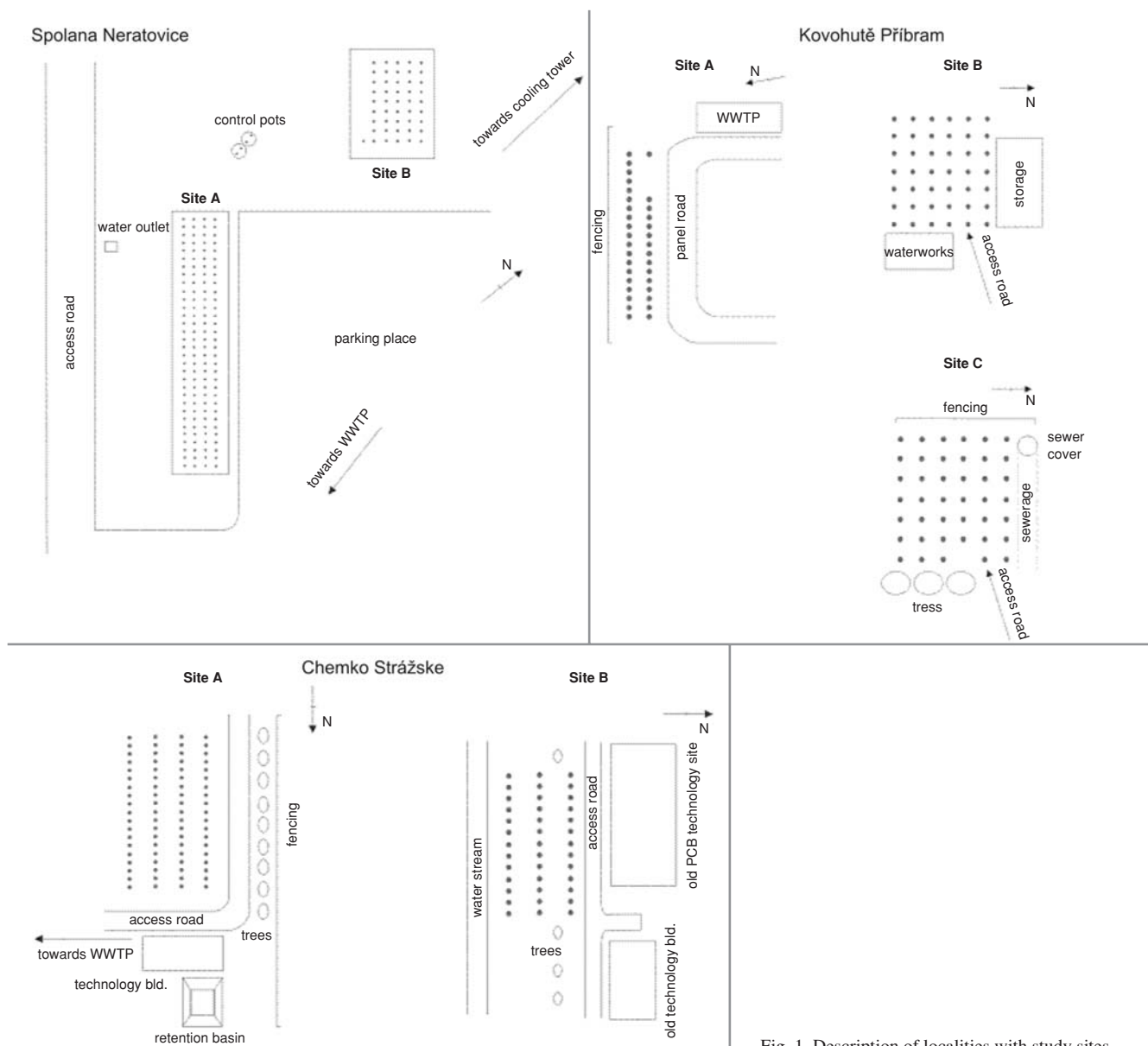
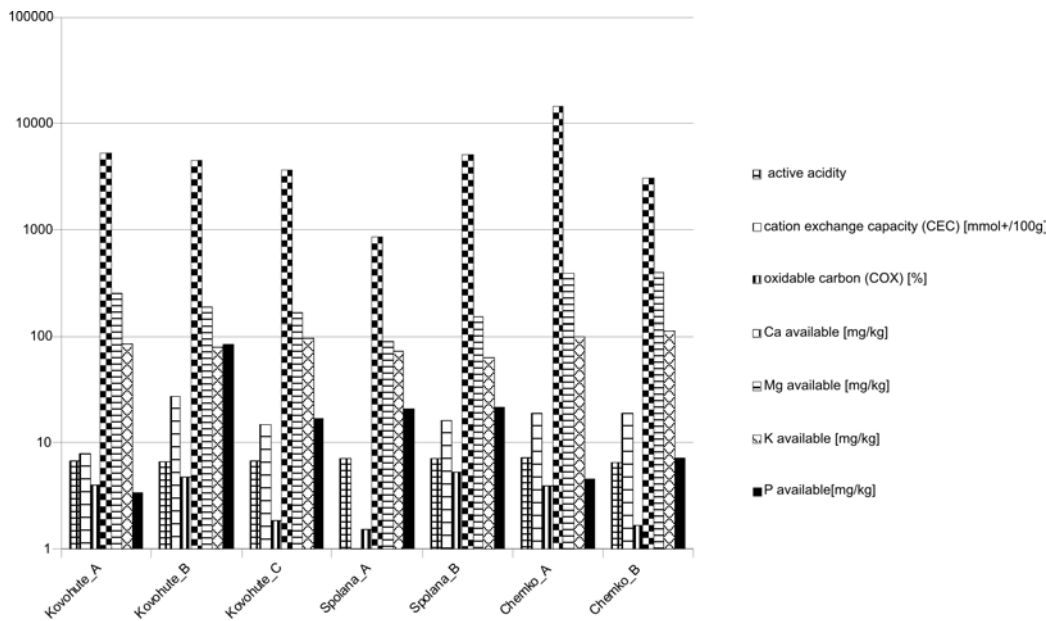


Fig. 1. Description of localities with study sites.



sweets, marmalade etc.) have been driven out the chemistry. In 1947, the production of viscose has been started and, since this time, chemical production in Spolana was gradually expanded to chlorine and sodium hydroxide by electrolysis, bone glue, fat, cellophane, hydrochloric acid, caprolactam and PVC.

Chemko Strážske

A chemical factory in East Slovakia near the village of Strážske (48°52'32.15"N, 21°48'56.57"E) has been in operation since 1952. In the past, the factory has been producing explosives and intermediates for military and civil purposes such as PCB. However, in the 90's, the situation in the market has changed and the economically unviable production of explosives have been suspended in 2003 and finally terminated on 1 January 2005. At present, the subsidiaries – Chemza, a.s. and Hnojivá, a.s. – produce industry-leading chemical products of inorganic and organic chemistry and their intermediates, nitric acid and nitration mixtures for the production of nitrogen fertilisers.

Soil sampling and chemical analyses

To assess general soil properties in study areas, available Ca, K, Mg, P in mg.kg⁻¹ of dry soil, cation exchange capacity in mmol/100 g of dry biomass, pH value in water solution, and percentage of oxidable carbon were determined from seven samples (see Appendix 1). To describe substrate toxicity in detail, 22 soil samples were further taken in Kovohutě and Spolana and contents of heavy metals (As, Cd, Cu – for Kovohutě only, Hg, Pb, Zn) in mg.kg⁻¹ of dry soil were measured (see Appendix 2). The content of polychlorinated biphenyls PCBs was measured for Chemko Strážské only.

To determine the basic soil properties, the samples were analyzed in a commercial laboratory. Analyses were carried out according to standards ISO 14235 (organic carbon), ISO 13536 (CEC), ISO 11277 (soil granularity), ČSN ISO 10390 (exchangeable pH) and public notice of the Czech Ministry of Agriculture Nr. 275/1998 Coll. (determination of available nutrients).

The dried soils were digested in 5 cm³ of acid mixture of HClO₄ and HNO₃ (15/85% v/v) in digestion glass tubes

overnight. Digestion was completed by a gradual increase of temperature from 60 to 195°C. Digestion protocol: 3 h (60°C); 1 h (100°C); 1 h (120°C); 3 h (195°C). After cooling, HCl (20%), 2.5 cm³ was added, whirl mixed and warmed to 80°C for one hour. The final volume was brought between 5 to 10 cm³ accurately. The heavy metal content was measured by AAS (SensAA, GBC, Australia) (Barazani et al. 2004).

PCB soil samples were extracted by n-hexane, polar phase was separated and polar compounds were removed by silicagel. Aliquot amounts of samples were analyzed on a Hewlett-Packard 5890 gas chromatograph with an electron capture detector and a fused silica capillary column (30 m, 0.32 mm inner diameter) coated with 0.25 µm immobilized phase of methyl-polysiloxane (5% phenyl) and nitrogen as a carrier gas (at the constant pressure 9 psi). The temperature program was 70°C for 0.5 min, then 25°C.min⁻¹ until 200°C, 10°C.min⁻¹ until 260°C, then isothermally 3.7 min, 25°C.min⁻¹ until 300°C, then isothermally 1 min. The sample amount injected was 1 µl in splitless mode. All of the seven congeners were identified by comparing their retention times with the retention times of corresponding peaks of PCB Congener Standard – RESTEK.

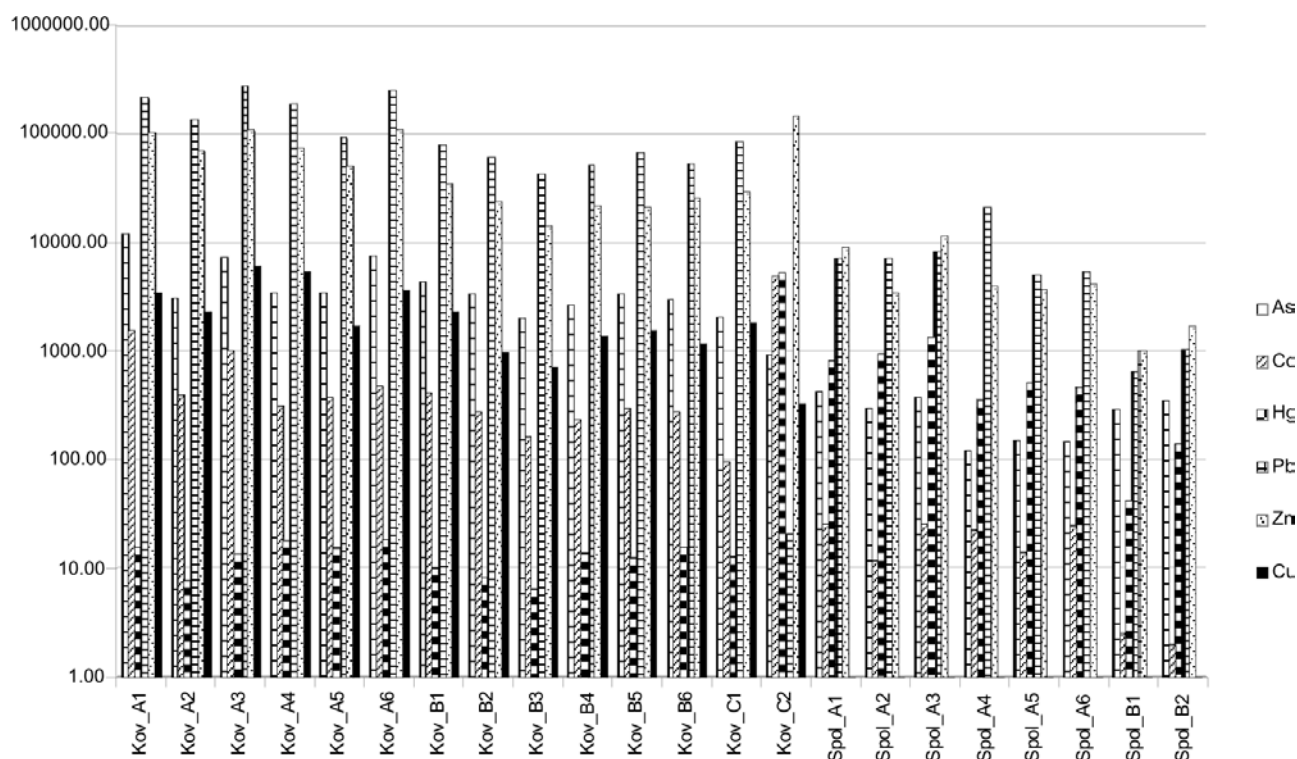
Botanical field survey

Within each locality, all vascular plant species were recorded in 6-14 samples of 2×2 m (the uneven number of samples was due to the varying sizes of each study area). The data on species composition were sampled twice in vegetation period (late summer 2007 and spring 2008) in order to cover important plant phenophases, an additional sampling for whole areas was performed so the final number of plots for classification reached 36.

Species data adjustment

To substitute missing information on selected soil properties, Ellenberg indicator values (Ellenberg et al. 1992) were used. These values are proxies for evaluation of site conditions for light, nutrients (productivity), continentality, temperature, soil reaction and moisture.

To employ species functional groups, their life forms according to C. Raunkjær (i.e. hemicryptophytes were classi-



Appendix 2. Content of six heavy metals from two study sites (Kovohutě and Spolana) investigated at 22 sample plots. The data are presented as mg.kg^{-1} of dry biomass*10 and shown in the logarithmic scale. Copper content was not measured at Spolana site.

fied – species with germinating buds above or at the soil ground – usually rhizomatous or tussock species, phanerophytes – trees and shrubs, terophytes – annuals surviving the unfavourable period in seeds, geophytes – species with vegetative organs in ground).

To assess original distribution areas of the species they were further classified as natives and non-natives (including neophytes, i.e. those brought to Europe after 1500 and archeophytes as those introduced before this date) according to Pyšek et al. (2002), and species number per plot was added.

Nomenclature of plant taxa follows Kubát et al. (2002) and all data adjustment was performed in the JUICE software (Tichý 2002).

Statistical analysis

To classify all 36 systematically sampled presence/absence data, a divisive classification TWINSPAN (Hill 1979) for all study areas was performed in the JUICE software (Tichý 2002). Diagnostic species typical of each study area were selected based on statistical tendency (measured by phi coefficient) of co-occurring species, which are more frequent than would be expected randomly (Bruehlheide 2000; see Petřík and Bruehlheide 2006 for using of species floristic data).

To discern trends in the composition of species and passive variables (i.e. indicator values, species number, native status, and life form) at all three study sites, the unconstrained linear ordination Principal Component Analysis (PCA) in CANOCO for Windows 4.0 (Ter Braak and Šmilauer 2002) was launched. For the assessment of species composition-environment correlations (i.e. the data set with environmental variables of soil properties and those containing species occurrences), a linear ordination method Redundancy Detrended Analysis (RDA) was performed for two sites with complete soil data as the length of gradients were

too short. To filter out the spatial dependency of the data, the localities as covariables were used. To find the minimum number of statistically significant environmental variables, a manual forward selection procedure with 999 Monte Carlo significance permutation tests was used. The eigenvalues and percentages of floristic variance explained by all canonical axes were then calculated.

RESULTS

Soil samples

The basic chemical soil properties were comparable in all study areas (Appendix 1). The lowest values of oxidable carbon was found at all plots (namely plot C in Kovohutě, plot A in Spolana and plot B in Chemko) followed by lower cation exchange capacity and bases (except of Mg and P where the values varied independently of other bases) in humus layer. The content of two base cations was significantly different from the other at Chemko site (for Mg the F value = 27.46, $P = 0.0046$, for K the F value is 11.225, $P = 0.0229$).

All sites differed significantly in content of As ($F = 14.395$, $P = 0.003$), Pb ($F = 12.244$, $P = 0.0023$) and Zn ($F = 12.777$, $P = 0.0019$). Compared to Spolana sites, Kovohutě was rich in all heavy metals investigated, with only one exception (plot C2) with low lead content (see Appendix 2). The arsenic reached up to 1180 ppm and, cadmium, up to 479 ppm was measured in one soil sample. The only exception was mercury (510 ppm), cadmium (479 ppm) and zinc (14380 ppm) at plot number C2 with extraordinary high values. Interestingly, this plot had the lowest amount of lead residua from all sites.

PCBs content was recognized only in Chemko and its values differ up to ten times between plots within this study area. Values of the PCBs in mg.kg^{-1} of dry soil are as

follows: 0.027, 0.046, 0.058, 0.063, 0.077, 0.11, 0.17, 0.18 at the first plot and 0.38, 0.43 at the second plot.

Species composition at study sites

Totally, 237 plant vascular species were found in three study areas (117 in Chemko, 133 in Kovohutě and 105 in Spolana). The three study areas differed in their species composition and every site was well represented by its own characteristic species (see Table 1). In Chemko site the species were typical of vegetation of moist meadows and pastures, whereas in Kovohutě the vegetation was similar to dry and mesic meadows with many ruderal plants. Quite unique vegetation has been developed in Spolana where under regular disturbances many annual and alien species occur (see further).

The results of PCA (Fig. 2) show gradients in the occurrence of natives/non-natives (including neophytes and archeophytes), species number, prevailing life forms and indicator values for plant species investigated. There were logically large differences among species composition of sites studied rather than the differences in their internal variability. The only exception was vegetation of Kovohutě of which the composition was most diverse and could be classified into two groups alongside the second PCA axis due to the significant gradient in productivity (or nutrient) based on indicator values and prevailing life form of plants. The position of Kovohutě vegetation in ordination space was well associated with high amount of natives and hemicryptophytes/phanerophytes (both are intercorrelated variables) and high supply for moisture. On the opposite gradient wing there is the relatively species-rich and unified vegetation of Spolana with many neophytes/archeophytes, with life-form of terophytes, with higher demand to light/continentality/temperature (mutually correlated variable) and soil reaction. The central position in ordination space was occupied by Slovak Chemko plots, represented by a relatively higher occurrence of geophytes.

Correlation of species composition with soil properties

Considering the sites as covariables, from five heavy metals analysed only the content of Pb was shown to be significant for vegetation composition of the two sites compared (see Fig. 3) and totally explained 13.8% of total variance in species data at the significance level $P = 0.002$ ($F = 3.04$), and was highly positively correlated (inter set correlation coefficient = 0.81) with the first canonical axis.

DISCUSSION

Soil properties and species composition correlations

The basic soil properties are apparently similar at all study sites and could be characterised as neutral up to slightly alcalic soil with slightly to moderately humose soil with high content of bases and varying cation exchange capacity. As the soil conditions are similar (except of two basic cations content), no specific plants were found to be correlated with these properties.

There were significant group of species recognised to be bound with higher content of Pb ions (see Fig. 2). To prove these species to be tolerant to high heavy metal content, habitats with similar conditions with ore-washery or ash-slag deposits in the Czech Republic could be compared

TABLE 1. Species composition at three study sites (column 1 – Chemko, column 2 – Kovohutě, 3 – Spolana) classified by TWINSpan divisive method and sorted according the decreasing fidelity based on phi coefficient $\phi \geq 0.50$ (**). Species characteristic for each column are ranked by a decreasing fidelity. Phi coefficient standardized to all group size of the total data set was used. Diagnostic species with probability of observed occurrence concentration in the given column (group of samples) higher than $P < 0.01$ (Fisher's exact test) were excluded. Number of sample plots for every study area is listed with species frequency in each column.

Study area no. No. of relevés	1 9	2 17	3 10
<i>Acer negundo</i>	78**	.	.
<i>Pastinaca sativa</i>	78**	.	.
<i>Galium album</i>	89**	18	.
<i>Dactylis glomerata</i>	89**	12	20
<i>Symphytum officinale</i>	56**	.	.
<i>Potentilla anserina</i>	56**	.	.
<i>Erigeron annuus</i>	100**	.	80
<i>Silene vulgaris</i>	.	88**	.
<i>Festuca rubra</i>	22	82**	.
<i>Holcus lanatus</i>	.	59**	.
<i>Rumex acetosa</i>	33	88**	.
<i>Cerastium holosteoides</i>	44	94**	.
<i>Capsella bursa-pastoris</i>	.	76**	20
<i>Hemiaria glabra</i>	.	53**	.
<i>Vicia cracca</i>	.	53**	.
<i>Trifolium repens</i>	11	65**	.
<i>Populus tremula</i>	11	65**	10
<i>Senecio vulgaris</i>	.	.	100**
<i>Papaver dubium</i> agg.	.	6	80**
<i>Crepis foetida</i> ssp. <i>rheoadifolia</i>	.	.	70**
<i>Arenaria serpyllifolia</i>	.	.	60**
<i>Datura stramonium</i>	.	.	60**
<i>Lactuca serriola</i>	11	.	70**
<i>Viola arvensis</i>	.	12	70**
<i>Fallopia convolvulus</i>	11	6	70**
<i>Digitaria ischaemum</i>	.	.	50**
<i>Senecio jacobaea</i>	22	.	70**
<i>Achillea millefolium</i>	100	94	100
<i>Taraxacum</i> sect. <i>Ruderalia</i>	89	88	100
<i>Daucus carota</i>	78	59	90
<i>Arrhenatherum elatius</i>	89	53	40
<i>Poa pratensis</i>	67	53	60
<i>Tanacetum vulgare</i>	78	35	70
<i>Poa compressa</i>	67	29	90
<i>Poa palustris</i>	78	35	50
<i>Calamagrostis epigejos</i>	67	18	90
<i>Medicago lupulina</i>	67	29	70
<i>Cirsium arvense</i>	89	6	80
<i>Tripleurospermum inodorum</i>	11	53	70
<i>Urtica dioica</i>	89	47	.
<i>Artemisia vulgaris</i>	44	41	50
<i>Plantago major</i> agg.	56	53	.
<i>Elytrigia repens</i>	78	6	50
<i>Plantago lanceolata</i>	67	18	40
<i>Veronica arvensis</i>	.	41	60
<i>Equisetum arvense</i>	56	35	10
<i>Arabidopsis thaliana</i>	.	59	20
<i>Ranunculus repens</i>	78	24	.
<i>Trifolium pratense</i>	33	47	.
<i>Securigera varia</i>	.	18	70
<i>Carex hirta</i>	56	18	10
<i>Lolium perenne</i>	56	18	10
<i>Agrostis capillaris</i>	22	41	.
<i>Hypericum perforatum</i>	22	12	50
<i>Silene latifolia</i>	44	24	10
<i>Fraxinus excelsior</i>	11	41	10
<i>Verbascum thapsus</i>	.	41	20
<i>Echium vulgare</i>	.	29	40
<i>Ranunculus acris</i>	11	41	.

TABLE 1. Cont.

Study area no.	1	2	3
No. of relevés	9	17	10
<i>Chenopodium album</i> agg.	.	12	60
<i>Stellaria media</i>	.	29	30
<i>Erodium cicutarium</i>	.	12	60
<i>Glechoma hederacea</i>	22	29	.
<i>Tussilago farfara</i>	33	24	.
<i>Rumex crispus</i>	56	.	20
<i>Lotus corniculatus</i>	22	18	20
<i>Lamium album</i>	22	24	10
<i>Aegopodium podagraria</i>	.	41	.
<i>Heracleum sphondylium</i>	.	41	.
<i>Setaria pumila</i>	56	.	10
<i>Arctium minus</i>	33	12	10
<i>Rumex obtusifolius</i>	11	29	.
<i>Rubus sect. Corylifolii</i>	11	24	10
<i>Sonchus oleraceus</i>	11	.	50
<i>Oenothera species</i>	11	.	50
<i>Salix caprea</i>	.	35	.
<i>Verbascum nigrum</i>	.	6	50
<i>Myosotis stricta</i>	.	12	40
<i>Leontodon autumnalis</i>	44	6	.
<i>Galium aparine</i>	22	.	30
<i>Calystegia sepium</i>	44	6	.
<i>Conyza canadensis</i>	22	.	30
<i>Trifolium campestre</i>	11	6	30
<i>Betula pendula</i>	.	29	.
<i>Poa nemoralis</i>	.	29	.
<i>Pinus sylvestris</i>	.	29	.
<i>Knautia arvensis</i>	.	29	.
<i>Campanula patula</i>	.	12	30
<i>Lythrum salicaria</i>	44	.	.
<i>Mentha longifolia</i>	44	.	.
<i>Rorippa sylvestris</i>	44	.	.
<i>Picris hieracioides</i>	44	.	.
<i>Geum urbanum</i>	44	.	.
<i>Potentilla argentea</i>	22	6	10
<i>Cirsium vulgare</i>	11	.	30
<i>Rubus mollis</i>	44	.	.
<i>Chenopodium polyspermum</i>	11	18	.
<i>Crepis biennis</i>	22	12	.
<i>Senecio viscosus</i>	11	.	30
<i>Myosotis arvensis</i>	11	18	.
<i>Convolvulus arvensis</i>	11	.	30
<i>Spergularia rubra</i>	.	24	.
<i>Poa annua</i>	.	24	.
<i>Moehringia trinervia</i>	.	24	.
<i>Rumex acetosella</i>	.	24	.
<i>Geranium pusillum</i>	.	18	10
<i>Carex ovalis</i>	.	24	.
<i>Vicia tetrasperma</i>	.	24	.
<i>Lamium amplexicaule</i>	.	.	40
<i>Cerastium semidecandrum</i>	.	.	40
<i>Solanum nigrum</i>	.	.	40
<i>Linum perenne</i>	.	.	40
<i>Ajuga reptans</i>	33	.	.
<i>Echinochloa crus-galli</i>	11	.	20
<i>Lysimachia nummularia</i>	33	.	.
<i>Carpinus betulus</i>	33	.	.
<i>Quercus robur</i>	11	12	.
<i>Agrostis stolonifera</i>	33	.	.
<i>Persicaria maculosa</i>	11	.	20
<i>Leucanthemum vulgare</i> agg.	.	18	.
<i>Trifolium arvense</i>	.	18	.
<i>Picea abies</i>	.	18	.
<i>Hieracium sabaudum</i>	.	18	.
<i>Vicia sepium</i>	.	18	.
<i>Cirsium palustre</i>	.	18	.
<i>Viola tricolor</i>	.	18	.

TABLE 1. Cont.

Study area no.	1	2	3
No. of relevés	9	17	10
<i>Cardaminopsis halleri</i>	.	18	.
<i>Vicia hirsuta</i>	.	12	10
<i>Sisymbrium altissimum</i>	.	.	30
<i>Bromus sterilis</i>	.	.	30
<i>Polygonum aviculare</i>	.	.	30
<i>Cerastium glutinosum</i>	.	.	30
<i>Papaver argemone</i>	.	.	30
<i>Deschampsia cespitosa</i>	22	.	.
<i>Cichorium intybus</i>	22	.	.
<i>Bidens frondosa</i>	22	.	.
<i>Prunus avium</i>	22	.	.
<i>Anagallis arvensis</i>	22	.	.
<i>Epilobium</i> sp.	22	.	.
<i>Rumex confertus</i>	22	.	.
<i>Veronica persica</i>	11	6	.
<i>Amaranthus</i> sp.	11	.	10
<i>Rosa canina</i> s.l.	11	.	10
<i>Malus domestica</i>	22	.	.
<i>Epilobium lamyi</i>	22	.	.
<i>Acer pseudoplatanus</i>	22	.	.
<i>Humulus lupulus</i>	11	.	10
<i>Melilotus albus</i>	11	.	10
<i>Oxalis fontana</i>	22	.	.
<i>Myosoton aquaticum</i>	22	.	.
<i>Arctium lappa</i>	11	.	10
<i>Salix purpurea</i>	.	12	.
<i>Atriplex patula</i>	.	12	.
<i>Erysimum cheiranthoides</i>	.	12	.
<i>Trifolium hybridum</i>	.	12	.
<i>Hypochaeris radicata</i>	.	6	10
<i>Linaria vulgaris</i>	.	12	.
<i>Veronica chamaedrys</i>	.	12	.
<i>Malva neglecta</i>	.	12	.
<i>Polygonum arenastrum</i>	.	6	10
<i>Salix fragilis</i>	.	12	.
<i>Erysimum durum</i>	.	12	.
<i>Campanula rapunculoides</i>	.	12	.
<i>Trifolium dubium</i>	.	12	.
<i>Reseda lutea</i>	.	.	20
<i>Vicia sativa</i>	.	.	20
<i>Papaver rhoeas</i>	.	.	20
<i>Arctium</i> sp.	.	.	20
<i>Rumex thyrsoiflorus</i>	.	.	20
<i>Thlaspi arvense</i>	.	.	20
<i>Oenothera biennis</i>	.	.	20
<i>Bromus tectorum</i>	.	.	20
<i>Nepeta cataria</i>	.	.	20
<i>Rubus caesius</i>	.	.	20
<i>Eragrostis minor</i>	.	.	20
<i>Amaranthus albus</i>	.	.	20

Species presented only once:

In column 1: *Potentilla reptans*; *Sonchus arvensis*; *Persicaria amphibia*; *Barbarea vulgaris*; *Plantago media*; *Inula britannica*; *Lathyrus pratensis*; *Euonymus europaea*; *Robinia pseudacacia*; *Solidago canadensis*; *Clematis vitalba*; *Comus sericea*; *Eupatorium cannabinum*; *Comus sanguinea*; *Agrostis gigantea*; *Lathyrus tuberosus*; *Viola odorata*; *Festuca pratensis*; *Prunella vulgaris*; *Clinopodium vulgare*; *Prunus padus*;

In column 2: *Carex* sp.; *Hieracium* species; *Persicaria lapathifolia*; *Setaria viridis*; *Larix decidua*; *Salix alba*; *Microrrhinum minus*; *Crepis capillaris*; *Festuca arundinacea*; *Epilobium montanum*; *Erigeron acris* s.l.; *Sedum sexangulare*; *Phleum pratense*; *Galinsoga quadriradiata*; *Artemisia absinthium*; *Euphorbia helioscopia*; *Sinapis arvensis*; *Carex muricata* agg.; *Sonchus asper*; *Carex brioidea*; *Trisetum flavescens*; *Pimpinella saxifraga*; *Torilis japonica*; *Sedum acre*; *Stellaria graminea*; *Astragalus glycyphyllos*; *Odontites vernus*;

In column 3: *Tragopogon pratensis*; *T. dubius*; *Vulpia myuros*; *Holosteum umbellatum*; *Centaurium erythraea*; *Elymus caninus*; *Euphorbia esula*; *Vicia villosa*; *Chenopodium botrys*; *C. pedunculare*; *Amaranthus retroflexus*; *Digitaria sanguinalis*

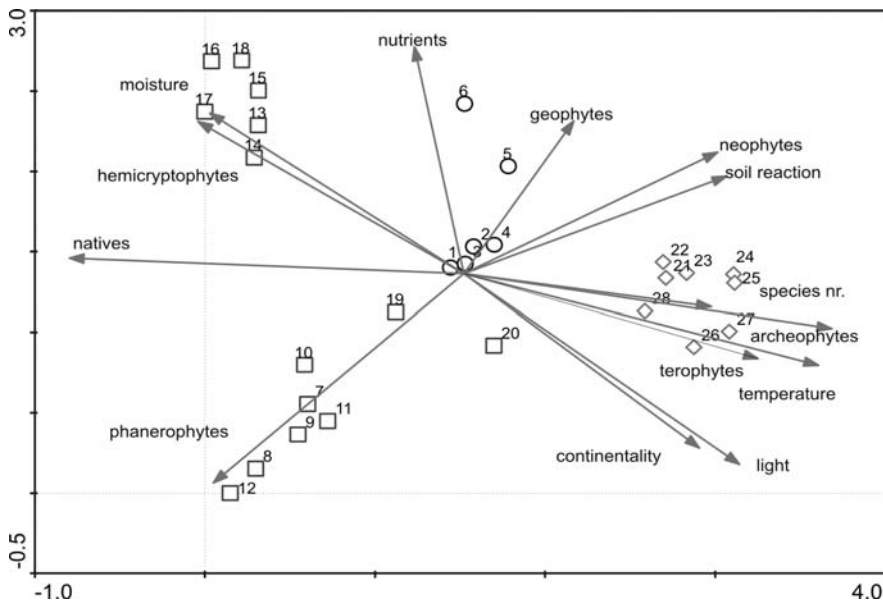


Fig. 2. Results of linear ordination method Principal Component Analysis (PCA) processed in CANOCO program and based on 28 samples from three study sites depicting pattern in their species composition. The Ellenberg indicator values (for light, nutrients, continentality, temperature, soil reaction, moisture), life form according to C. Raunkjær (hemicryptophytes, phanerophytes, terophytes, geophytes), species number per each plot, and native status according to Pyšek et al. (2002) were projected into ordination space as passive variables.

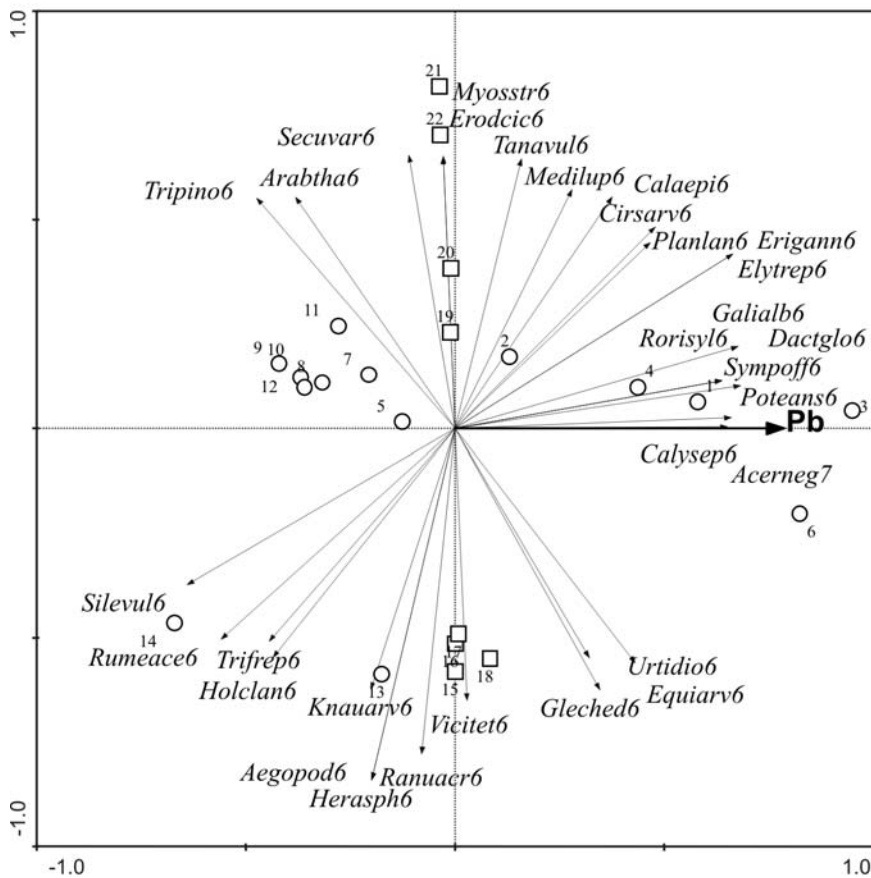


Fig. 3. Results of linear ordination method Redundancy Detrended Analysis (RDA) processed in CANOCO program with forward selection of variables (999 permutations tested by Monte Carlo test). Two study sites are shown (Kovohutě – circles, Spolana – squares) with arrow showing correlation of species composition with content of Pb, a heavy metal, which passed through selection. Only 31 species best fitting with variables are shown. Abbreviations (6 means herbs, 7 means juveniles of tree): Acerneg7 – *Acer negundo*, Calaepi6 – *Calamagrostis epigejos*, Čirsarv6 – *Cirsium arvense*, Dactglo6 – *Dactylis glomerata*, Elytrep6 – *Elytrigia repens*, Equiarv6 – *Equisetum arvense*, Erigann6 – *Erigeron annuus*, Gleched6 – *Glechoma hederacea*, Medilup6 – *Medicago lupulina*, Planlan6 – *Plantago lanceolata*, Poteans6 – *Potentilla anserina*, Rorisyl6 – *Rorripa sylvestris*, Sympoff6 – *Symphytum officinale*, Tanavul6 – *Tanacetum vulgare*, Urtidio6 – *Urtica dioica*, Galialb6 – *Galium album*, Calysep6 – *Calystegia sepium*, Rumeace6 – *Rumex acetosa*, Ranuacr6 – *Ranunculus acris*, Arabtha6 – *Arabidopsis thaliana*, Holclan6 – *Holcus lanatus*, Silevul6 – *Silene vulgaris*, Tripino6 – *Tripleurospermum inodorum*, Trifrep6 – *Trifolium repens*, Secuvar6 – *Securigera varia*, Aegopod6 – *Aegopodium podagraria*, Herasph6 – *Heracleum sphondylium*, Knauarv6 – *Knautia arvensis*, Vicitet6 – *Vicia tetrasperma*, Myosstr6 – *Myosotis stricta*, Erodci6 – *Erodium cicutarium*.

(see review done by Vaňková and Kovář 2004). Preliminary analysis shows that the species composition differs profoundly between these areas, and is characterized by only common species with wide ecological niches, which are able to colonize the toxic substrate.

The content of other heavy metals was not shown to be significantly associated with species composition. This is maybe due to the low number of sites. Similarly, no correlation between PCBs and plants was found, but there is lack of studies dealing with similar topic.

The variation in species composition explained by content of heavy metals in soil is not the only factor, and other unknown factors could be important for species occurrence in the sites (i.e. management – regular mowing, soil seed bank, species pool of neighbouring area) and without further experiments we cannot disentangle their effect, at all. For example, the enriching effect of neighbouring landscape for species diversity of species-poorer sedimentation basins was proved by Vaňková and Kovář (2004).

Possible remedial measures for sites

The soil substrate is suitable for vegetation and is colonized by various plant species, apart of high toxic contents. Thus, no action is needed to reduce the initial toxicity of the sites studied. The obtained results may give us the possibility to select plant species for phytoremediation purposes. The plants suitable for phytoremediation should have not only a high ability to accumulate large amount of heavy metals or organic compounds, but also must be able to survive on soils with low mineral content and must produce a relatively high biomass (Cheng 2003a, b).

Since purification capacity of plants is species-specific and depends also on soil properties, we recommend studying the effects of nutrient or biosolids amendments at the sites studied (usually, the concentrations of trace elements and bioavailability by plants increase after fertilizing, see Adriano et al. 2004; Wu et al. 2004; Zahra et al. 2008). In addition, the role of mycorrhiza could play a role and enhance plant growth at toxic substrates.

CONCLUSIONS

Phytoremediation is relatively cost-effective compared to total removal of contaminated soil. However, when selecting hyper-accumulators, we must consider several factors (hyper-accumulator biomass production, their toxicity for wildlife, distribution of contaminants in their roots and shoots, long length of cropping cycle, enhancement of biological processes in the initial substrate, accumulation factor – uptake of minerals by plants compared to soil content (see Baker et al. 2000). Currently, we are not able to recognize such plants as the analyses of leaves content are under analyses. However, based on many studies, the most suitable species for phytostabilization, the grasses (Poaceae family) or tussock species with developed root system such as *Calamagrostis epigejos* and *Elytrigia repens* or *Artemisia vulgaris* (Asteraceae) are recommended as good bioindicators in the study areas (see e.g. Rebele et al. 1993, Lehmann and Rebele 2004a, b). Such species fulfil most of the criteria mentioned above and they are very frequent in order to assess variability.

ACKNOWLEDGEMENT

This work was supported by project NPV II (2B06187), NPV II (2B08058) and by the institutional long-term research plan AV0Z60050516 (Academy of Sciences of the Czech Republic). P. P. was supported by the Biodiversity Research Centre (grant nr. LC 06073 funded by Ministry of Education, Youth and Sports). We thank the anonymous reviewer for valuable comments.

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