

## THE PARASITE FAUNA OF BROWN TROUT *SALMO TRUTTA*, WHITE FISH *COREGONUS LAVARETUS*, AND GRAYLING *THYMALLUS THYMALLUS* FROM WATER BODIES OF NORTH KARELIA (RUSSIA)

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**ABSTRACT.** The data on parasite fauna of brown trout *Salmo trutta*, whitefish *Coregonus lavaretus*, and grayling *Thymallus thymallus* from lake-river system Paanajarvi-Olanga (White Sea basin) are presented. As lake-river systems on this area remain in natural conditions, without any anthropogenic pressure, the parasite fauna of Salmonidae in these water bodies had not been changed and has an archetypal status.

**Key words:** *Coregonus lavaretus*, Karelia, parasite fauna, *Salmo trutta*, *Thymallus thymallus*.

### INTRODUCTION

Paanajarvi-Olanga system is situated at the North Karelia (Russia) in the territory of National Park Paanajarvi, which borders on Finland and Murmansk region. This lake-river system belongs to the Basin of the Kovda River – one of the largest rivers of the White Sea basin. Paanajarvi-Olanga is characterized by the presence of water bodies where Salmonidae found to be a basis of ichthyocenosis.

Being an oligotrophic lake, Paanajarvi is closer to the ultraoligotrophic status than other water bodies of Karelia. Therefore, Paanajarvi can be regarded as a model of a periglacial lake where the parasite fauna of Salmonids had formed.

### MATERIAL AND METHODS

The object of research were parr aged 3+ and adult fish of brown trout, white fish, grayling from Paanajarvi lake. The material was collected and treated following the generally accepted technique (Bykhovskaya-Pavlovskaya 1985). Fifty specimens from each locality were studied.

Describing the structure of parasite fauna we used two following terms: „core of parasite fauna” and „crossing sector of parasite fauna”. „The core of parasite fauna” is formed by the species presented in each host species under study. „Crossing sector of parasite fauna” is founded by the species, which occur in parasite fauna at least two hosts.

Two parameters of infection were analysed: prevalence and abundance (as defined by Margolis et al. 1982).

### RESULT AND DISCUSSION

Analysing the infection of brown trout parr, white fish, and grayling fry (Table 1) we can trace the first stages of formation of the salmonid parasite fauna.

Table 1. The parasite fauna of parr of Salmonidae fish: prevalence – (%)/abundance

| Species of parasite               | Brown trout | White fish | Grayling |
|-----------------------------------|-------------|------------|----------|
| <i>Hexamita truttae</i>           | –           | –          | 27/+     |
| <i>Chloromyxum schurovi</i>       | 13/+        | –          | –        |
| <i>Capriniana piscium</i>         | 47/0.07     | –          | –        |
| <i>Apiosoma campanulatum</i>      | –           | –          | 27/0.004 |
| <i>A. megamicronucleatum</i>      | 7/0.002     | –          | –        |
| <i>Trichodina pediculus</i>       | 7/0.003     | –          | –        |
| <i>Trichodinella epizootica</i>   | –           | 13/0.015   | 7/0.0006 |
| <i>Scyphidia sp.</i>              | –           | –          | 7/0.001  |
| <i>Tetraonchus borealis</i>       | –           | –          | 50/2.0   |
| <i>Discocotyle sagittata</i>      | –           | 67/7.0     | 7/0.1    |
| <i>Triaenophorus crassus</i>      | –           | –          | 7/0.3    |
| <i>Proteocephalus longicollis</i> | –           | 93/10.0    | –        |
| <i>Crepidostomum farionis</i>     | 20/0.5      | 13/0.2     | 27/0.7   |
| <i>C. metoecus</i>                | 7/0.1       | –          | –        |
| <i>Ichthyocotylurus erraticus</i> | 7/0.1       | 64/6.0     | 7/0.1    |
| <i>Diplostomum spathaceum</i>     | 7/0.06      | –          | 7/0.1    |
| <i>D. volvens</i>                 | 73/2.1      | 27/3.0     | 27/0.3   |
| <i>D. pseudobaueri</i>            | 7/0.1       | –          | –        |
| <i>Raphidascaris acus l.</i>      | –           | 13/1.5     | 60/1.8   |
| <i>Rhabdochona denudata</i>       | –           | –          | 7/0.1    |
| <i>Cystidicoloides tenuissima</i> | 67/2.0      | 7/0.1      | 100/75.0 |
| <i>Capillaria salvelini</i>       | 33/0.8      | –          | 7/0.1    |
| <i>Echinorinchus salmonis</i>     | 13/0.8      | –          | 20/0.3   |
| <i>Salmincola coregonorum</i>     | –           | 13/0.3     | –        |
| <i>S. extumenscen</i>             | –           | 13/0.1     | –        |
| <i>S. thymalli</i>                | –           | –          | 20/0.3   |
| No. of parasite species           | 13          | 10         | 17       |
| No. of fish studied               | 15          | 15         | 15       |

The core of their parasite fauna is formed by the following species: *Crepidostomum farionis*, *Ichthyocotylurus erraticus*, *Diplostomum volvens*, and *Cystidicoloides tenuissima* (Fig. 1)

It is necessary to take into consideration that these three salmonid fishes feed mostly on benthic organism. This fact determines a specific composition of the core including the species whose development goes on with the participation of bottom

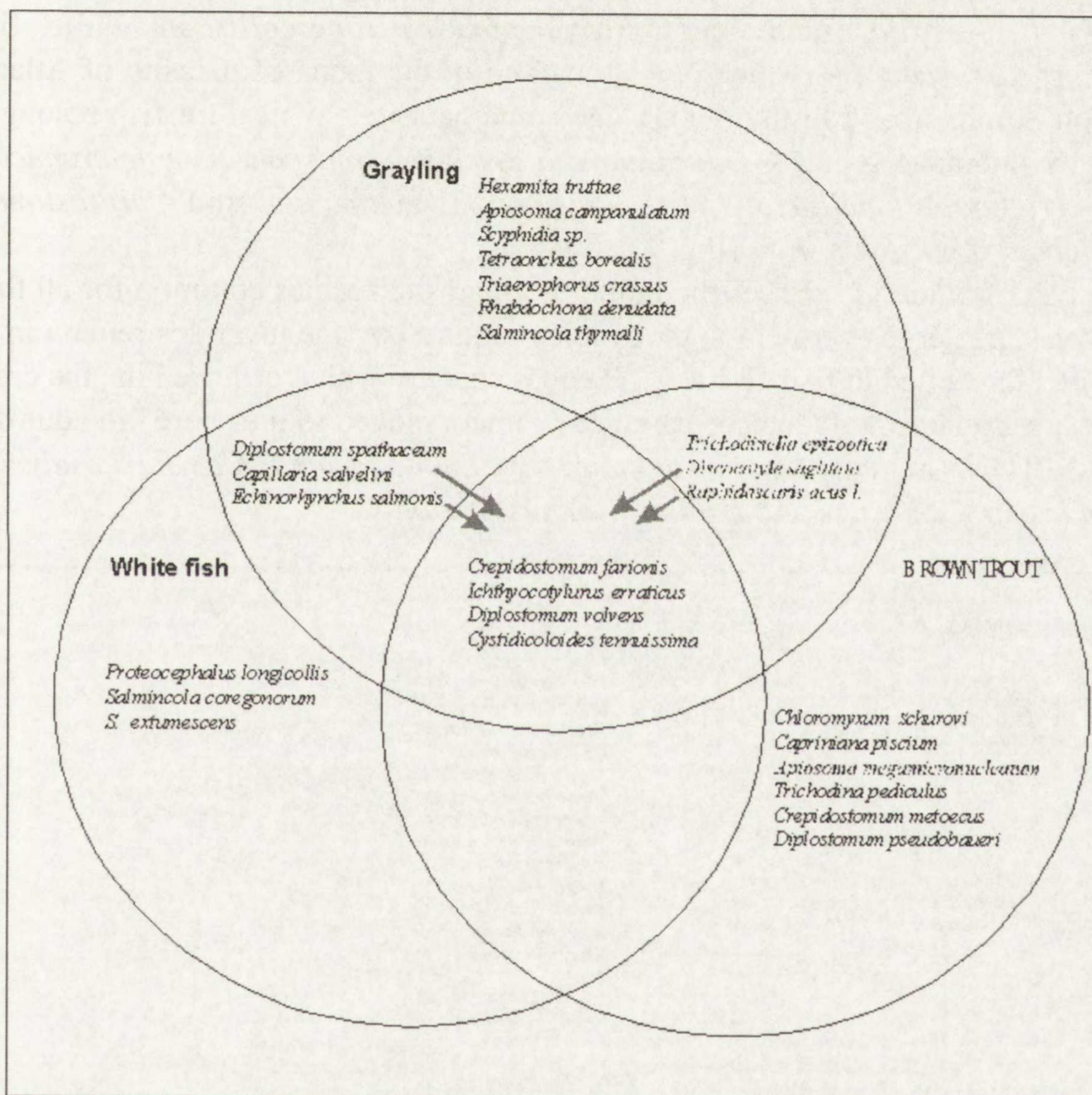


Fig. 1. Formation of Salmonidae parr parasite fauna from lake-river system Paanajrvi-Olanga

invertebrates. The species which are the basis of the parasite fauna of Salmonidae belong to boreal-foothills and arctic freshwater complexes. Only *D. volvens* is widely specific.

The species, observed in two hosts, are located in the crossing sectors of parasite fauna (Fig. 1). The parasites common for white fish and grayling are typical species for Salmonidae (*Capillaria salvelini*, *Echinorhynchus salmonis*) and non-specific digenean *D. spathaceum*.

Common for white fish and grayling parr were: the infusoria *Trichodinella epizootica*, the typical monogeneans of Salmonidae – *Discocotyle sagittata* and widely spread nematode *Rhabdiascaris acus*.

There are also some species, which were found in parr of only one host species, determining the specific elements of parasite fauna of each host under study. These are: *Hexamita truttae*, *Apiosoma campanulatum*, *Scyphidia sp.*, *Tetraonchus borealis*, *Rhabdochona demidata*, and *Salmincola thymalli* in grayling; *Salmincola core-*

gonorum, *S. extumescens*, and *Proteocephalus longicollis* in white fish; *Chloromyxum schurovi* (which is also marked in the fauna of parasite of Atlantic Salmon (Shulman and Ieshko 2003), and some parasites, typical for fish belonging to the Salmonidae group – *Capriniana piscium*, *Apiosoma megamicronucleatum*, *Trichodina pellucidus* *Crepidostomum metoecus*, and *Diplostomum pseudobaueri* in brown trout (Fig. 1).

In fish of older age classes the number of parasite species common for all three hosts was greater. Nevertheless, the species acquired by the juveniles remain in the parasite fauna of adult fish (Table 2). Besides species, which occurred in „the crossing sector of parasitic fauna” of parr, some others moved to the „core” in adult fish (Fig. 2). These are: *Discocotyle sagittata*, *Cystidicola farionis*, *Capillaria salvelini*, *Rhapidascaaris acus* (l.), and *Echinorhynchus salmonis*.

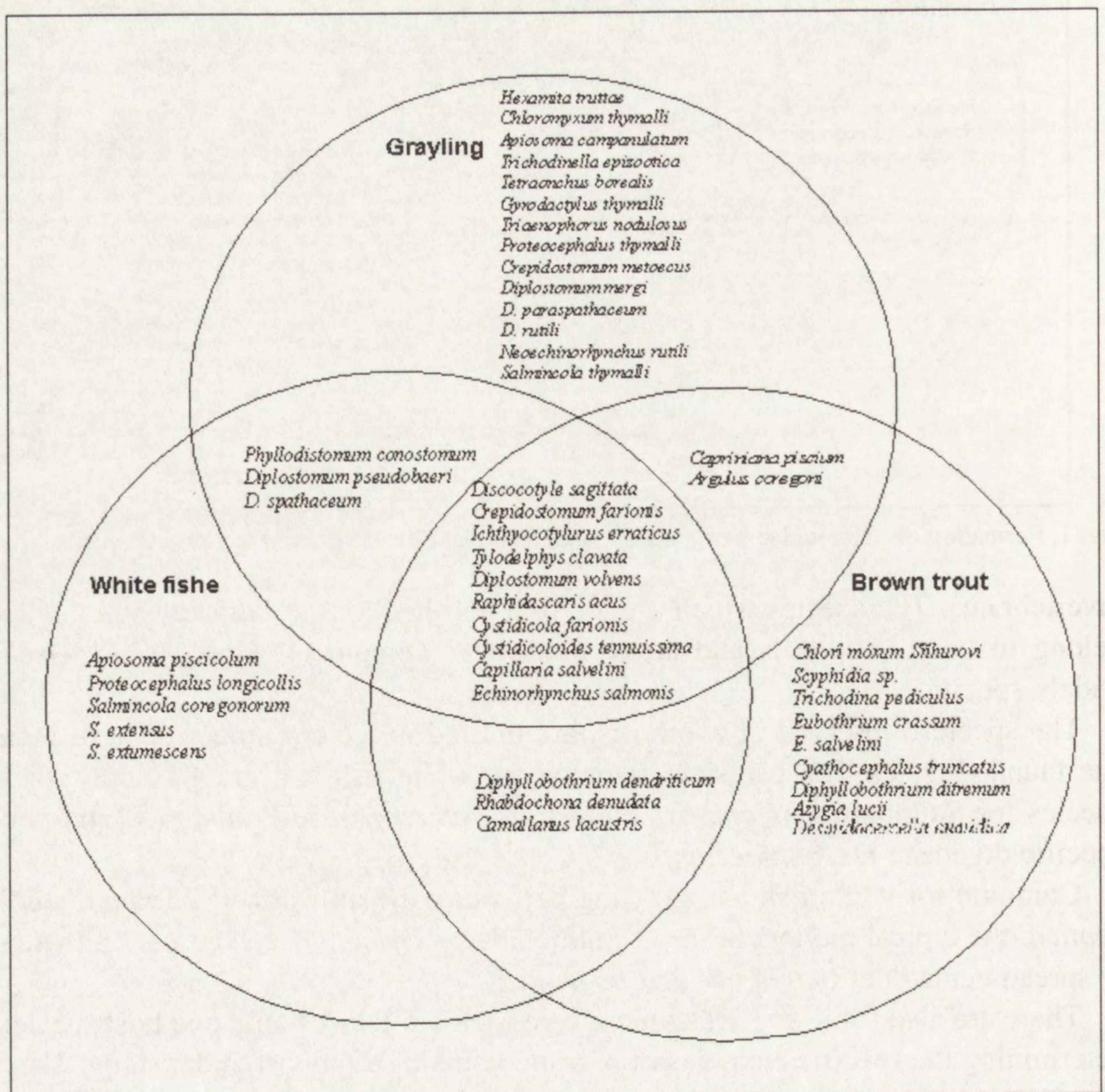


Fig. 2. Formation of parasite fauna of Salmonidae adults parasite fauna from lake-river system Paanajrvi-Olanga

Table 2. The parasite fauna of adult Salmonidae fish: prevalence (%)/abundance

| Species of parasite                | Brown trout | White fish | Grayling |
|------------------------------------|-------------|------------|----------|
| <i>Hexamita truttae</i>            | —           | —          | 61/+     |
| <i>Chloromyxum schurovi</i>        | 47/+        | —          | —        |
| <i>Ch. thymalli</i>                | —           | —          | 5/+      |
| <i>Capriniana piscium</i>          | 20/0.26     | —          | 44/35.3  |
| <i>Scyphidia</i> sp.               | 20/0.001    | —          | —        |
| <i>Apiosoma campanulatum</i>       | —           | —          | 6/0.3    |
| <i>A. piscicolum</i>               | —           | 30/0.11    | —        |
| <i>Trichodina pediculus</i>        | 13/0.8      | —          | —        |
| <i>Trichodinella epizootica</i>    | —           | —          | 27/0.3   |
| <i>Tetraonchus borealis</i>        | —           | —          | 61/17.0  |
| <i>Gyrodactylus thymalli</i>       | —           | —          | 16/1.4   |
| <i>Discocotyle sagittata</i>       | 20/0.5      | 25/0.7     | 61/5.7   |
| <i>Triaenophorus nodulosus</i>     | —           | —          | 6/0.1    |
| <i>Eubothrium crassum</i>          | 33/6.3      | —          | —        |
| <i>E. salvelini</i>                | 47/14.2     | —          | —        |
| <i>Cyathocephalus truncatus</i>    | 7/0.1       | —          | —        |
| <i>Diphyllobotrium dendriticum</i> | 7/0.1       | 19/0.1     | —        |
| <i>D. ditremum</i>                 | 7/0.1       | —          | —        |
| <i>Proteocephalus longicollis</i>  | —           | 56/38.4    | —        |
| <i>P. thymalli</i>                 | —           | —          | 33/0.8   |
| <i>Phyllodistomum conostomum</i>   | —           | 19/1.5     | 11/0.1   |
| <i>Azygia lucii</i>                | 13/0.3      | —          | —        |
| <i>Crepidostomum farionis</i>      | 20/4.6      | 6/0.1      | 39/62    |
| <i>C. metoecus</i>                 | —           | —          | 11/109   |
| <i>Ichthyocotylurus erraticus</i>  | 20/0.5      | 63/159     | 6/0.5    |
| <i>Tylodelphys clavata</i>         | 7/0.1       | 63/38.0    | 17/1.1   |
| <i>Diplostomum mergi</i>           | —           | —          | 5/0.1    |
| <i>D. paraspathaceum</i>           | —           | —          | 5/0.1    |
| <i>D. pseudobaeri</i>              | —           | 25/0.6     | 33/1.2   |
| <i>D. rutili</i>                   | —           | —          | 11/0.1   |
| <i>D. spathaceum</i>               | —           | 25/0.4     | 17/0.1   |
| <i>D. volvens</i>                  | 33/0.6      | 88/6.1     | 11/0.1   |
| <i>Rhaphidascaris acus</i>         | 53/1.9      | —          | —        |
| <i>R. acus</i> l.                  | —           | 88/41      | 61/11.6  |
| <i>Rhabdochona denudata</i>        | 7/0.1       | 6/0.1      | —        |
| <i>Cystidicola farionis</i>        | 33/1.5      | 63/7.0     | 61/30.5  |
| <i>Cystidicoloides tenuissima</i>  | 47/1.9      | 38/2.0     | 100/321  |
| <i>Desmidocercella numidica</i>    | 13/0.1      | —          | —        |
| <i>Camallanus lacustris</i>        | 13/01       | 6/1.6      | —        |
| <i>Capillaria salvelini</i>        | 40/3.5      | 38/5.0     | 28/0.3   |
| <i>Neoechinorhynchus rutili</i>    | —           | —          | 11/0.2   |
| <i>Echinorhynchus salmonis</i>     | 53/3.9      | 38/16.0    | 80/8.2   |
| <i>Salmincola coregonorum</i>      | —           | 13/0.3     | —        |
| <i>S. extensus</i>                 | —           | 6/0.1      | —        |
| <i>S. extumescens</i>              | —           | 13/0.1     | —        |
| <i>S. thymalli</i>                 | —           | —          | 66/6.2   |
| <i>Argulus coregoni</i>            | 13/0.1      | —          | 6/0.3    |
| No. of parasite species            | 24          | 21         | 29       |
| No. of fish studied                | 15          | 16         | 18       |

In general the core is formed by the species, whose development is connected with benthic organisms; except is nematode *R. acus*, which can be received by fish as a result of preying on both other fish and some invertebrates: Chironomidae and Oligocheta. There are differences in quantity of these species in the parasite fauna of white fish, brown trout and grayling. The grayling is higher infected by nematode *Cystidicoloides tenuissima* than other fish. The same was observed in parr. Our study (Table 2) and the study of the parasite fauna of European grayling in water bodies of European North showed that it is the main definitive host of this parasite. And only in the water bodies, where grayling is absent or not numerous, other species of Salmonidae fish play role of the main host, supporting the size of this parasite population (Dorovskih 1996).

*Ichthyocotylurus erraticus* is the dominating species in the parasite fauna of white fish both the parr and adults. Besides, the parasite fauna of this host is characterized by high level of abundance of the species, developing *via* bottom invertebrates.

In the parasite fauna of brown trout many species show high level of prevalence and abundance. It is impossible to determine the dominating species.

*Phyllodistomum conostomum*, *D. pseudobaeri*, and *D. spathaceum*, which are included into parasite fauna of white fish and grayling are connected with benthic organisms. However, in fish parasite fauna studied in other water bodies of the system, these species have been also found in brown trout (Barskaya et al. 2001); similar *Capriniana piscium* and *Argulus coregoni*, discovered in grayling and brown trout, can be also found in white fish.

The similarity in ecology of brown trout and white fish, being manifested, among other, by the presence of zooplankton in their diet, is emphasized by the presence of the common species *Diphyllobothrium dendriticum* in parasite fauna of those fish.

Therefore, the relationships between parasites and their common hosts, established in juveniles, persist in the fish of older age groups, beside new ones, characteristic for adult fish only.

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