

G. JURKOWSKA, J. W. DLUGOSZ, G. RYDZEWSKA, A. ANDRZEJEWSKA *

THE EFFECT OF PROSTACYCLIN ANALOGUE—ILOPROST ON THE PANCREAS REGENERATION AFTER CAERULEIN-INDUCED ACUTE PANCREATITIS IN RATS

Gastroenterology Department, Anatomopathology Department*, University Medical School, Bialystok, Poland

Prostacyclin (PGI₂) and its stable analogue iloprost (I) exert beneficial effect in acute pancreatitis (AP). The aim of the study was to evaluate the role of iloprost in pancreas regeneration after AP in rats. Methods: AP was induced in male Wistar rats by s.c. injections of caerulein 12 µg/kg t.i.d. for 2 days. Rats were divided into four groups: control + saline (C); control + iloprost (C/I); AP + saline (AP); AP + I (AP/I). Rats were treated for 7 or 14 days. I (Schering AG) was given at the dose 1 µg/kg b.w., i.p., t.i.d. After the rats were killed, the pancreata were weighed and their protein, DNA, RNA, chymotrypsin, α-amylase contents were evaluated. Light microscopic examination of representative pieces of pancreas was performed. Results: Acute pancreatitis resulted in pancreas destruction observed even 7 days after the onset of the disease. The significant decrease of pancreatic weight, RNA and chymotrypsin contents were observed in AP rats when compared to C. The improvement of pancreatic histology and significant increase of DNA content were found in I treated (during 7 days) AP rats in comparison to untreated AP group. Two weeks after pancreatitis induction the pancreas regeneration occurred in both pancreatitis groups and it was connected with pancreas hypertrophy. Treatment with I resulted in slight not significant increase of some of cellular hypertrophy indices when compared to AP untreated animals. Healthy rats injected with I during 7 days showed significant elevation of DNA content in comparison to C. When treatment with I was prolonged up to 14 days such hyperplastic effect was not observed. Our results suggest, that treatment with iloprost exerts temporary hyperplastic influence on the pancreas of healthy rats and pancreas regenerating after caerulein-induced pancreatitis.

Key words: acute pancreatitis, regeneration, nucleic acids, prostaglandins, iloprost.

INTRODUCTION

Pancreas regeneration after partial resection, duct ligation injury, toxic injury and acute pancreatitis has been described (1). The time course of the regenerative response has been studied extensively but mechanisms and factors involved in this process are still poorly clarified (2).

The prostanoid imbalance has been found to be an important factor in the pathogenesis of acute pancreatitis (3). The protective effect of different natural

prostaglandins and their analogues on the course of acute pancreatitis has been shown (4—6). The mechanisms responsible for their beneficial influence involve: the stabilization of lysosomal membranes within acinar cells (7, 8), inhibition of intracellular activation of pancreatic digestive enzymes (9), improvement of blood supply (10). Prostacyclin and its analogue iloprost has been also shown to exert platelet antaggregatory influence. Beside that, both PGI₂ and iloprost inhibit the leukocytes chemotaxis and decrease the release of lysosomal hydrolases and the generation of oxygen free radicals from inflammatory cells, agents potentially noxious to the endothelial and acinar cells (11). Prostaglandins have been also shown to exert the trophic effect on gastrointestinal mucosa (12, 13) and liver (14). Contrary, the inhibitors of prostaglandin production exert some inhibiting action both on DNA synthesis and mitotic activity in the regenerative response of the rat liver after partial hepatectomy (15).

The goal of present study was to investigate the role of chemically stable carbacyclin analogue of PGI₂ — iloprost on the pancreas regeneration after caerulein induced acute pancreatitis in rats.

MATERIAL AND METHODS

Animals

Male Wistar rats (n = 52) weighing 240—260 g were used in the study. The animals were given a standard chow diet. Care was provided in accordance with the procedures outlined in the Guide for the Care and Use of Laboratory Animals (NIH publication # 85—23, 1985).

Experimental design

Acute pancreatitis was induced by s.c. injections of caerulein 12 µg · kg body weight⁻¹ every 8 h for 2 days. This analogue of cholecystokinin (CCK) was dissolved in gelatin (16% w/v) to prolong its absorption (2). The control animals were injected with saline in gelatin s.c. Beginning from day 3, rats were allocated into four treatment groups (6—8 rats in each):

- Control rats injected with saline (C).
- Control animals treated with PGI₂ stable analog iloprost (C/I).
- Rats with AP injected with saline (AP).
- Rats with AP treated with iloprost (AP/I).

Rats were treated for 7 or 14 days. Iloprost (I) (Schering AG, Germany) was given i.p. at a dose of 1 µg per kg b.w. three times per day (t.i.d.) The animals were fed *ad libitum* with a standard diet during whole experiment with a 12 hour light-dark cycle. After an overnight fast with free access to water rats were decapitated. The pancreas was dissected as quickly as possible, free of connective tissue, fat, lymph nodes and weighed. The representative fragments of pancreatic tissue from each rat were fixed in 10% formaline, embedded in paraffin, sectioned at 5-µm slices and stained with hematoxylin and eosin for light microscopic examinations.

Biochemical assays

The pieces of pancreatic tissue were homogenized using motor-driven ground-glass homogenizer either in 0.6 mol/L perchloric acid for nucleic acids determinations or in the ice-cold 0.1 mol/L Tris-HCl buffer (pH = 8.0) for protein, α -amylase and chymotrypsin assays.

DNA and RNA were extracted according to Mainz et al. (16). DNA was assayed according to Volkin and Cohn (17) using calf thymus DNA as a standard. RNA was measured after overnight hydrolysis in 0.3 mol/L KOH according to Munro and Fleck (18). Tissue α -amylase and chymotrypsin activities were determined according to the Bernfeld (19) and Hummel (20), respectively. Protein content was measured according to Lowry et al. (21).

All reagents except soluble starch (Merck, Darmstadt) were purchased from Sigma Chemical Co.

Statistical analysis

All values were expressed as means \pm SEM. Statistical significance of the difference between the groups was assessed using t test for unpaired observations and Fischer test. $P < 0.05$ was considered statistically significant.

RESULTS

Biochemical assays

Pancreatic weight (PW) decreased significantly in untreated rats with acute pancreatitis (AP) when compared to C (by 11%) 7 days after the pancreatitis induction (*Table 1*). Such decrease of PW was not observed after the treatment with iloprost in AP/I rats comparing to C, however PW in this group was significantly lower than in the control rats injected with I (C/I) ($p < 0.05$). Healthy animals injected with I during 7 days had higher PW than C rats (increase, by 8.8%, ns).

No significant differences of PW between healthy and pancreatitis rats were found 14 days after the onset of pancreatitis.

Pancreatic protein contents in both groups with AP (untreated or treated with I) tended to decrease 7 days after the induction of acute pancreatitis when compared to control (by 16% and 12% respectively), however differences were not significant (*Table 1*). Treatment with I during two weeks resulted in significantly higher protein content in AP rats as compared to C injected with I (when data were expressed per 100 g of b.w., the protein content increased also in comparison to C animals, data not shown).

Table 1. Pancreatic weight, total protein, enzymes and nucleic acids contents in caerulein-induced acute pancreatitis in rats treated with iloprost *. Means \pm SE are reported.

Parameter	Control (C)	Control-Iloprost (C/I)	Acute Pancreatitis (AP)	AP-Iloprost (AP/I)
P. weight (mg)				
7 days	847 \pm 44	930 \pm 38	747 \pm 29 *	823 \pm 34 °
14 days	1036 \pm 85	826 \pm 32	940 \pm 44	962 \pm 77
Protein (mg)				
7 days	87.8 \pm 8.8	94.3 \pm 10.4	73.7 \pm 7.8	77.1 \pm 8.9
14 days	90.2 \pm 18.1	88.0 \pm 9.8	108.6 \pm 7.3	119.5 \pm 11.1 °
Amylase (U)				
7 days	26769 \pm 3349	31963 \pm 3473	22386 \pm 4672	23690 \pm 3692
14 days	15663 \pm 2347	10760 \pm 1468	16238 \pm 2211	20933 \pm 3489 °
Chymotrypsin (U)				
7 days	362.7 \pm 52.8	425.2 \pm 115.5	126.0 \pm 25.2 *	242.4 \pm 35.3 */**
14 days	159.8 \pm 27.8	159.4 \pm 27.2	199.0 \pm 24.7	232.9 \pm 56.6
RNA (mg)				
7 days	24.3 \pm 1.4	24.2 \pm 2.2	18.7 \pm 1.2 *	18.9 \pm 1.2 */°
14 days	24.5 \pm 1.7	21.8 \pm 1.3	27.2 \pm 0.9	23.5 \pm 1.9
DNA (mg)				
7 days	4.49 \pm 0.22	6.12 \pm 0.40 *	4.85 \pm 0.08	5.4 \pm 0.17 */**
14 days	4.65 \pm 0.24	4.67 \pm 0.35	4.07 \pm 0.15	4.65 \pm 0.28
RNA/DNA				
7 days	5.4 \pm 0.19	4.09 \pm 0.50 *	3.87 \pm 0.29 *	3.53 \pm 0.31 *
14 days	5.3 \pm 0.43	4.69 \pm 0.12	6.70 \pm 0.37 *	5.23 \pm 0.61

* Iloprost (I) was injected at a dose of 1 μ g/kg b.w. tid i.p. for 7 or 14 days.

* values significantly different from control ($p < 0.05$).

** values significantly different from acute pancreatitis untreated rats ($p < 0.05$).

° values significantly different from control rats treated with I ($p < 0.05$).

Healthy animals injected with iloprost showed higher α -amylase content in comparison to C group (the difference was significant when data were expressed per 100 g of b.w., data not shown). After 14 days of treatment with I, α -amylase content increased significantly in AP rats vs. their control (C/I) (*Table 1*).

Pancreatic chymotrypsin content, 7 days after the pancreatitis induction was significantly lower in both groups with AP in comparison to C ($p < 0.05$), (*Table 1*). When I was injected to AP rats the chymotrypsin content significantly increased (by 48%) when compared to untreated animals ($p < 0.05$). PGI₂ analogue tended to increase pancreatic chymotrypsin content in healthy animals by 14.7% in comparison to C (ns). No significant differences were found in this parameter between all groups 14 days after the onset of pancreatitis.

Acute pancreatitis resulted in the significant decrease of pancreatic RNA content (in both groups) when compared to C, even 7 days after the induction of inflammation ($p < 0.01$), (*Table 1*). Iloprost did not change RNA content in healthy rats after 7 days of treatment (in comparison to C). No significant differences were found between treated and untreated groups 14 days after the pancreatitis induction.

Highly significant increase ($p < 0.005$) of pancreatic DNA content was observed after 7 days in healthy animals treated with I when compared to C and in AP rats injected with I when compared to the control group or untreated rats with AP (*Table 1*). After 14 days of experiment DNA contents in all groups were similar.

RNA/DNA ratio decreased in AP group 7 days after induction of the disease when compared to the control group. Treatment with I both healthy and AP rats was followed by proportional lowering of this ratio in both treated groups. Two weeks after the pancreatitis induction RNA/DNA ratio increased in AP rats in comparison to C ($p < 0.05$).

Using total DNA content as a measure of the cell number, it can be seen that iloprost treatment during 7 days induces pancreatic growth both in healthy and in AP animals, causing significant hyperplasia (*Table 2*). This influence of iloprost has not been observed after 14 days of treatment (*Table 2*). The cellular hypertrophy was established by dividing the total contents of RNA, protein, amylase and chymotrypsin by the total content of DNA. Pancreas regeneration after acute pancreatitis was connected with cellular hypertrophy 14 days after the pancreatitis induction. The indices of cellular hypertrophy (protein/DNA, amylase/DNA, chymotrypsin/DNA) in AP rats were slightly augmented by I treatment, but at this interval of time the difference did not achieved the level of statistical significance (*Table 2*).

Table 2. The effect of iloprost treatment on indices of pancreatic tissue hyperplasia and cellular hypertrophy in the rats with caerulein-induced acute pancreatitis[#]. Means \pm SEM are reported.

Parameter	Control-Iloprost (C/I)	Acute pancreatitis (AP)	AP-Iloprost (AP/I)
Hyperplasia			
DNA			
7 days	1.36 *	1.08	1.20 */**
14 days	1.00	0.87	1.00
Hypertrophy			
RNA/DNA			
7 days	0.75	0.71	0.65
14 days	0.88	1.26 *	0.97
Protein/DNA			
7 days	0.79	0.77	0.72
14 days	0.94	1.38 *	1.49
Amylase/DNA			
7 days	0.87	0.75	0.71
14 days	0.71	1.19	1.38
Chymotrypsin/DNA			
7 days	0.93	0.32	0.55
14 days	1.05	1.46 *	1.53

[#] Values are the ratios of experimental groups to control group after 7 and 14 days of treatment with iloprost (I) at a dose of 1 μ g/kg b.w. tid i.p. for 7 or 14 days. Total DNA content represents the pancreas hyperplasia. Cellular hypertrophy was established by dividing total contents of RNA, protein, amylase and chymotrypsin by the total content of DNA. Values significantly greater than 1 (in comparison to control) were accepted as a indices of cellular hypertrophy.

* Values significantly greater than 1, were significantly different from controls ($p < 0.05$).

** Values significantly different in comparison with acute pancreatitis rats ($p < 0.05$).

Light microscopic examination

Fig. 1 shows the normal pancreas appearance in control rats. The iloprost treatment of healthy rats resulted in little increase of eosinophilic granules within acinar cells (to similar extent after 7 and 14 days) when compared to control untreated rats (not shown).

Seven days after the induction of pancreatitis the oedema and small inflammatory infiltration in untreated rats were present. The acinar cells showed some signs of lesion (cytoplasmic degeneration and vacuolization) and a few dead acinar cells were also found (Fig. 2). The treatment with I resulted in the improvement of histological changes, however vacuolization of acinar cells and small inflammatory infiltration with polymorphonuclear cells were still observed (Fig. 3).

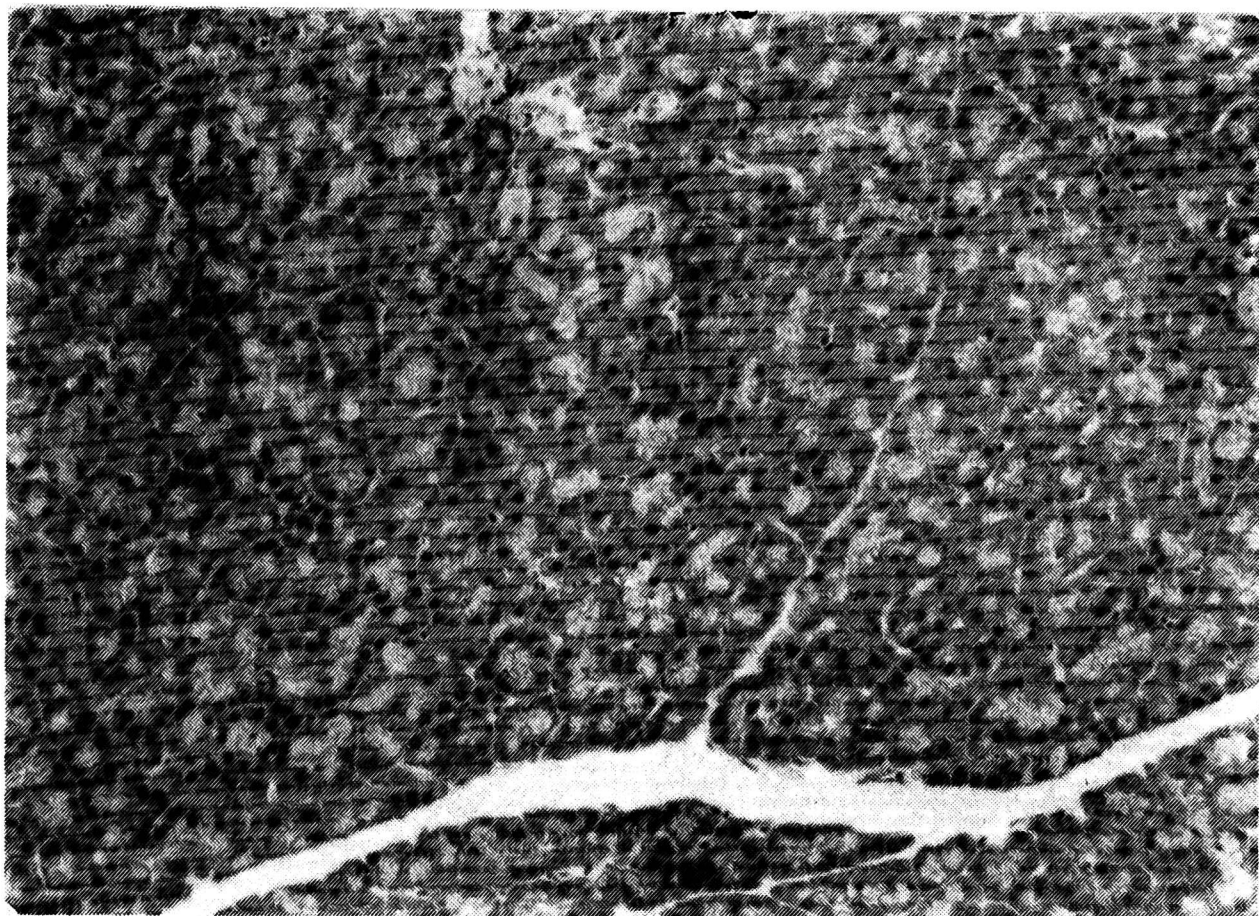


Fig. 1. Normal pancreas of rat from Control group (C 7 days). Mag. $\times 160$.

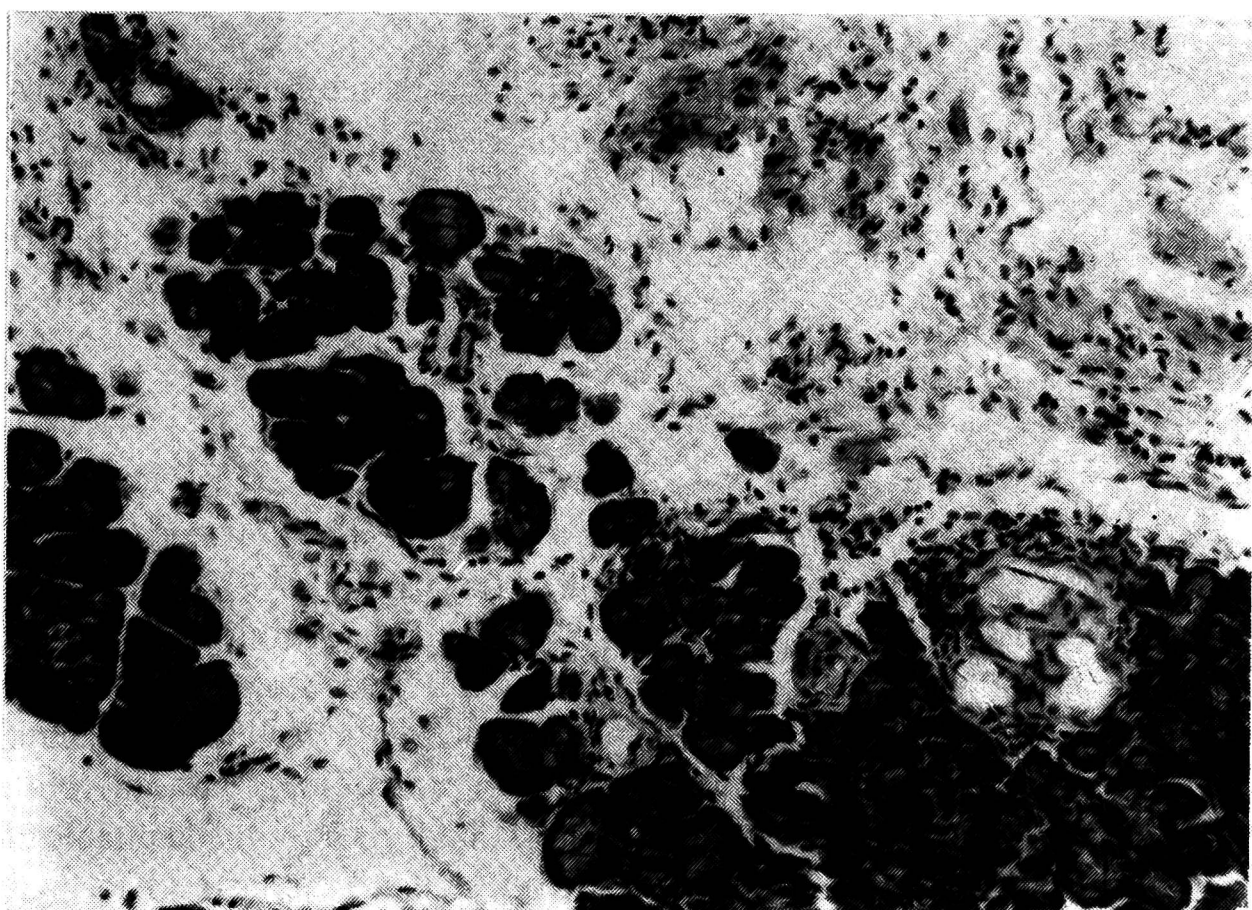


Fig. 2. Caerulein-induced pancreatitis — 7 days (AP 7 days). Oedema and inflammatory cell infiltration, signs of cytoplasmic degeneration, vacuolization and sporadic necrosis of acinar cells.
Mag. $\times 160$.

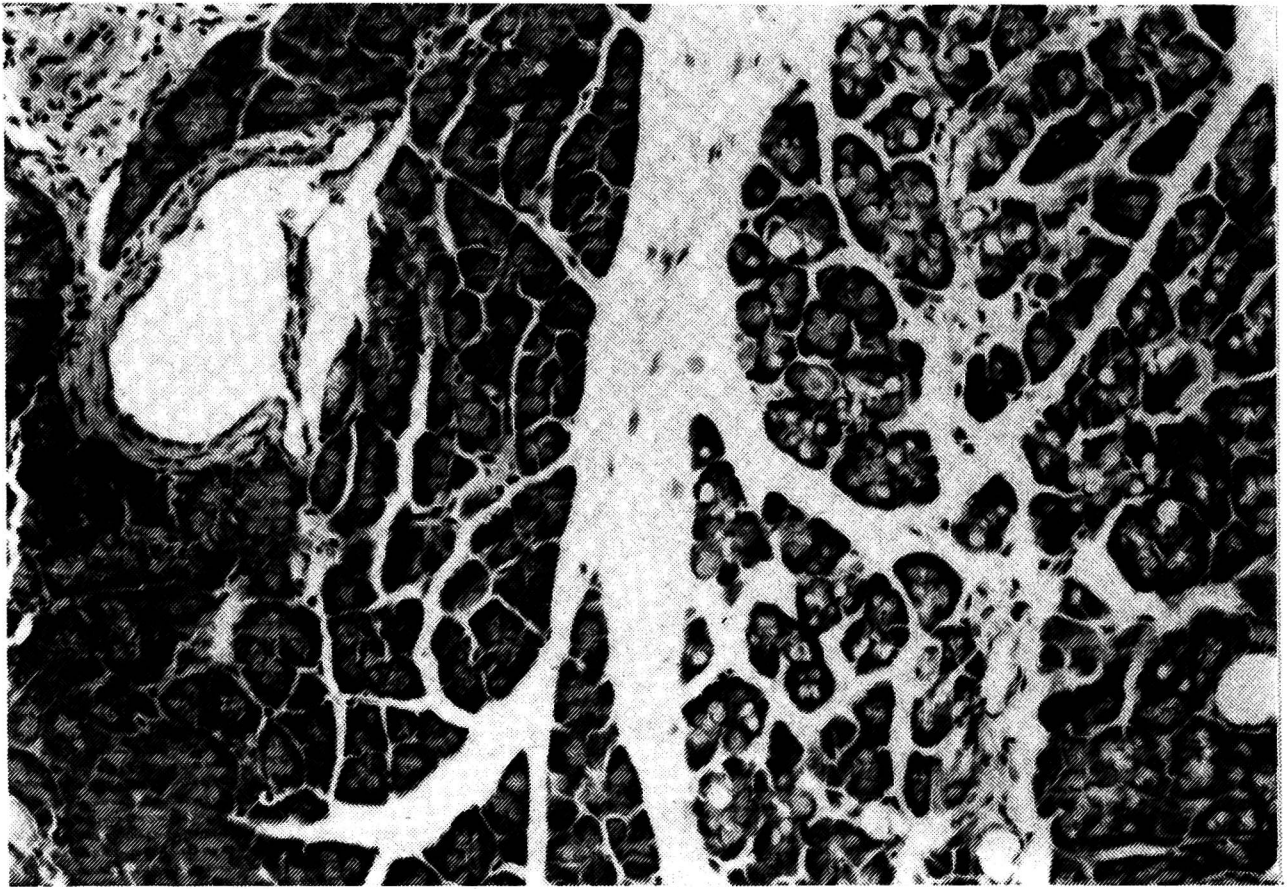


Fig. 3. Caerulein-induced pancreatitis — 7 days, treated with iloprost 1 $\mu\text{g}/\text{kg}$ b.w. t.i.d. (AP/I 7 days). Intersititial oedema and leucocyte infiltration less advanced than in untreated group. The vacuolization and some degenerative changes of acinar cells can be observed but cell necrosis is really scarce. Mag. $\times 160$.

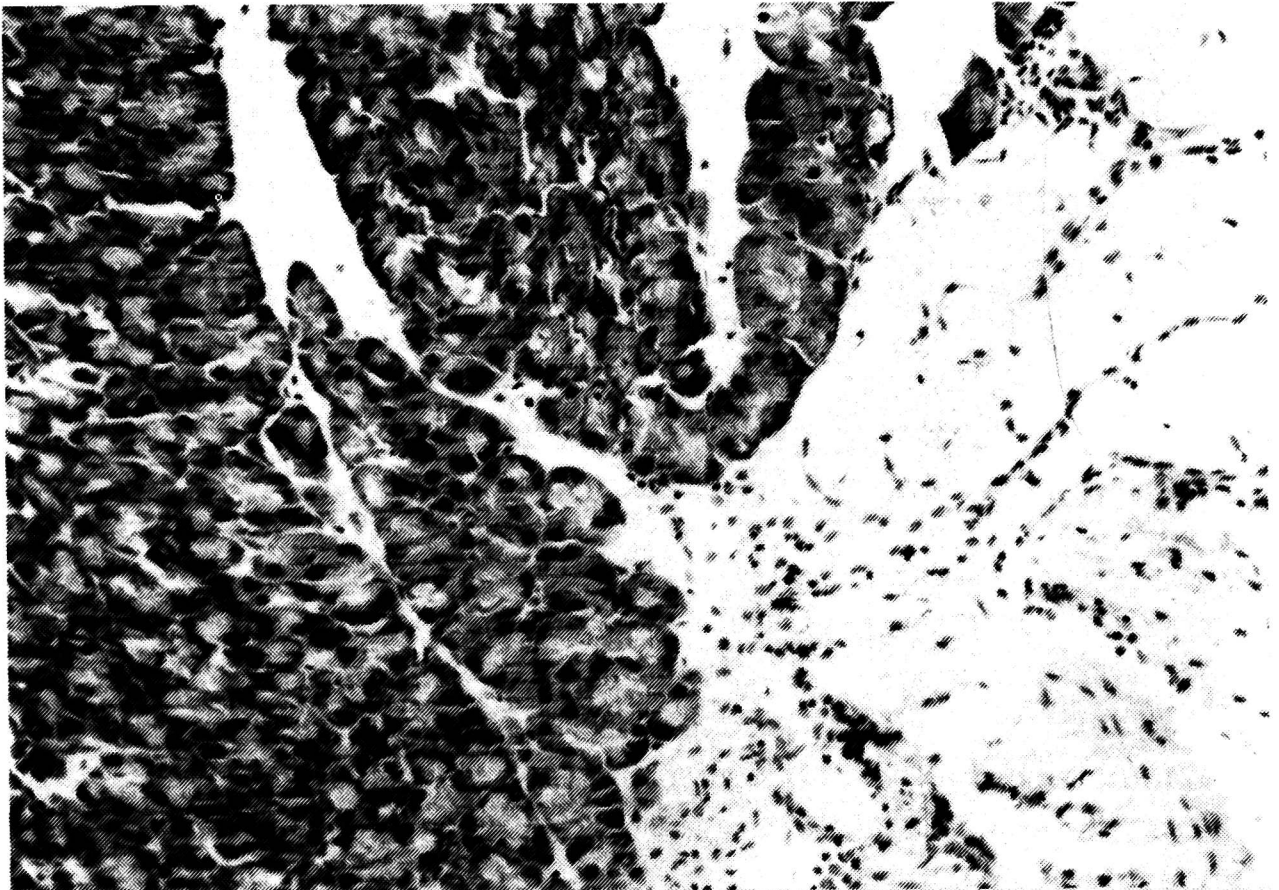


Fig. 4. Caerulein-induced pancreatitis — 14 days (AP 14 days). Traces of oedema and inflammatory cell infiltration. Small degree disarrangement of acinar cell morphology. Discrete degenerative changes of acinar cells. Evident improvement in comparison to AP lasting 7 days. Mag. $\times 160$.

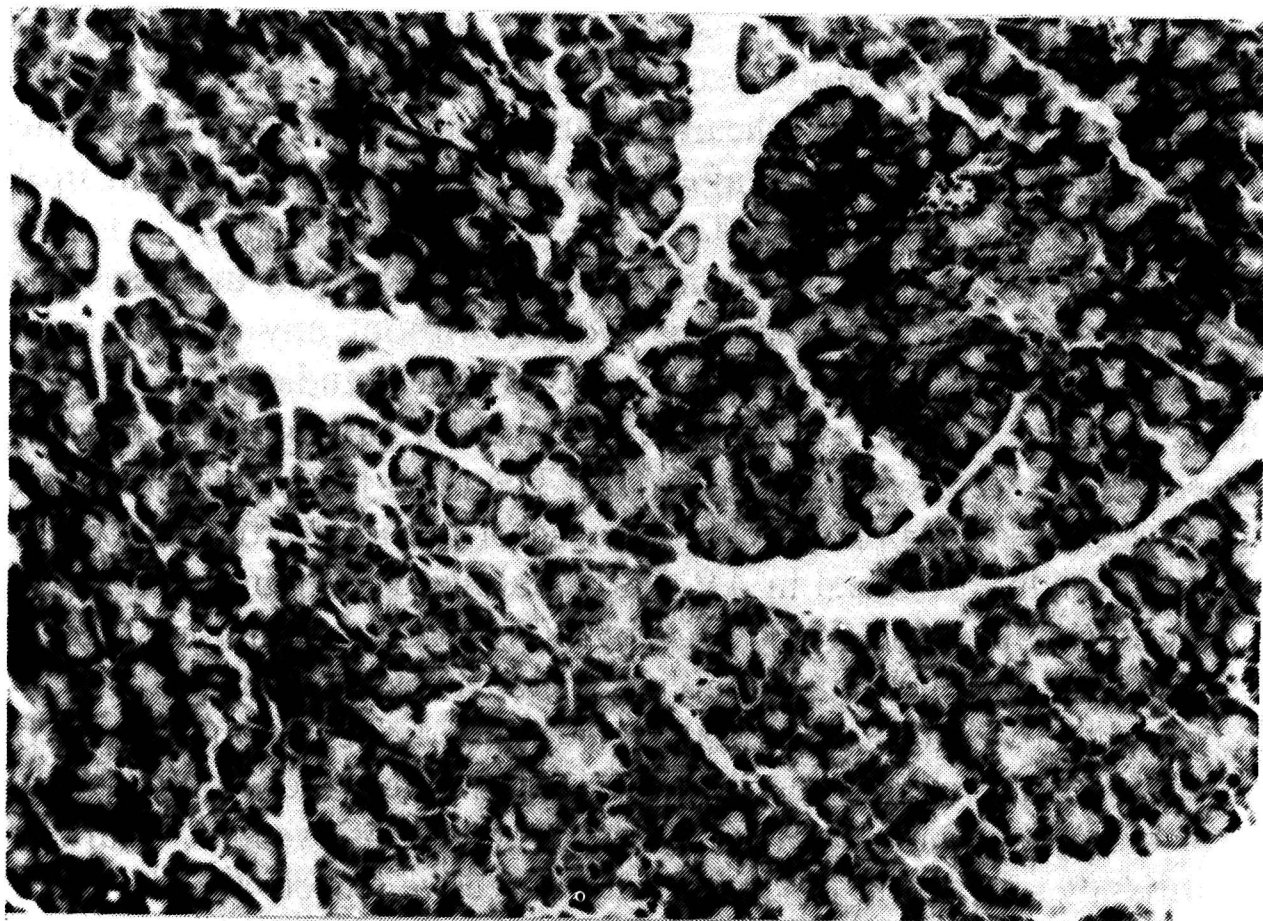


Fig. 5. Caerulein-induced pancreatitis — 14 days, treated with iloprost 1 $\mu\text{g}/\text{kg}$ b.w. t.i.d. (AP/I 14 days). Inflammatory infiltration and oedema is virtually absent. Minimal degenerative changes of acinar cells. Evident improvement in comparison to AP rats treated with I during 7 days (AP/I 7 days) as well as to 14-days AP group without treatment (AP 14 days). Mag. $\times 160$.

After 14 days quite a large amount of eosinophilic granules within acinar cells were found in AP group. Small number of inflammatory cells was also present (*Fig. 4*). In AP group treated with I during 14 days the inflammatory infiltration was virtually absent and the number of eosinophilic granules in acinar cells was increased as compared to AP untreated rats (*Fig. 5*).

DISCUSSION

In our previous study (2) it was found, that DNA content returned to control values 13 days after AP induction. Thus, at present study we have chosen 7 and 14 days as time points for trophic parameters investigation. Our results indicate that even 7 days after the onset of acute pancreatitis the signs of pancreas destruction are still present. Iloprost treatment during seven days resulted in the significant increase of DNA content in AP rats. This hyperplastic effect seen in the first phase of regeneration was not observed after 14 days of treatment. Pancreatic trophic parameters of AP untreated rats 14 days after the onset of pancreatitis induction reached control values, supporting that spontaneously occurring regeneration after acute pancreatitis was completed. The prostacyclin analogue given during 14 days after the

induction of AP showed only slight tendency to increase the trophic parameters over values found in untreated AP rats.

The iloprost treatment of healthy rats during 7 days also resulted in pancreatic hyperplasia, however after prolongation of the treatment up to 14 days this effect was not observed.

We have not yet any explanation for such DNA content increases (in both groups of rats treated with I during 7 days) without any changes in RNA content but similar effect was also found in other studies (22, 23).

Prostaglandins, known cytoprotective agents for gastrointestinal tract have been used also in the treatment of AP (24). The most promising prostaglandin derivative seems to be prostacyclin, because of broad spectrum of biological actions, potentially beneficial in AP. PGI₂ stabilized the pancreatic lysosomal membranes in bile-induced AP in dogs (7). PGI₂ and iloprost increased survival rate of rats with taurocholate pancreatitis (25, 26).

Iloprost has five times longer half-time in blood circulation than PGI₂ (27) and possess a wide range of potentially beneficial activities in AP. It protects the lysosomal membrane integrity, as it was observed in taurocholate pancreatitis (9). It exerts a direct cytoprotective effect as was found in experimental ulcerogenesis, at a dose of 1 µg/kg b.w. (28). Iloprost also improves microcirculation and the treatment with this agent could counteract the imbalance of PGI₂/TXA₂ occurring in AP resulting in the improvement of histopathological changes in AP (25).

The trophic effect of different prostaglandins on gastrointestinal mucosa has been described. Fitzpatrick et al. (12) found that 16, 16-dimethyl-PGE₂ (ig) increased ornithine decarboxylase (ODC) activity by 2 hr, and chronic administration of PGE₂ up to one week increased RNA, DNA and protein content in the rat duodenal mucosa in association with the elevation of ODC activity. PGI₂ analogue (TRY-200) was also shown to stimulate ODC activity within duodenum, jejunum and ileum to similar extent as a refeeding (potent trophic factor), however endogenous prostaglandins seems to play no role in feeding-associated induction of ODC activity (29).

Short term prostacyclin treatment evoked significant elevation of DNA content in gastric fundic mucosa, while RNA and protein remained unchanged (22). Long term treatment (40 and 80 days) with PGI₂ (po) results in thickening of gastric fundic mucosa and was connected with the cellular hyperplasia as DNA content increased significantly in comparison to untreated rats (protein and RNA contents remained unchanged) (23). These results partly correspond to the effect of PGI₂ analogue treatment on pancreas of healthy and AP rats obtained by us.

In our study iloprost treatment during 7 days resulted in pancreas hyperplasia of healthy and AP rats. The lack of hyperplastic or evident trophic influence of iloprost after 14 days in both groups could result from the pancreas

„adaptation” to this treatment. This adaptation may result from the receptor desensitization due to often, prolonged drug application, analogically as has been shown for decreased platelet sensitivity to PGI₂ infusion during 7 days (decreased receptor affinity with increased number of binding sites) (30). Modesti et al. (31) have found that after short term of iloprost infusion (6 h) the reversible reduction of platelet PGI₂ sensitivity occurred, and was related to a contemporary down-regulation of PGI₂ receptors. We did not find in the literature the explanation for the exact mechanisms of PGI₂ influence on pancreas growth/regeneration.

In view of fact, that PGI₂ analogue exerts potentially beneficial effect in acute pancreatitis, the possible mechanisms of its influence on pancreas growth and regeneration require to be investigated.

REFERENCES

1. Elsasser H-P, Lutcke H, Kern HF. Acinar and duct cell replication and regeneration. In Go VLW, Gardner JD, Brooks FP, Lebenthal E, DiMagno ED, Scheel GA, (eds.). The exocrine pancreas biology, pathobiology and diseases. New York: Raven, 1986: pp. 45—53.
2. Jurkowska G, Grondin G, Masse S, Morisset J. Soybean trypsin inhibitor and cerulein accelerate recovery of cerulein-induced pancreatitis in rats. *Gastroenterology* 1992; 102: 550—562.
3. Van Ooijen B, Kort WJ, Zijlstra FJ, Vincent JE, Wilson JHP, Westbroek DL. Prostanoid imbalance in experimental acute necrotizing pancreatitis in rats. *Scand J Gastroenterol* 1988; 23: 193—198.
4. Buscail L, Bussenot I, Bouisson M et al. Protective effect of micoprostol, a synthetic prostaglandin E₁ analog on caerulein-induced acute pancreatitis in rats. *Pancreas* 1990; 5: 171—176.
5. Pozsar J, Berger Z, Simon K, Kovacsai A, Marosi E, Pap A. Biphasic effect of prostaglandin E₁ on the severity of acute pancreatitis induced by a closed duodenal loop in rats. *Pancreas* 1996; 2: 159—164.
6. Robert A, Lum JT, Lancaster C, Olafson AS, Kolbasa KP, Nezamis JE. Prevention by prostaglandin of caerulein-induced pancreatitis in rats. *Lab Invest* 1989; 60: 677—691.
7. Gabryelewicz A, Dlugosz J, Brzozowski J et al. Prostacyclin: effect on pancreatic lysosomes in acute experimental pancreatitis in dogs. *Mt Sinai J Med* 1983; 50: 218—224.
8. Manabe T, Hirano T, Ando K, Yotsumoto F, Tobe T. Effect of prostaglandin E₂ on cellular, lysosomal and mitochondrial fragility in caerulein-induced pancreatitis in rats. *Hepato-Gastroenterol* 1993; 40: 463—466.
9. Dlugosz JW, Wroblewski E, Poplawski C, Gabryelewicz A. Does iloprost affect acute pancreatitis with ethanolic coetiology in rats?. *Gut* 1995; 37 (suppl. 2): A28 (334).
10. Homma T, Malik KU. Effect of secretin and caerulein in canine pancreas; relation to prostaglandins. *Am J Physiol* 1983; 224: G 660—667.
11. Tremoli E, Colli S, Paoletti R. Mode of action of PGI₂ and its stable derivatiae on platelets and leucocytes. *Thromb Res* 1990; suppl. 11: 33—42.
12. Fitzpatric LR, Gaginella TS, Haddox MK, Johnson LR. Prostaglandin-mediated trophic affects on the rat duodenum: The role of polyamines. *Proc Soc Exp Biol Med* 1988; 189: 201—205.

13. Uribe A, Johansson C. Initial kinetic changes of prostaglandin E₂-induced hyperplasia of the rat small intestinal epithelium occur in the villous compartments. *Gastroenterology* 1988; 94: 1335—1342.
14. Andries PG, Whitefield JF, Armato U. Stimulation of DNA synthesis and mitosis of hepatocytes in primary cultures of neonatal rat liver by arachidonic acid and prostaglandins. *Exp Cell Res* 1981; 134: 265—272.
15. Rixon RH, Whitefield JF. An early mitosis determining event in regenerating rat liver and its possible mediation by prostaglandins or thromboxane. *J Cell Physiol* 1982; 113: 281—288.
16. Mainz DL, Black O, Webster PD. Hormonal control of pancreatic growth. *J Clin Invest* 1973; 52: 2300—23004.
17. Volkin E, Cohn WE. Estimation of nucleic acids. *Biochem Anal* 1954; 91: 287—303.
18. Munro HF, Fleck A. Recent developments in the measurements of nucleic acids in biological materials. *Analyst* 1966; 91: 78—88.
19. Bernfeld P. Amylase α and β . *Methods Enzymol* 1955; 1: 149—158.
20. Hummel BC. A modified spectrophotometric determination of chymotrypsin, trypsin and thrombin. *Can J Biochem* 1959; 37: 1393—1399.
21. Lowry OH, Rosenbrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem* 1951; 193: 59—70.
22. Varro V, Balint GA, Karacsony G, Nafradi J. Prostacyclin and gastroduodenal protein synthesis in normal and ulcerated gastric mucosa of the rat. European Gastro Club, Erlangen, 26—29 Oct. 1983. *Acta Hepato-Gastroent* 1984; 31: 96.
23. Balint GA, Karacsony G, Varro V. The effect of long-term prostacyclin treatment on the protein, DNA and RNA content of rat gastric fundic mucosa. *Agents Action* 1985; 16: 404—406.
24. Stanfield NJ, Kakkar VV. Prostaglandins and acute pancreatitis: experimental and clinical studies; *Br J Surg* 1983; 70: 573—576.
25. Van Ooijen B, Kort WJ, Tinga CJ, Wilson P. Significance of tromboxane A₂ and prostaglandin I₂ in acute necrotising pancreatitis in rats. *Dig Dis Sci* 1990; 35: 1078—1084.
26. Jurkowska G, Dlugosz J, Gabryelewicz A, Andrzejewska A. The time course of liver DNA and RNA alterations in acute experimental pancreatitis in rats — a possible mechanism of prostacyclin (PGI₂) protection. *Hepato-Gastroenterol* 1989; 36: 249—254.
27. Krause W, Nieuweboer B. Pharmacokinetics and biotransformation of the prostacyclin analogue, ZK 36 374. III. Development of a radioimmunoassay and its application to the pharmacokinetics of ZK 36 374 in the rat. *Prostagl Leucotr Med*. 1983; 10: 289—299.
28. Zengil H, Onuk E, Ercan ZS, Turker RK. Protective effect of iloprost and UK 38 485 against gastric mucosal damage induced by various stimuli. *Prostagland Leuk Med* 1987; 30: 61—67.
29. Kuwayama H, Naito T. Effects of prostaglandins on ornithine decarboxylase activity in rat small intestine. *Dig Dis Sci* 1993; 38: 1087—1090.
30. Steurer G, Fitscha P, Sinzinger H. The platelet rebound phenomenon during PGI₂-infusion occurs at the receptor level. *Folia Haematol Leipzig* 1988; 4, S: 435—438.
31. Modesti PA, Fortini A, Poggesi L, Boddi M, Abbate R, Gensini GF. Acute reversible reduction of PGI₂ platelet receptors after iloprost infusion in man. *Thromb Res* 1987; 48: 663—669.

Received: May 6, 1996

Accepted: September 9, 1996

Author's address: G. Jurkowska, Department of Gastroenterology, University Medical School, M.C. Sklodowskiej 24 a, 15-276 Bialystok, Poland.