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Community structure of macrozoobentos on artificial reef made from reef rubble and split rock in Tunda Island, province of Banten, Indonesia

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ABSTRACT

Tunda Island has underwater beauty that has the potential to be developed into a marine tourism object. One important part of this is the fringing reef around the island. Unfortunately, this has been severely degraded. This study aims to determine if reef rubble and split rock artificial reefs have the ability to be a living space for macrozoobenthos and to measure the potential community structure. The research was conducted in August 2018 - November 2018 in the eastern part of Tunda Island, and included monitoring the initial conditions, artificial reef creating and positioning, gathering and analyzing macrozoobenthos data. The macrozoobenthos data collection was carried out from September to November 2018 as 4 observations, and was obtained using the Belt Transect method. In the analysis, the community structure data retrieved was divided into 3 categories: diversity, abundance and dominance. The diversity index obtained ranged from 0.43 – 7.65, meaning that it is of low to moderate diversity. The abundance of macrozoobenthos ranged from 1 - 18 individuals /m² on the rubble type artificial reef and 2.13 – 4.26 individuals /m² on the split type artificial reefs, with the most common class being gastropods. On rubble type artificial reefs, 14 macrozoobenthos genera were found to be common, while 13 genera were observed in the split type. The genus found was *Culcita* sp., *Chicoreus* sp., *Diadema* sp., *Chelidonura* sp., *Thrombus* sp., *Chromodoris* sp., *Cymatium* sp., *Trochus* sp., *Terebra* sp., *Dardanus* sp., *Malea* sp., *Oliva* sp., *Opheodosoma* sp., *Actinopyga* sp., *Conus* sp., *Enoplometopus* sp., *Cypracea* sp., *Lambis* sp., and *Phylidia* sp. The dominance index value was in the range of 0.18-1.00 or low to high on both rubble and split types

Keywords: Macrozoobenthos, Split rock, Tunda Island, Reefs Rubble, Artificial Coral Reef Community Structure

1. INTRODUCTION

Tunda Island, which is surrounded by the Java Sea, is the outermost region of the Serang Regency, and is geographically located at 106°50'00" - 105°51'51" BT and 5°56'15" - 5°59'00" LS. The main village is Warga Sara and it is part of Tirtayasa District. Tunda Island has underwater beauty that has the potential to be developed into a marine tourism object. An important aspect of this is its fringing reef. In 2016, conditions were discovered where dead coral or rubble was piled up and scattered around the island, even making natural breakwaters in the eastern part of the island. Moreover, the condition of the coral reefs in the southern part of the island of Tunda was assessed as poor. In the coral reef ecosystem, there are associated biota such as reef fish and macrozoobenthos. Macrozoobenthos have low motility, so these are often used as environmental indicators. One way to rehabilitate the macrozoobenthos ecosystem is to create and place artificial coral reefs. Artificial reefs are man-made habitats placed on the bottom of waters and are usually made from heaps of materials such as used tires, cement or concrete molds and bamboo. Because materials like split rock and used tires are difficult to find on Tunda Island, the coral detritus on the island have been used for artificial coral reef construction. Artificial coral reefs can mimic some of the characteristics of natural reefs so that they can attract species of marine organisms to live and settle upon them. Hence, functional artificial reefs can provide new habitat for macrozoobenthos and can reduce waves action so as to provide a coastal safeguard. Macrozoobenthos response to changes in environmental conditions come in the form of changes in the structure of abundance, diversity and biomass. Therefore, by establishing macrozoobenthos community structure on these artificial reefs and comparing this with that associated with natural coral reefs in the area, a benchmark can be ascertained so as to reveal the success of artificial reef establishment.

2. METHODS

The research location was in the eastern waters of Tunda Island, Banten (Figure 1). The research was conducted in August 2018 - November 2018, including location determination, measurement of physical and chemical parameters of the water, data collection on macrozoobenthos community structure and data analysis. The created artificial coral reefs were lowered on August 12, 2018, and macrozoobenthos data was collected four times: September 2, September 23, October 14 and November 18, 2018, at depths between 3 to 6 m.

The methods used in this study were the survey method and Purposive Sampling. The data points were carefully selected to exclude the influence of the surrounding natural coral reefs. At each station, 6 pieces of artificial reef were deployed (3 pieces of rubble composition and 3 pieces of Split composition) with a minimum distance of 2 to 5m between the artificial reef components (Figure 2). At each research station, data was collected at 10:00 WIB for 3 months every 3 weeks. Before the artificial reef was installed, observations were made of the initial conditions. Measurement of physical parameters was taken in the form of temperature, current velocity and turbidity. The chemical parameters of water quality taken were DO (dissolve oxygen), salinity and pH. Research on the macrozoobenthos community was carried out using the Belt Transect Method. Herein, macrozoobenthos abundance is based on the number of Macrozoobenthos found with a 2 m radius of an observation station. Macrozoobenthos abundance can be calculated by formula:

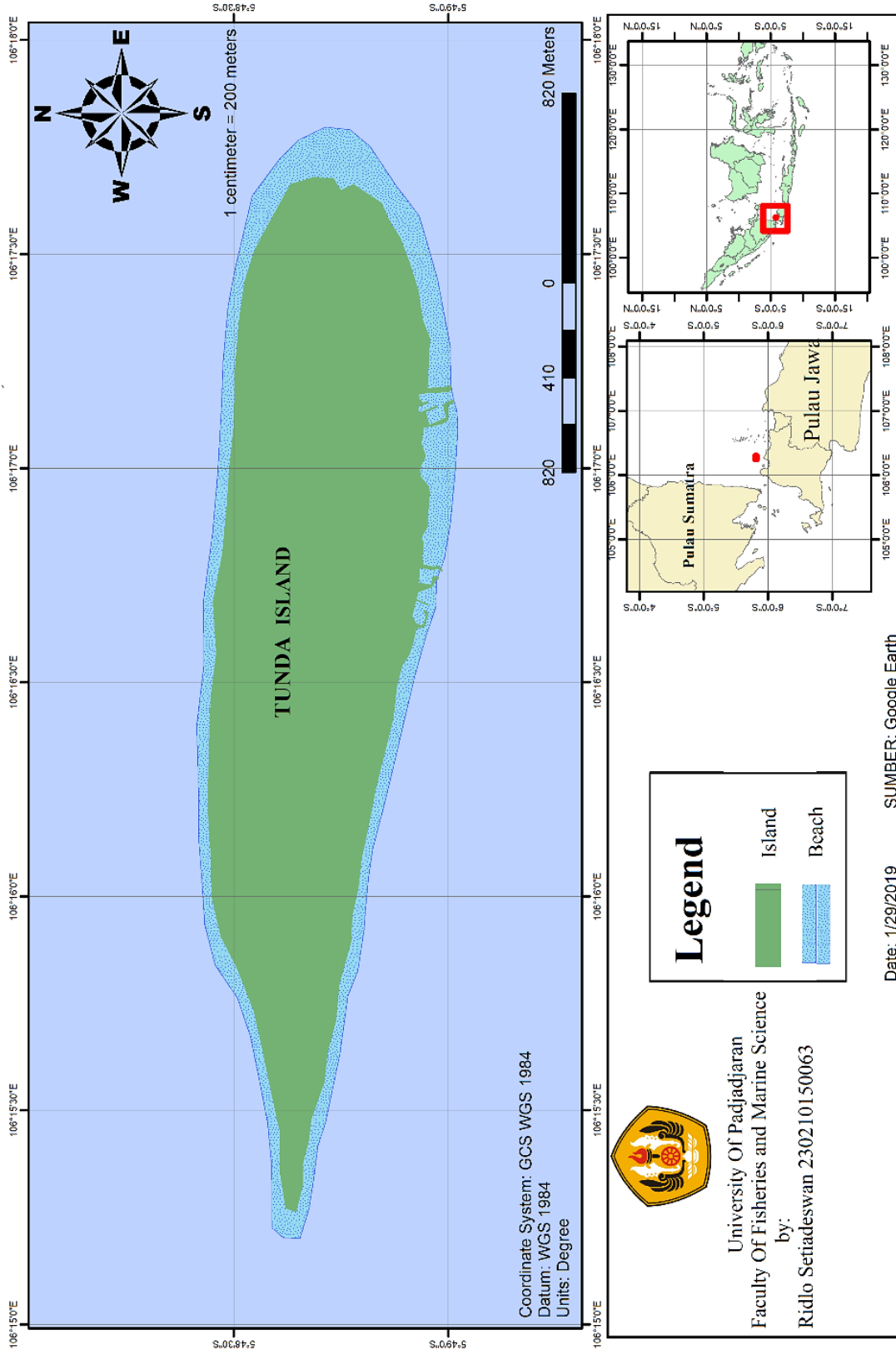


Figure 1. Research location.

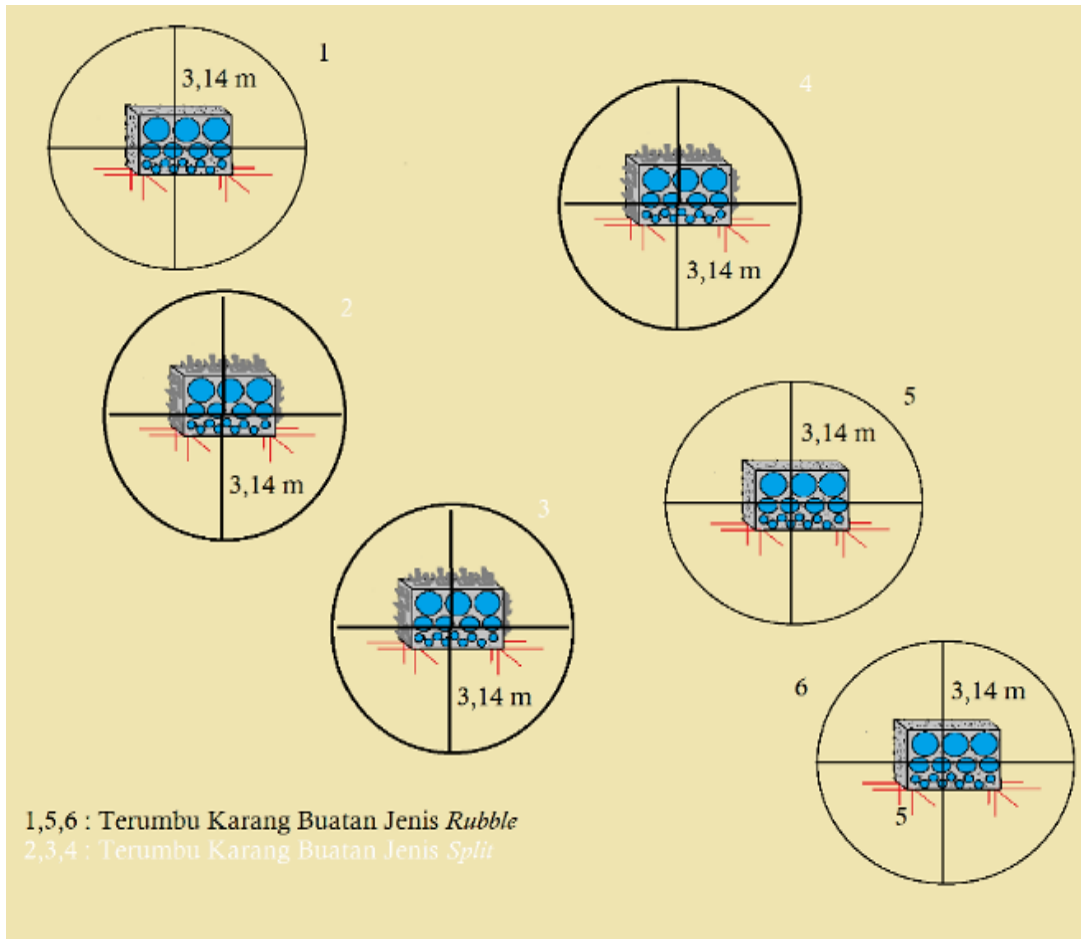


Figure 2. Illustration of Artificial Reef Placement

The Diversity Index is used to obtain a mathematical description of the organism's population. This aims to facilitate the analysis of information on the number of individuals of each species within a community. Macrozoobenthos diversity is calculated by the Shannon-Wiener Index using the following formula:

$$K = \frac{\text{total individual}}{\text{area of oservation}}$$

where:

K = Individuals richness (ind/m²)

The Diversity Index is used to get a description of the organism's population mathematically. This aims to facilitate the analysis of information on the number of individuals in each species in a community. Macrozoobenthos diversity is calculated by the Shannon-Wiener Index using the following formula:

$$H' = -\sum_{i=1}^n p_i \ln p_i$$

Information:

H' = Shannon-Wiener Index

p_i = Comparison between the number of i (n_i) individual species; (i : 1,2, .. n) with the total number of individuals (N)

The Macrozoobenthos diversity index category is:

$H' < 1$ = small diversity

$1 < H' < 3$ = medium diversity

$H' > 3$ = high diversity

To measure the dominance of a type used the Simpson Dominance Index as follows:

$$D = (n_i / N)^2$$

Description:

D : Dominance Index

N_i : Number of i -type individuals

N : Number of individuals of all types

The range of the Dominance Index values are:

$0.00 < C < 0.30$: low dominance

$0.30 < C < 0.60$: moderate dominance

$0.60 < C < 1.00$: high dominance

3. DISCUSSION

3. 1. Water Conditions at the Research Site

The results of measurements of water quality (Table 1) show that the waters as assessed by the seawater quality standards for aquatic biota issued by the Ministry of Environment in Minister of Environment Decree No.51 of 2004 are in good condition.

3. 2. Macrozoobenthos Community Structure

The community structure of macrozoobenthos is observed in the form of abundance data, diversity index and macrozoobenthos dominance index.

3. 2. 1. Macrozoobenthos Diversity

On the artificial coral reefs, 19 genera of macrozoobenthos were found in the four observations. Genus found include *Calucita* sp., *Chicoreus* sp., *Diadema* sp., *Chelidonura* sp., *Thrombus* sp., *Chromodoris* sp., *Cymatium* sp., *Trochus* sp., *Terebra* sp., *Dardanus* sp., *Malea* sp., *Oliva* sp., *Opheodosoma* sp., *Actinopyga* sp., *Conus* sp., *Enoplometopus* sp., *Cyperaceae*

sp., *Lambis* sp. and *Phylidia* sp. A community is said to have high species diversity if there are many species, with individual numbers of each species relatively evenly distributed. If a community consists of only a few species with an uneven number of individuals, then the community has a low diversity index.

Table 1. Physical and Chemical Parameter Value of Aquatic Each Observation

Parameter	Observation data					Quality standards*
	1	2	3	4	5	
Temperature (°C)	27-28	29	29	29-30	29-30	28-30
pH	8	8	8	8	7-8	7-8,5
DO (mg/l)	5,4-5,6	5,1-5,6	3,8-4,2	4	8,3	>5
Brightness (%)	100%	100%	100%	100%	100%	>5m
Salinity (ppt)	33,5-34	32	31-32	32-33	32	33-34
Current Speed (m/detik)	0.31					-

Table 2. Diversity index

Data number	1	2	3	4
H' Rubble	0.69	0	1.83	1.61
H' split	0.00	0.95	1.23	1.83

In this study, we found one genera that was common, namely *chicoreus* spp. This is a mollusk phylum that is very successful in adapting to fluctuating conditions. Molluscs (Bivalvia and Gastropoda) and Polychaeta are common in the estuary benthic community.

3. 2. 2. Abundance of Macrozoobenthos

On the artificial coral reefs, both rubble and Split species were found to have an abundance of ± 0.78 ind / m² and ± 0.75 ind / m² in artificial coral cover of 2.78 m². The index values of each observation can be found in Table 3.

Table 3. Found macrozoobenthos abundance.

Observation	Type	Genus	Amount	Type	Genus	Amount
1	Rubble	<i>Culcita</i> sp.	1	Split	<i>Diadema</i> sp.	5
		<i>Chicoreus</i> sp.	1			
		Total	2		Total	5
2	Rubble	<i>Chelidonura</i> sp.	1	Split	<i>Diadema</i> sp.	3
					<i>Chicoreus</i> sp.	1
					<i>Strombus</i> sp.	1
		Total	1		Total	5
3	Rubble	<i>Chicoreus</i> sp.	3	Split	<i>Culcita</i> sp.	1
		<i>Culcita</i> sp.	2		<i>Chicoreus</i> sp.	6
		<i>Phylidia</i> sp.	1		<i>Terebra</i> sp.	1
		<i>Chromodoris</i> sp.	1		<i>Cymatium</i> sp.	1
		<i>Actinopyga</i> sp.	1		<i>Dardanus</i> sp.	1
		<i>Cymatium</i> sp.	1			
		<i>Trochus</i> sp.	1			
		Total	10		Total	10
4	Rubble	<i>Culcita</i> sp.	1	Split	<i>Cypracea</i> sp.	1
		<i>Malea</i> sp.	1		<i>Chicoreus</i> sp.	3
		<i>Oliva</i> sp.	1		<i>Lambis</i> sp.	1
		<i>Opheodosoma</i> sp.	3		<i>Malea</i> sp.	1
		<i>Actinopyga</i> sp.	9		<i>Phylidia</i> sp.	1
		<i>Canus</i> sp.	1		<i>Cymatium</i> sp.	1
		<i>Chicoreus</i> sp.	1		<i>Canus</i> sp.	2
		<i>Enoplometopus</i> sp.	1			
		Total	18		Total	10

In the 3rd round of observations, we found one species of the nudibranch order, *Chelidonura* spp. The presence of nudibranch is due to the emergence of nudibranch food on the artificial coral reefs, namely sponges. Almost all Nudibranchia members are carnivorous and predators of sponges, soft corals, anemones, sea pens, Bryozoa, ascidians, hydroids and other Nudibranch eggs. In addition, some species are parasitic.

3. 2. 3. Dominance Index

Dominance will occur in a community that has low diversity. In our study, the dominance index value approaches 0 (zero). This means that no type dominates. This shows that the macrozoobenthos habitat is viable. Dominance data can be seen in Table 4.

Table 4. Dominance index obtained

	Observation			
	1	2	3	4
<i>Rubble</i>	0.50	1.00	0.18	0.30
<i>Split</i>	1.00	0.44	0.40	0.18

The dominance that occurs in this study is in the low range because macrozoobenthos generally prefer sandy substrate types, whereas in this study area the dominant substrate type is rocky. Hence, the living conditions are less suitable. States that the type of substrate greatly influences the presence of macrozoobenthos, and puts forward that fine sand to muddy substrates are most suitable to promote the macrozoobenthos community.

4. CONCLUSIONS AND SUGGESTION

4. 1. Conclusions

On artificial coral reefs, 19 genera of macrozoobenthos were found during the study. Genus found include *Culcita* spp., *Chicoreus* sp., *Diadema* sp., *Chelidonura* sp., *Strombus* sp., *Chromodoris* sp., *Cymatium* sp., *Trochus* sp., *Terebra* sp., *Dardanus* sp., *Malea* sp., *Oliva* sp., *Opheodosoma* sp., *Actinopyga* sp., *Conus* sp., *Enoplometopus* sp., *Cypraea* sp., *Lambis* sp. and *Phylidia* sp.

The abundance of macrozoobenthos is not significantly different on either rubble and split types of coral reefs, with the abundance ranging from 0.85 - 7.65 individuals /2.34m² on artificial coral types of rubble and 2.41 – 7.48 individuals / 2,34 m² on split type artificial reefs, with the most common class being gastropods.

The dominance that occurs in this study is in the low range, this comes about because most types of macrozoobenthos generally live on sandy substrate types, whereas in the study area the substrate is rocky or hard. The dominance index values are in the range of 0.18-1.00 for both rubble and artificial split types

The value of the Macrozoobentos community structure on rubble-made coral reefs does not differ greatly from split types. This is because the water quality values do not differ. Over all, however, macrozoobenthos are more adaptable to rubble types of artificial coral than to reefs of the artificial split type.

4. 2. Suggestion

Further research needs to be done on various aspects of the macrozoobenthos community structure, and must be extended in time. In addition, more observation must occur so as to achieve better comparison between artificial and natural reef types. Periodic monitoring is also needed to preserve the coral reef ecosystem in Tunda Island.

References

- [1] Hughes TP, Baird AH, Bellwood DR, Card M, Connolly SR, Folke C, et al. Climate change, human impacts, and the resilience of coral reefs. *Science* 2003; 301(5635), 929–33.
- [2] Hoegh-Guldberg O, Mumby PJ, Hooten AJ, Steneck RS, Greenfield P, Gomez E, et al. Coral Reefs under Rapid Climate Change and Ocean Acidification. *Science* 2007; 318(5857), 1737–42.
- [3] Frieler K, Meinshausen M, Golly A, Mengel M, Lebek K, Donner SD, et al. Limiting global warming to 2°C is unlikely to save most coral reefs. *Nature Climate Change* 2013; 3(2): 165–70.
- [4] Donner SD, Skirving WJ, Little CM, Oppenheimer M, Hoegh-Guldberg O. Global assessment of coral bleaching and required rates of adaptation under climate change. *Glob Change Biol.* 2005; 11(12): 2251–65. 10.1111/j.1365-2486.2005.01073.x
- [5] Chollett I, Mumby PJ, Muller-Karger FE, Hu CM. Physical environments of the Caribbean Sea. *Limnol Oceanogr.* 2012; 57(4): 1233–44. 10.4319/lo.2012.57.4.1233
- [6] Indra, Asep Sahidin, Zahidah dan Yuli Andriani, Macrozoobenthos Community Structure in Cijulang River Pangandaran District, West Java Province, Indonesia. *World Scientific News* 128(2) (2019) 182-196
- [7] Syifa Hanifah, Asep Sahidin, Herman Hamdani, Lintang Permata Sari Yuliadi, Diversity of Chlorophyta on Karapyak Beach, Pangandaran, West Java Province, Indonesia. *World Scientific News* 117 (2019) 158-174
- [8] Gardner TA, Cote IM, Gill JA, Grant A, Watkinson AR. Long-term region-wide declines in Caribbean corals. *Science* 2003; 301: 958–60.
- [9] Jokiel PL, Coles SL. Response of Hawaiian and other Indo-Pacific reef corals to elevated temperature. *Coral Reefs.* 1990; 8: 155–62.
- [10] D'Croz L, Mate JL, Oke JE. Responses to elevated sea water temperature and UV radiation in the coral *Porites lobata* from upwelling and non-upwelling environments on the Pacific coast of Panama. *Bull Mar Sci.* 2001; 69(1): 203–14.

- [11] Aronson R, Precht W, Toscano M, Koltjes K. The 1998 bleaching event and its aftermath on a coral reef in Belize. *Marine Biology*. 2002;141 (3): 435–47.
- [12] Wooldridge S, Done T, Berkelmans R, Jones R, Marshall P. Precursors for resilience in coral communities in a warming climate: a belief network approach. *Marine Ecology Progress Series*. 2005; 295: 157–69.
- [13] Donner SD, Knutson TR, Oppenheimer M. Model-based assessment of the role of human-induced climate change in the 2005 Caribbean coral bleaching event. *Proceedings of the National Academy of Sciences of the United States of America* 2007; 104(13): 5483–8. 10.1073/pnas.0610122104
- [14] Oliver TA, Palumbi SR. Do fluctuating temperature environments elevate coral thermal tolerance? *Coral Reefs*. 2011; 30(2): 429–40. 10.1007/s00338-011-0721-y
- [15] Barshis DJ, Ladner JT, Oliver TA, Seneca FO, Traylor-Knowles N, Palumbi SR. Genomic basis for coral resilience to climate change. *Proceedings of the National Academy of Sciences* 2013; 110(4): 1387–92
- [16] Fadliyan R, Rachim, Mega L, Syamsudin, Indah Riyantini, Lintang P. S. Yuliadi, Influence of Indian Ocean Dipole Phenomena Towards the Eddy Variability in Southern Java Ocean. *World Scientific News* 132 (2019) 121-131
- [17] Pineda J, Starczak V, Tarrant A, Blythe J, Davis K, Farrar T, et al. Two spatial scales in a bleaching event: Corals from the mildest and the most extreme thermal environments escape mortality. *Limnol Oceanogr*. 2013; 58(5): 1531–45. 10.4319/lo.2013.58.5.1531
- [18] Castillo KD, Ries JB, Weiss JM, Lima FP. Decline of forereef corals in response to recent warming linked to history of thermal exposure. *Nature Climate Change*. 2012; 2(10): 756–60.
- [19] Carilli J, Donner SD, Hartmann AC. Historical temperature variability affects coral response to heat stress. *Plos ONE*. 2012; 7(3): e34418 10.1371/journal.pone.0034418
- [20] van Woesik R, Houk P, Isechal AL, Idechong JW, Victor S, Golbuu Y. Climate-change refugia in the sheltered bays of Palau: analogs of future reefs. *Ecol Evol*. 2012; 2(10): 2474–84. 10.1002/ece3.363
- [21] Fine M, Gildor H, Genin A. A coral reef refuge in the Red Sea. *Glob Change Biol*. 2013; 19(12): 3640–7. 10.1111/gcb.12356
- [22] Lirman D, Fong P. Is proximity to land-based sources of coral stressors an appropriate measure of risk to coral reefs? An example from the Florida Reef Tract. *Marine Pollution Bulletin*. 2007; 54(6): 779–91
- [23] Darling ES, Alvarez-Filip L, Oliver TA, McClanahan TR, Côté IM. Evaluating life-history strategies of reef corals from species traits. *Ecology Letters*. 2012; 15(12): 1378–86. 10.1111/j.1461-0248.2012.01861.x
- [24] Grime JP, Pierce S. *The evolutionary strategies that shape ecosystems*: John Wiley & Sons; 2012.

- [25] Darling ES, McClanahan TR, Côté IM. Life histories predict coral community disassembly under multiple stressors. *Global Change Biology*. 2013; 19(6): 1930–40. 10.1111/gcb.12191
- [26] Alvarez-Filip L, Dulvy NK, Côté IM, Watkinson AR, Gill JA. Coral identity underpins architectural complexity on Caribbean reefs. *Ecological Applications*. 2011; 21(6): 2223–31.
- [27] Greenstein B, Curran H, Pandolfi J. Shifting ecological baselines and the demise of *Acropora cervicornis* in the western North Atlantic and Caribbean Province: a Pleistocene perspective. *Coral Reefs*. 1998; 17(3): 249–61
- [28] Buglass S, Donner SD, Alemu JB. A study on the recovery of Tobago's coral reefs following the 2010 mass bleaching event. *Marine Pollution Bulletin*. 2016; 104(1): 198–206
- [29] Van Woesik R, Sakai K, Ganase A, Loya Y. Revisiting the winners and the losers a decade after coral bleaching. *Mar Ecol Prog Ser*. 2011; 434: 67–76.
- [30] Green D, Edmunds P, Carpenter R. Increasing relative abundance of *Porites astreoides* on Caribbean reefs mediated by an overall decline in coral cover. *Marine Ecology Progress Series*. 2008; 359: 1–10. 10.3354/meps07454
- [31] Cramer KL, Jackson JB, Angioletti CV, Leonard-Pingel J, Guilderson TP. Anthropogenic mortality on coral reefs in Caribbean Panama predates coral disease and bleaching. *Ecology Letters*. 2012; 15(6): 561–7. 10.1111/j.1461-0248.2012.01768.x
- [32] Cramer K, editor Changes in coral communities and reef environments over the past few centuries in Caribbean Panama. Proceedings from the 2010 AGU Ocean Sciences Meeting; 2010: American Geophysical Union, 2000 Florida Ave., N. W. Washington DC 20009 USA.
- [33] Cramer KL, Leonard-Pingel JS, Rodríguez F, Jackson JB. Molluscan subfossil assemblages reveal the long-term deterioration of coral reef environments in Caribbean Panama. *Marine Pollution Bulletin*. 2015; 96(1): 176–87
- [34] Game ET, McDonald-Madden E, Puotinen ML, Possingham HP. Should we protect the strong or the weak? Risk, resilience, and the selection of marine protected areas. *Conservation Biology*. 2008; 22(6): 1619–29. 10.1111/j.1523-1739.2008.01037.x
- [35] Bell PRF, Elmetri I, Lapointe BE. Evidence of Large-Scale Chronic Eutrophication in the Great Barrier Reef: Quantification of Chlorophyll a Thresholds for Sustaining Coral Reef Communities. *AMBIO*. 2014; 43(3): 361–76. 10.1007/s13280-013-0443-1
- [36] Polónia ARM, Cleary DFR, de Voogd NJ, Renema W, Hoeksema BW, Martins A, et al. Habitat and water quality variables as predictors of community composition in an Indonesian coral reef: a multi-taxon study in the Spermonde Archipelago. *Science of The Total Environment*. 2015; 537: 139–51. 10.1016/j.scitotenv.2015.07.102
- [37] Gittleman JL, Kot M. Adaptation: statistics and a null model for estimating phylogenetic effects. *Systematic Biology*. 1990; 39(3): 227–41.
- [38] Oksanen J, Blanchet FG, Kindt R, Legendre P, Minchin PR, O'Hara R, et al. Package 'vegan'. *R Packag ver*. 2013; 254: 20–8.

- [39] Done TJ. Patterns in the distribution of coral communities across the central Great Barrier Reef. *Coral Reefs*. 1982; 1(2): 95–107.
- [40] Thompson D, Van Woesik R. Corals escape bleaching in regions that recently and historically experienced frequent thermal stress. *Proceedings of the Royal Society of London B: Biological Sciences*. 2009; 276(1669): 2893–901.
- [41] McClanahan TR, Ateweberhan M, Omukoto J. Long-term changes in coral colony size distributions on Kenyan reefs under different management regimes and across the 1998 bleaching event. *Marine Biology*. 2008; 153(5): 755–68.
- [42] Grottoli AG, Rodrigues LJ, Palardy JE. Heterotrophic plasticity and resilience in bleached corals. *Nature*. 2006; 440(7088): 1186–9. 10.1038/nature04565
- [43] Anthony K. Coral suspension feeding on fine particulate matter. *Journal of Experimental Marine Biology and Ecology*. 1999; 232: 85–106.
- [44] Aronson RB, Macintyre IG, Wapnick CM, O'Neill MW. Phase shifts, alternative states, and the unprecedented convergence of two reef systems. *Ecology*. 2004; 85(7): 1876–91
- [45] Hughes TP, Tanner JE. Recruitment failure, life histories, and long-term decline of Caribbean corals. *Ecology*. 2000; 81(8): 2250–63
- [46] Wild C, Hoegh-Guldberg O, Naumann MS, Colombo-Pallotta MF, Ateweberhan M, Fitt WK, et al. Climate change impedes scleractinian corals as primary reef ecosystem engineers. *Marine and Freshwater Research*. 2011; 62(2): 205–15. 10.1071/MF10254
- [47] Van Woesik R, Tomascik T, Blake S. Coral assemblages and physico-chemical characteristics of the Whitsunday Islands: evidence of recent community changes. *Marine and Freshwater Research*. 1999; 50(5): 427–40.
- [48] West K, Van Woesik R. Spatial and temporal variance of river discharge on Okinawa (Japan): inferring the temporal impact on adjacent coral reefs. *Marine Pollution Bulletin*. 2001; 42(10): 864–72.
- [49] Fabricius KE. Effects of terrestrial runoff on the ecology of corals and coral reefs: review and synthesis. *Marine Pollution Bulletin*. 2005; 50(2): 125–46
- [49] Marubini F, Atkinson MJ. Effects of lowered pH and elevated nitrate on coral calcification. *Marine Ecology Progress Series*. 1999; 188: 117–21.
- [50] Wooldridge S. A new conceptual model for the enhanced release of mucus in symbiotic reef corals during 'bleaching' conditions. *Marine Ecology Progress Series*. 2009; 396: 145–52. 10.3354/meps08310
- [51] Szmant AM. Nutrient enrichment on coral reefs: is it a major cause of coral reef decline? *Estuaries*. 2002; 25:743–66.
- [52] Wooldridge SA. Water quality and coral bleaching thresholds: Formalising the linkage for the inshore reefs of the Great Barrier Reef, Australia. *Marine Pollution Bulletin*. 2009; 58(5): 745–51. 10.1016/j.marpolbul.2008.12.013
- [53] Vega Thurber RL, Burkepille DE, Fuchs C, Shantz AA, McMinds R, Zaneveld JR. Chronic nutrient enrichment increases prevalence and severity of coral disease and bleaching. *Global Change Biology*. 2014;20(2):544–54. 10.1111/gcb.12450

- [54] Carilli JE, Prouty NG, Hughen KA, Norris RD. Century-scale records of land-based activities recorded in Mesoamerican coral cores. *Marine Pollution Bulletin*. 2009; 58(12): 1835–42. 10.1016/j.marpolbul.2009.07.024
- [55] Carilli JE, Norris RD, Black BA, Walsh SM, McField M. Local stressors reduce coral resilience to bleaching. *Plos ONE*. 2009; 4(7): e6324 10.1371/journal.pone.0006324
- [56] Hunte W, Wittenberg M. Effects of eutrophication and sedimentation on juvenile corals. *Marine Biology*. 1992; 114(4): 625–31. 10.1007/bf00357259
- [57] Carrillo L, Johns EM, Smith RH, Lamkin JT, Largier JL. Pathways and Hydrography in the Mesoamerican Barrier Reef System Part 1: Circulation. *Continental Shelf Research*. 2015; 109: 164–76. 10.1016/j.csr.2015.09.014.
- [58] Sheng J, Tang L. A numerical study of circulation in the western Caribbean Sea. *Journal of Physical Oceanography*. 2003; 33(10): 2049–69.
- [59] Sheng J, Tang L. A two-way nested-grid ocean-circulation model for the Meso-American Barrier Reef System. *Ocean Dynamics*. 2004; 54(2): 232–42.
- [60] Paris CB, Chérubin LM, Cowen RK. Surfing, spinning, or diving from reef to reef: effects on population connectivity. *Marine Ecology Progress Series*. 2007; 347: 285–300
- [61] Andrefouet S, Mumby PJ, Mcfield M, Hu C, Muller-Karger RE. Revisiting coral reef connectivity. *Coral Reefs*. 2002; 21: 43–8.
- [62] Prouty N, Hughen K, Carilli J. Geochemical signature of land-based activities in Caribbean coral surface samples. *Coral Reefs*. 2008; 27(4):727–42
- [63] Cooper TF, Uthicke S, Humphrey C, Fabricius KE. Gradients in water column nutrients, sediment parameters, irradiance and coral reef development in the Whitsunday Region, central Great Barrier Reef. *Estuarine, Coastal and Shelf Science*. 2007; 74(3): 458–70. 10.1016/j.ecss.2007.05.020
- [64] Jackson JB, Kirby MX, Berger WH, Bjorndal KA, Botsford LW, Bourque BJ, et al. Historical overfishing and the recent collapse of coastal ecosystems. *Science*. 2001; 293(5530): 629–37.
- [65] Perry C, Larcombe P. Marginal and non-reef-building coral environments. *Coral Reefs*. 2003; 22(4): 427–32.
- [66] Woesik R, Houk P, Isechal AL, Idechong JW, Victor S, Golbuu Y. Climate-change refugia in the sheltered bays of Palau: analogs of future reefs. *Ecology and Evolution*. 2012; 2(10): 2474–84. 10.1002/ece3.363
- [67] Graham NAJ, Cinner JE, Norström AV, Nyström M. Coral reefs as novel ecosystems: embracing new futures. *Current Opinion in Environmental Sustainability*. 2014; 7: 9–14. 10.1016/j.cosust.2013.11.023