M. DZIEDZICKA-WASYŁEWSKA, W. KOLASIEWICZ, Z. ROGÓŻ, W. MARGAS, J. MAJ

THE ROLE OF DOPAMINE D₂ RECEPTOR IN THE BEHAVIORAL EFFECTS OF IMIPRAMINE — STUDY WITH THE USE OF ANTISENSE OLIGONUCLEOTIDES

Department of Pharmacology, Institute of Pharmacology, Polish Academy of Sciences, Kraków, Poland

protein, e.g. receptor subtype, thus may help to uncover its behavioral and/or biochemical function. In the present study we demonstrated the utility of this approach for studying the role of dopamine D₂ receptors in the anti-immobility effect of imipramine in the forced swimming test. Following intracerebroventricular (i.e.v.) administration of phosphorothioate oligonucleotide complementary to mRNA encoding for dopamine D₂ receptors (D₂ antisense ODN; 1 nmol/1µl H₂O, twice a day for 5 days) to the rats, the decrease in the locomotor activity (shortened total distance travelled and decrease in vertical activity, without differences in the stereotypic movements of animals), as well as the decrease of specific binding of [³H]raclopride in the striatum and limbic forebrain were observed. At the same time, i.e.v. administration of D₂ antisense ODN reversed the effect of imipramine in the forced swimming test, what may indicate that the dopamine D₂ receptors play a significant role in the behavioral anti-immobility effects of imipramine.

Antisense strategies have a potential to specifically block the production of a given

Key words: dopamine D_2 antisense oligonucleotides (D_2 antisense ODN), imipramine, locomotor activity, forced swimming test, binding of $[^3H]$ raclopride, rats

INTRODUCTION

The role of dopamine D_2/D_3 receptors in the mechanism of action of antidepressant drugs (ADs) has been shown in our previous studies (1—5). However, a persistent methodological problem associated with these studies is that the dopaminomimetic drugs used may have actions on other CNS receptors (e.g. amphetamine with the indirect effect on α_1 -adrenergic receptors) so that their effects may not be specific to their dopaminomimetic activity.

dopamine receptor subtypes recently cloned (6) which were previously classified into two families (D_1 and D_2) according to their pharmacological profiles (7). Although selective drugs easily discriminate between these two families, drugs that usefully discriminate between family members (D_1 versus D_5 ; D_2 versus D_3 and D_4) are not yet available. These include one of the classic dopamine D_2 receptor agonist, quinpirole, which now has been shown to bind also to dopamine D_3 receptor (8) as well as several of the most widely used antipsychotics (9).

Indeed, the synaptic actions of dopamine are mediated by at least five distinct

It is well known that a possible effect in the forced swimming test is produced by various substances which stimulate the locomotor activity. An effect which is not accompanied with the locomotor hyperactivity is regarded as a specific antidepressant-like effect. The ADs-induced reduction of the immobility time in the forced swimming test in rats is considered to be mainly due to activation of the dopamine system. However, despite the studies done by Delini-Stula *et al.* (10), who have shown the antagonism of anti-immobility effect of levoprotiline by haloperidol and sulpiride, but not by prazosin, the critical involvement of dopamine D_2/D_3 receptors was not clearly proved.

Therefore, since the selectivity of widely used drugs acting at the level of dopamine receptors should be reconsidered, the antisense oligonucleotides (ODNs), complementary to the specific mRNA sequences encoding dopaminergic receptors have been supposed to become a tool for blocking specific gene expression, thus helping to elucidate the specific role of appropriate proteins (e.g. receptors) in certain experimental paradigms. The major advantage of this approach is the — theoretically — relatively simple, rational design and synthesis of ODNs that should bind only to specific nucleic acid sequence. The problem of rapid degradation of ODNs in tissues has been partially obviated by chemically modifying ODNs. Phosphorothioate ODNs are less likely to be degraded by nucleases (11) or RNAase H (12) than their unmodified counterparts. This modification increases their half-lives to more than 24 h (13). Since ODNs do not pass an intact blood-brain barrier (12, 14), intracerebroventricular (i.c.v.) or intracerebral injection have become

In the present study we designed the *antisense* sequence complementary to mRNA encoding for dopamine D_2 receptors (phosphorothioate D_2 antisense ODN) in order to see how important this subtype of dopamine receptors is for the effect of imipramine in forced swimming test. The *antisense* sequence has been chosen following several published reports indicating its efficacy in decreasing the dopamine D_2 receptor density as well as its function manifested in certain behavioural paradigms (16—19).

a standard in brain research (15).

MATERIALS AND METHODS

The experiments were carried out on rats (male Wistar, 250—270 g). The animals had free access to food and water before experiment and were kept in a constant room temperature $(22 \pm 1^{\circ}\text{C})$ on a natural day-night cycle. The experiments were performed in accordance with the ethical requirements.

D₂ Antisense ODN

Based on the cDNA sequence for the dopamine D₂ receptor (20, 21) a 20-mer phosphorothioate oligodeoxynucleotide was designed and purchased (molecular biology grade) from Biometra GmBH, Germany. The antisense was targeted to the area of the dopamine D₂ receptor cDNA sequence that bridges initiation codon (from -10 to +10: 5'-GTG GAT CCA TTG GGG CAG TG—3'). This selected target sequence has relatively low homology with any of the other known cDNA sequences found in the National Center for Biotechnology Information (BLAST 2.0).

Intracerebroventricular administration of D2 antisense ODN

The rats were operated under pentobarbital anaesthesia (30 mg/kg i.p.), 24 h before and after the operation the antibiotics (1ml/kg, Linco-Spectin, Upjohn) were administered. They were implanted chronically and bilaterally with stainless guide cannulae 9.0 mm long (0.4 mm o.d.), according to the method described by Paxinos and Watson (22). After 7-day postoperative period, D_2 antisense ODN solution (1 nmol in 1 μ l of H_2O) was administered 10 times in 12-h intervals to conscious unrestrained animals via an internal cannula (10.6—11.0 mm long; 0.3 mm o.d.) that extended beyond the guide cannula to the lateral ventricle (AP -0.4—0.8, L 1.2—1.5, V 2.0—2.4). Antisense solution or solvent was administered in a volume of 1 μ l over 2 min with a 2 microliter Hamilton syringe. The internal cannula was withdrawn 1 min before the termination of the injection. Postoperatively, rats were housed singly with food mash and water provided ad libitum.

[3H] Raclopride binding

For binding experiments the rats were sacrificed 12 h after the last dose of D₂ antisense ODN, their brains were quickly removed and striata and limbic forebrains were dissected out, frozen on dry ice and stored until used. Specific binding of [³H]raclopride was performed, according to the method described previously (5).

Locomotor activity

At 48 h before the test animals were adapted for 24 h to the experimental room. After adaptation, 12 h after the last i.e.v. administration of D₂ antisense ODN, each animal was placed individually into one of four plexiglass cages. Four Animal Activity Meter (Auto-Track System, Columbus Instruments, USA) was used. The animals were observed continously for 30 min.

The Auto-Track system consists of plexiglass monitor cages $(40 \times 40 \times 22 \text{ cm})$ surrounded by horizontal and vertical infrared sensors non-detectable by the animal. The monitor cages are connected to a Digiscan activity monitors. Data for the following variables of locomotor activity detected by the Digiscan, were collected in an IBM PC/XT compatible computer system: DT — total distance travelled by the animal (cm), BSM — number of stereotypic movements (all tiny

movements not linked to the displacement, i.e. grooming, scratching and sniffing around) and V — vertical activity, i.e. single ambulations on cage walls as well as typical rearing with sniffing and looking around.

Forced swimming test

The total immobility time was measured 12 h after the last i.c.v. administration of D₂ antisense ODN.

The total immobility time of rats was assessed according to Porsolt *et al.* (23) during a 5-minute observation period. Imipramine (Polfa, Poland; 10 mg/kg) was given three times at 24, 5 and 1 h before the test to naive and operated animals.

Histological analysis

After completion of the experiment, the rats were deeply anaesthetized with pentobarbital (450 mg/kg i.p.), perfused intracardially with isotonic saline solution followed by perfusion with 4% paraformaldehyde. The brains were taken out and stored in the fixative (8% formaldehyde solution). Coronal sections were cut from the frozen tissue at 90 µm, photographed and the position of all injection cannula tips was checked. Only those animals with histologically confirmed proper injection sites were used in the data analysis.

Statistical analysis

The results are expressed as means ± SEM. A statistical evaluation was carried out by Student's t-test, or by an analysis of variance (one-way ANOVA) followed by Newman-Keuls test. In biochemical studies ANOVA followed by Duncan's test was used.

RESULTS

Locomotor activity

I.c.v. administration of D_2 antisense ODN decreased all measured parameters of rat locomotor activity, i.e. shortened the total distance travelled (DT) and caused decrease in vertical activity (V). At the same time no differences in the stereotypic movements of animals (BSM) were observed (Fig. 1).

Binding studies

(Fig. 2).

Specific binding of [3H]raclopride was decreased following i.c.v. administration of D₂ antisense ODN, both in the striatum as well as in the limbic forebrain. The effect was more pronounced in the latter brain region

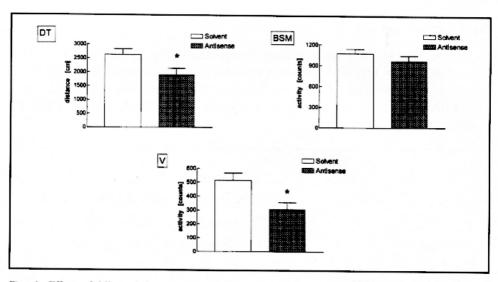


Fig. 1. Effect of bilateral intraventricular (lateral ventricle) injections of D₂ antisense ODN (1 nmol/1 μl; n = 5) or solvent (1 μl; n = 9) on spontaneous activity of the rats. DT — distance travelled [cm]; BSM — burst of stereotypic movements (i.e.: grooming, scratching, sniffing around) [counts]; V — vertical activity (i.e.: rearings, climbing up cage walls) [counts]. The total observation time was 30 min.

*p<0.05 vs solvent-receiving group, unpaired t-Student test.

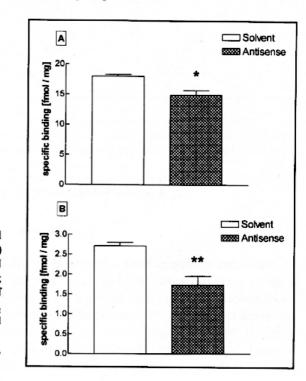
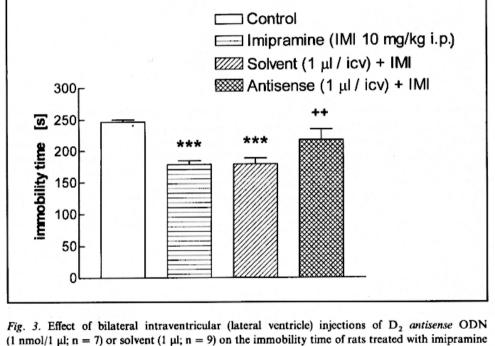


Fig. 2. Effect of bilateral intraventricular (lateral ventricle) injections of D2 antisense ODN (1 nmol/1 μ l; n = 6) or solvent (1 μ l; n = 7on specific binding of [3H]raclopride to dopamine receptors in the rat striatum (A) and limbic forebrain (B).

*p<0.05 vs solvent-receiving group, ANOVA followed by Duncan's test.

Forced swimming test

Imipramine (10 mg/kg) significantly shortened the immobility time. The effect was almost identical in intact animals and in the animals which were operated and received i.c.v. administration of the solvent. However, i.c.v. administration of D, antisense ODN reversed the anti-immobility effect of imipramine (Fig. 3).



(IMI, 10 mg/kg i.p.; n = 16). Control — animals receiving solvent (2ml/kg i.p.) without imipramine (n = 16).

***p<0.001 vs control, ++p<0.01 vs solvent-receiving group, ANOVA (number of groups = 4; F = 21.57, R squared = 0.5952), followed by Newman-Keuls test.

DISCUSSION

Antisense strategies have a potential to specifically block the production of a given receptor subtype, thus may help to uncover its behavioral and/or biochemical function. The number of studies using antisense strategy for clarifying the behavioral function of dopamine D2 receptors increases in recent years. It has been shown that i.c.v. administrating the D2 antisense ODN spontaneous locomotor activity, inhibited quinpirole-induced reduced

locomotor activation and elicited catalepsy in rats (24), inhibited rotations

induced by quinpirole in 6-OH-DA lesioned mice (16, 17), supressed — after intrastriatal administration — stereotyped sniffing elicited by apomorphine in the rat (25) and significantly influenced — after intranigral administration — the motor actions of cocaine (26).

It is widely accepted that normal, physiological functions of dopamine D₂ receptors (also pharmacological blockade or stimulation of these receptors) are responsible for locomotor activation (so called "forward" locomotor activity, measured as distance travelled) and for exploratory activity (measured as a number of rearings). On the other hand, behavioral activity such as grooming or scratching and spiffing are believed to be regulated by dopamine

as a number of rearings). On the other hand, behavioral activity such as grooming or scratching and sniffing, are believed to be regulated by dopamine D_1 receptors. This notion seems to be confirmed in the present study, since we observed significant decrease in locomotor (DT) and exploratory (V) activity, without any changes in BSM, after i.c.v. administration of D_2

The results obtained in the present work demonstrate the utility of

antisense ODN.

this approach for the studying the role of dopamine D2 receptors in anti-immobility effect of imipramine in the forced swimming test. This test, initially described by Porsolt (23), although bears oversimplification of a highly complex illness as depression is, has been widely used in the studies of action of ADs as well as in order to find compounds with novel antidepressant activity. It has been shown that various compounds which stimulate the locomotor activity are also active in the forced swimming test, and the effect which is not accompanied with locomotor hyperactivity is regarded as specific antidepressant effect. The i.c.v. administration of D, antisense ODN induces locomotor hypoactivity, therefore it is justified to interprete its reversal of anti-immobility effect of imipramine as a result of diminished synthesis of dopamine D2 receptor. Especially, since in biochemical experiments we observed the decreased specific binding of [3H]raclopride, D₂ receptor antagonist, following i.c.v. administration of D, antisense ODN, which may be interpreted as a reduction in the level of D, receptors. The degree of this decrease is relatively small compared to the reduction in behavioral changes. However, other investigators have already reported that small changes in the density of dopamine receptors may be associated with profound changes in dopamine-mediated behavioral responses (19). Since administration of D2 antisense ODN causes a decreased synthesis in D2 receptors, it would cause a proportionally greater decrease

Therefore, it may be concluded that, indeed, the dopamine D_2 receptors play a significant role in the behavioral anti-immobility effects of imipramine.

by the binding of an antagonist used as radioligand.

in the functional receptors compared with the total pool of receptors, thereby resulting in a large decrease in biological function but a relatively small decrease in the total pool of receptors, which is — in fact — detected

Acknowledgement: This research was supported by Grant No 4.P05F 012 14 from the State Committee for Scientific Research, Warszawa, Poland.

REFERENCES

- Maj J, Rogóż Z, Skuza G, Sowińska H. Repeated treatment with antidepressant drugs increases the behavioral response to apomorphine. J Neural Transm 1984; 60: 273—282.
- Maj J, Rogóż Z, Skuza G, Sowińska H. Antidepressants given repeatedly increase the behavioral effects of dopamine D₂ agonists. J Neural Transm 1989; 78: 1—8.
- Dziedzicka-Wasylewska M, Rogoż R, Klimek V, Maj J. Repeated administration of antidepressant drugs affects the levels of mRNA coding for D₁ and D₂ dopamine receptors in the rat brain. J Neural Transm 1997; 104: 515—524.
- Maj J, Dziedzicka-Wasylewska M, Rogoż R, Rogóż Z. Effect of antidepressant drugs administered repeatedly on the dopamine D₃ receptors in the rat brain. Eur J Pharmacol 1998; 351: 31—37.
- Rogoż R, Dziedzicka-Wasylewska M. Effects of antidepressant drugs on the dopamine D₂/D₃ receptors in the rat brain differentiated by agonist and antagonist binding an autoradiographic analysis. Naunyn-Schmiedeberg's Arch Pharmacol 1999; 359: 178—186.
- Sibley DR, Monsma FJ. Molecular biology of dopamine receptors. Trends Pharmacol Sci 1992; 13: 61—69.
- 7. Kebabian JW, Calne DB. Multiple receptors for dopamine. Nature 1979; 277: 93-96.
- Sokoloff P, Giros B, Martres MP, Bouthenet ML, Schwartz JC. Molecular cloning and characterization of a novel dopamine receptor (D₃) as a target for neuroleptics. *Nature* 1990; 347: 146—151.
- Van Tol HHM, Bunzow JR, Guan HC, Sunahara RK, Seeman P, Niznik HB, Civelli O. Cloning of the gene for a human dopamine D₄ receptor with high affinity for the antipsychotic clozapine. Nature 1991; 350: 610—614.
- Delini-Stula A, Radeke E, van Riezen H. Enhanced functional responsiveness of the dopaminergic system the mechanism of anti-immobility effects of antidepressants in the bahavioral despair test in the rat. Neuropharmacol 1988; 27: 943—947.
- Whitesell L, Geselowitz D, Chavany C, Fahmy B, Walbridge S, Alger JR, Neckers LM. Stability, clearance and disposition of intraventricularly administered oligodeoxynucleotides: implications for therapeutic application within the central nervous system. *Proc Natl Acad Sci USA* 1993; 90: 4665—4669.
- 12. Agrawal S, Temsamani J, Tang JY. Pharmacokinetics, biodistribution, and stability of oligonucleotide phosphorothioates in mice. Proc Natl Acad Sci USA 1991; 88: 7595—7599.
- oligonucleotide phosphorothicates in mice. Proc Natl Acad Sci USA 1991; 88: 1595—1599.

 13. Crooke RM, Graham MJ, Cooke ME, Crooke ST. In vitro pharmacokinetics of
- phosphorothioate antisense oligonucleotides. J Pharmacol Exp Ther 1995; 275: 462—473.

 14. Wahlestedt C. Antisense oligonucleotide strategies in neuropharmacology. Trends Pharmacol
- Sci 1994; 15: 42—46.

 15. Crooke ST, Bennet CF. Progress in antisense oligonucleotide therapeutics. Ann Rev Pharmacol
- Toxical 1996; 36: 107—129.

 16. Weiss B, Zhou LW, Zhang SP, Qin ZH. Antisense oligodeoxynucleotide inhibits D₂ dopamine
- receptor-mediated behavior and D₂ messenger RNA. Neuroscience 1993; 55: 607—612.

 17. Zhou LW, Zhang SP, Qin ZH, Weiss B. In vivo administration of an oligodeoxynucleotide antisense to the D₂ dopamine receptor mRNA inhibits D₂ dopamine receptor-mediated behavior and the expression of D₂ dopamine receptors in mouse striatum. J Pharmacol Exp

Ther 1994; 268: 1015-1023.

- Zhang SP, Zhou LW, Morabito M, Lin RCS, Weiss B. Uptake and distribution of fluorescein-labeled D₂ dopamine receptor antisense oligonucleotide in mouse brain. J Mol Neurosci 1996, 7: 13—28.
 Weiss B, Zhang SP, Zhou LW. Antisense strategies in dopamine receptor pharmacology. Life
- Sci 1997; 60: 433—455.

 20. Bunzow JR, Van DI HHM, Grandy DK et al. Cloning and expression of a rat D₂ dopamine
- receptor cDNA. Nature 1988; 336: 783—787.
 Mack KJ, Todd RD, O'Mallen KL. The mouse D₂ receptor gene: sequence homology with the rat and human genes and expression of alternative transcripts. J Neurochem 1991; 57: 795—801.
- Paxinos G, Watson C. The Rat Brain in Stereotaxic Coordinates. Academic Press, Sydney, 1986.
 Porsolt RD, Le Pichon M, Jaltre M. Depression: a new animal model sensitive to antidepressant treatments. *Nature* 1977; 266: 730-732.
- Zhang M, Creese I. Antisense oligonucleotide reduces brain dopamine D₂ receptors: behavioral corelates. Neurosci Lett 1993; 161: 223-226.
 Rajakumar N, Laurier L, Niznik HB, Stoessl AJ. Effect of intrasriatal infusion of D₂ receptor antisense oligonucleotide on apomorphine-induced behaviors in the rat. Synapse 1997; 26:
- 199—208.
 26. Silvia CP, King GR, Lee TH, Xue ZH, Caron MG, Ellinwood EH. Intranigral administration of D₂ dopamine receptor antisense oligodeoxynucleotides establishes a role for nigrostratal D₂ autoreceptors in the motor actions of cocaine. *Mol Pharmacol* 1994; 46: 51—57.
 - Received: May 4, 2000
 Accepted: July 6, 2000

 Author's address: M. Dziedzicka-Wasylewska, Dept of Pharmacology, Institute of Pharmacology, Polish Academy of Sci. 12 Smetna Street, 31-343 Kraków, Poland