

## **ANALYSIS OF INDEL POLYMORPHISM OF THE *PRNP* GENE IN WATER BUFFALO, *BUBALUS BUBALIS***

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**Abstract.** The aim of this study was to examine insertion-deletion polymorphisms (one being 12bp-long in intron 1 and second being 23bp-long in the promoter) in water buffalo. The blood samples were collected from two water buffalo herds (40 individuals). DNA was isolated using a commercial Master Pure DNA Purification Kit. After conducting 2 PCRs, and then electrophoreses in 4% agarose gel, it was found that there was no polymorphism in either *PRNP* 12 ins/del the *PRNP* 23 ins/del in the analyzed group of animals. All individuals were characterized by the ins/ins genotype for both polymorphisms. Because of the fact that four different genotypes were found in the Anatolian breed and only one genotype was identified in the breed examined in the present study, more breeds of this species should be included in further research.

**Key words:** *PRNP* gene, DNA polymorphism, prion disease, water buffalo

### **INTRODUCTION**

Spongiform encephalopathies are fatal diseases that attack the central nervous system of mammals. The greatest influence on their occurrence is exerted by environmental factors, although some individuals will never develop the disease since they exhibit genetic resistance.

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The main role in the pathogenesis of prion diseases is played by the endogenous glycosyl prion protein, containing no nucleic acids, but having the ability to multiply and infect. There are two forms of the prions – one not causing disease (PrPC) and the invasive form (PrPSc) [Momcilovic and Rasooly 2000, Brunelle et al. 2007].

These diseases include, among others, bovine spongiform encephalopathy (BSE), which is mainly characterized by irreversible changes in the brain, where the tissue transforms into a spongy structure. The first symptoms occur 4–5 years after infection, and death occurs within the next few months. The prion protein is encoded by the *PRNP* gene. In humans [Windle et al. 1996], cattle [Juling et al. 2006], mice [Carlson et al. 1988] and sheep [Hunter et al. 1997] susceptibility to prion diseases, and the time in which they develop, depend on the variant of the gene. The *PRNP* gene is normally expressed in the nervous tissue and tissues of other organs. It is always present in one copy in an organism [Liberski and Bratosiewicz 1996]. In cattle, it is located in the long arm of chromosome 13 and consists of 3 exons, of which only one (third) is expressed [Yoshimoto et al. 1992, Horiuchi et al. 1998].

Of 60 known mutations in the bovine *PRNP* gene, two are located in the regulatory region and play important roles. These are insertion-deletion polymorphisms, one 12bp-long in intron 1 [Juling et al. 2006] and second one 23bp-long in the promoter region [Sander et al. 2004, Haase et al. 2007], which determine the occurrence of two homozygous genotypes (ins/ins) and (del/del) and a heterozygous genotype (ins/del). These polymorphisms result in changes in the gene expression. Heterozygotes and homozygotes with deletion in both the first and the second polymorphism are highly susceptible to BSE, although the magnitude of this relationship depends on the breed [Juling et al. 2006, Strychalski et al. 2012, Zhao et al. 2012].

The aim of this study was to examine insertion-deletion polymorphisms (one being 12bp-long in intron 1 and second being 23bp-long in the promoter) in water buffalo.

## MATERIAL AND METHODS

The research material comprised the water buffalo blood from two herds. The first herd was kept in Poland, while the second one in Germany. For the first herd, the blood samples were collected from the external jugular vein into tubes with anticoagulant from 29 females (half of the herd) in October 2008. For the German herd (11 individuals), the material was collected from the tail vein in August 2010.

DNA was isolated using a commercial MasterPure DNA Purification Kit. This was followed by the two PCRs with the primers suggested by Juling et al. [2006] –

F 5'-CCTGTTGAGCGTGCTCGT-3' and R 5'-ACCTGCGGCTCCTCTACC-3' for ins/del 23bp (191bp/168bp) as well as F 5'-GGAAGTCACGTGAAGGCACT-3' and R 5'-CAAAGAGTTGGACAGGCACA-3' for the ins/del 12bp (215bp/203 bp). Thirty-two cycles were carried out under the following thermal conditions: denaturation at 94°C for 45 s (pre-denaturation for 5 min), primers annealing at 58°C for 45 s, elongation at 72°C for 45 s (in the last cycle for 7 min). The obtained products were separated by electrophoresis on a 4% agarose gel with ethidium bromide and visualized under UV light.

## RESULTS AND DISCUSSION

In the analyzed group of animals, there was no polymorphism in either *PRNP* 12 ins/del or the *PRNP* 23 ins/del. All individuals were characterized by the ins/ins genotype for both polymorphisms. Oztabak et al. [2009] examined 106 individuals of river buffalo of the Anatolian breed from various regions of Turkey. They found

Table 1. Comparison of the allele frequencies of the *PRNP* ins/del polymorphisms in different cattle breeds

Tabela 1. Porównanie częstości alleli z *PRNP* ins/del polimorfizmów u różnych ras bydła

Animal Rasa	Allele frequencies of 12 bp and 23 indel polymorphisms Częstość alleli 12 bp 23 indel polimorfizmów				Reference Piśmiennictwo
	12 bp in	12 bp del	23 bp in	23 bp del	
Korean Holstein	0.39	0.61	0.30	0.70	Jeong et al. 2006
Korean Hanwoo	0.44	0.56	0.40	0.60	
German Holstein	0.57	0.53	0.48	0.62	Juling et al. 2006
German Brown	0.86	0.14	0.65	0.35	
Fleckvieh	0.38	0.62	0.32	0.68	
USA beef cattle	0.37	0.63	0.26	0.74	Clawson et al. 2006
USA dairy cattle	0.63	0.47	0.38	0.62	
Polish Holstein	0.46	0.54	0.37	0.63	Czarnik et al. 2007
Abderdeen Angus	0.44	0.56	0.27	0.73	Kerber et al. 2008
Charolais	0.42	0.58	0.32	0.68	
Franqueiro	0.67	0.33	0.36	0.64	
Sout Anatolian Red	0.69	0.31	0.36	0.64	Ün et al. 2008
East Anatolian Red	0.72	0.28	0.40	0.60	
Anatolian Grey	0.80	0.20	0.62	0.38	
Japanese Black	0.42	0.58	0.23	0.77	Msalya et al. 2011
Japanese Brown	0.47	0.53	0.44	0.77	
Water Buffalo	1.00	–	1.00	–	Our research Badania własne

the presence of four different genotypes (ins12/del23, del12/del23, ins12/ins23, del12/ins23), of which the ins12/ins23 genotype had the highest frequency (0.86). Other genotypes were characterized by much lower frequencies: 0.08, 0.05 and 0.1 for ins12/del23, del12/del23 and ins12/del23, respectively. These results are similar to those obtained in the present study, in which all the individuals were homozygotes, whereas in cattle, disadvantageous alleles predominate, although their frequency depends on the breed. Most of the cattle breeds studied so far are presented in Table 1.

Sander et al. [2004, 2005] demonstrated that the ins/del23 polymorphism is strongly associated with susceptibility/resistance to BSE. However, in later studies Juling et al. [2006] and Kashkevich et al. [2007] showed the same effect of the ins/del12 polymorphism. The results obtained by the above-mentioned authors can also be applied to the water buffalo, because these animals belong to the same family as cattle.

Due to the fact that four different genotypes were found in the Anatolian breed and only one genotype was identified in the breed examined in the present study, more breeds of this species should be included in further research.

## REFERENCES

- Brunelle B.W., Hamir A.N., Baron T., Biacabe A.G., Richt J.A., Kunkle R.A., Cutlip R.C., Miller J.M., Nicholson E.M., 2007. Polymorphisms of the prion gene promoter region that influence classical bovine spongiform encephalopathy susceptibility are not applicable to other transmissible spongiform encephalopathies in cattle. *J. Anim. Sci.* 85, 3142–3147.
- Carlson G.A., Goodman P.A., Lovett M., Taylor B.A., Marshall S.T., Peterson-Torchia M., Westaway D., Prusiner S.B., 1988. Genetics and polymorphism of the mouse prion gene complex: control of scrapie incubation time. *Mol. Cell. Biol.* 8, 5528–5540.
- Clawson M.L., Heaton M.P., Kele J.W., Smith T.P.L., Harhay G.P., Laegreid W.W., 2006. Prion gene haplotypes of U.S. cattle. *BMC Genet.* 7, 51.
- Czarnik U., Zabołewicz T., Strychalski J., Grzybowski G., Bogusz M. and Walawski K., 2007. Deletion/insertion polymorphism of the prion protein gene (*PRNP*) in Polish Holstein-Friesian cattle. *J. Appl. Genet.*, 48, 69–71.
- Haase B., Doherr M.G., Seuberlich T., Drögemüller C., Dolf G., Nicken P., Schiebel K., Ziegler U., Groschup M.H., Zurbriggen A., Leeb, T., 2007. *PRNP* promoter polymorphisms are associated with BSE susceptibility in Swiss and German cattle. *BMC Genet.*, 8 (1), 15.
- Hourichi M., Ishiguro N., Nagasawa H., Toyoda Y., Shinagawa M., 1998. Genomic structure of the bovine PrP gene and complete nucleotide sequence of bovine PrP cDNA. *Anim. Genet.* 29, 37–40.
- Hunter N., Cairns D., Foster J.D., Smith G., Goldmann W., Donnelly K., 1997. Is scrapie solely a genetic disease? *Nature* 386, 137.

- Jeong B.H., Lee Y.J., Kim N.H., Carp R.I., Kim Y.S., 2006. Genotype distribution of the prion protein gene (*PRNP*) promoter polymorphism in Korean cattle. *Genome* 49, 1539–1544.
- Juling K., Schwarzenbacher H., Williams J. L., Fries R., 2006. A major genetic component of BSE susceptibility. *BMC Biol.*, 4:33, doi:10.1186/1741-7007-4-33.
- Kashkevich K., Humeny A., Ziegler U., Groschup M.H., Nicken P., Leeb T., Fischer C., Becker C–M., Schiebel K., 2007. Functional relevance of DNA polymorphisms within the promoter region of the prion protein gene and their association to BSE infection. *FASEB J.*, 21(7), 1547–1555.
- Kerber A.R., Hepp D., Passos D.T., Weimer T.A., 2008. Polymorphisms of two indels at the *PRNP* gene in three beef cattle herds. *Biochem. Genet.* 46, 1–7.
- Liberski P.P., Bartosiewicz J., 1996. Pasażowalne amyloidozy mózgowie czyli choroby wywołane przez priony: czy struktura czynnika scrapie jest już rzeczywiście znana? [Transmissible cerebral amyloidoses as diseases caused by prions: is the Scrapie factor structure really known?] *Post. Biochem.* 42 (4), 320–330 [in Polish].
- Momicilovic D., Rasooly A., 2000. Detection and analysis of animal materials in food and feed. *J. Food Protect.* 63 (11), 1602–1609.
- Msalya G., Shimogiri T., Ohno S., Okamoto S., Kawabe K., Minezawa M., Yoshizane Maeda Y., 2011. Evaluation of *PRNP* expression based on genotypes and alleles of two indel loci in the Medulla Oblongata of Japanese Black and Japanese Brown cattle. *PLoS ONE* 6(5), 18787.
- Oztabak K., Ozkan E., Soysal I., Paya I., Un C., 2009. Detection of prion gene promoter and intron1 indel polymorphisms in Anatolian water buffalo (*Bubalus bubalis*). *J. Anim. Breed Genet.* 126 (6), 463–467.
- Sander P., Hamann H., Drogemuller C., Kashkevich K., Schiebel K., Leeb T., 2005. Bovine prion proteingene (*PRNP*) promoter polymorphisms modulate *PRNP* expression and may be responsible for differences in bovine spongiform encephalopathy susceptibility. *J. Biol. Chem.* 280, 37408–37414.
- Sander P., Hamann H., Pfeiffer I., Wemheuer W., Brenig B., Groschup M.H., Ziegler U., Distl O., Leeb T., 2004. Analysis of sequence variability of the bovine prion protein gene (*PRNP*) in German cattle breeds. *Neurogenetics* 5, 19–25.
- Strychalski J., Czarnik U., Tadeusz Zabołewicz J., 2012. Abnormal segregation of alleles and haplotypes at the polymorphic site of the *PRNP* gene within promoter and intron 1 regions in Polish Holstein–Friesian cattle. *Biochem. Genet.* 50, 520–528.
- Ün C., Oztabak K., Ozdemir N., Tesfaye D., Mengi A., Schellander K., 2008. Detection of bovine spongiform encephalopathy related prion protein gene promoter polymorphism in Turkish local cattle. *Biochem. Genet.* 46, 820–827.
- Windl O., Dempster M., Estibeiro J.P., Lathe R., de Silva R., Esmonde T., Will R., Springbett A., Campbell T.A., Sidle K.C.L., Palmer M.S., Collinge J., 1996. Genetic basis of Creutzfeldt–Jakob disease in the United Kingdom: a systematic analysis of predisposing mutations and allelic variation in the *PRNP* gene. *Hum. Genet.* 98, 259–26.
- Yoshimoto J., Toshiyuki I., Naotaka I., Motohiro H., Masakatu I., Morikazu S., 1992. Comparative sequence analysis and expression of bovine PrP gene in mouse L-929 cells. *Virus Genes.* 6 (4), 343–356.

Zhao H., Liu L.L., Du S.H., Wang S.Q., Zhang Y.P., 2012. Comparative Analysis of the Shadoo Gene between Cattle and Buffalo Reveals Significant Differences. PLoS ONE 7(10), 46601.

## **ANALIZA POLIMORFIZMU TYPU INDEL GENU *PRNP* U BAWOŁÓW, *BUBALUS BUBALIS***

**Streszczenie.** Celem niniejszego badania była analiza polimorfizmów typu insercja-delecja (jednego o długości 12 pz w intronie I, drugiego o długości 23 pz w promotorze genu). Próbki krwi zostały pobrane od bawołów z dwóch stad (w sumie od 40 osobników). DNA zostało wyizolowane przy użyciu zestawu Master Pure DNA Purification Kit. Po wykonaniu dwóch reakcji PCR oraz przeprowadzeniu elektrofo-rezy na żelu agarozowym o stężeniu 4%, stwierdzony został brak polimorfizmu za-równy we fragmencie *PRNP* 12 ins/del, jak i we fragmencie *PRNP* 23 ins/del u ana-lizowanych osobników. U wszystkich osobników stwierdzono genotypy ins/del dla obydwu polimorfizmów. Ponieważ u rasy anatolijskiej zidentyfikowano cztery rodzaje genotypów, a w obecnie badanej rasie jedynie jeden rodzaj genotypu, konieczne jest przeanalizowanie obecności polimorfizmu także u innych ras.

**Słowa kluczowe:** gen *PRNP*, polimorfizm DNA, choroba prionowa, bawół

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