

## IMPACT OF BULK AND NANO-SIZED $Al_2O_3$ ON *Cynara scolymus* SEED GERMINATION AND SEEDLING GROWTH

Zakie Angizeh, Homa Mahmoodzadeh✉, Ali Es-haghi, Ali Akbar Ziroohi

Department of Biology, Mashhad Branch, Islamic Azad University, Mashhad, Iran

### ABSTRACT

**Background.** Nanoparticles have been applied worldwide, posing substantial effects on the environment and its living organisms. Plants as sessile organisms are always exposed to considerable fluctuations of nanoparticles concentrations. Here, the effects of different concentrations of bulk and nanosized  $Al_2O_3$  on seed germination and seedling growth of *Cynara scolymus* were investigated in a randomized completely design with four replications.

**Material and methods.** The experimental treatments included four concentrations of bulk  $Al_2O_3$  (10, 50, 100 and 500 ppm), four concentrations of nanosized  $Al_2O_3$  (10, 50, 100 and 500 ppm), and control without any  $Al_2O_3$ .

**Results.** The results indicated that among the *Cynara scolymus* germination indices, only mean germination time and weighted germination index was not affected by treatments. In addition, plumule length, radicle length, seedling fresh and dry weight were affected by bulk and nanosized  $Al_2O_3$  concentrations, significantly. Radicle length at 50 ppm concentration of nanosized  $Al_2O_3$  was higher than those of the untreated control and bulk  $Al_2O_3$  at 10, 100 and 500 ppm concentrations. It is concluded that treatment with bulk  $Al_2O_3$  treatments have more inhibitory effects on germination indices of *Cynara scolymus* in comparison to nanosized  $Al_2O_3$ .

**Conclusion.** Overall, we found through our study that bulk and nano sized  $Al_2O_3$  in high concentrations have a negative impact on the seed germination and seedling growth of *Cynara scolymus*. We saw an overall significant reduction in plumule length, seedling dry biomass, and vigor index of *Cynara scolymus* seedlings after being exposed to different concentrations of bulk and nano sized  $Al_2O_3$ , but no significant change in the mean germination time, weighted germination index and radicle length of the seedlings. The two common possibilities are either the bulk and nano sized  $Al_2O_3$  will adhere to the roots impeding any uptake of water and nutrients, or that the nanoparticles are up taken and translocated within the plant therefore causing toxicity internally.

**Key words:** *Cynara scolymus*,  $Al_2O_3$  nanoparticles, bulk  $Al_2O_3$ , germination indices, seedling biomass, seedling length

### INTRODUCTION

Nanotechnology is a fast-developing industry, posing substantial impacts on economy, society and environment. Particles in such a size (<100 nm) fall in the transitional zone between individual atoms or

molecules and the corresponding bulk material, which can modify the physicochemical properties of the material (e.g., performing exceptional feats of conductivity, reactivity, and optical sensitivity). Therefore, such materials can generate adverse biological effects in living cells (Nel *et al.*, 2006).

✉ [homa\\_mahmoodzadeh@yahoo.com](mailto:homa_mahmoodzadeh@yahoo.com)

There is an increasing amount of research on the toxicology of nanomaterials. Some researchers have shown the toxicity of nanoparticles, such as fullerene, carbon nanotubes and metal oxides to human cells, bacteria, and rodents (Hussain *et al.*, 2005; Jia *et al.*, 2005; Brunner *et al.*, 2006; Lam *et al.*, 2006; Soto *et al.*, 2006). Nano-aluminum is being used in increasing quantities as energetic material (Navrotsky, 2003; Argonide, 2004). Aluminum nanoparticles are the basis for superior fuels for space launch vehicles and other rockets or missiles. A single Space Shuttle launch requires approximately 226 800 kg of Al, all in the form of particles with sizes greater than 10 nm. Replacement by the faster-burning nano-aluminum particles in the Space Shuttle's solid rocket boosters will allow motors to be packed more efficiently which may reduce motor size and thereby increase mission payload. Virtually all food, water, and air contain some aluminum which nature is well adapted to handle. It is reported that Al is poorly absorbed by the body and efficiently eliminated; however, when absorption occurs, Al is distributed mainly in bone, liver, testicles, kidneys, and brain. Aluminum has been linked to Alzheimer's disease. The respiratory system appears to be the primary target following inhalation exposure to Al (Sharma and Mishra, 2005). Toxicity of dissolved aluminum to aquatic organisms has been reported (Ward *et al.*, 2006). Aluminum cation (Al<sup>3+</sup>) is very unfriendly to agriculture as it injures plant root cells and thus interferes with root growth and nutrient uptake in crops (Quing *et al.*, 2006). Most aluminum-containing compounds have low solubility in water unless the water is acidic. After investigating the phytotoxicity of nano-scale alumina (nano-Al<sub>2</sub>O<sub>3</sub>) powders with or without phenanthrene coating, Yang and Watts concluded that uncoated alumina particles inhibited root elongation of corn, cucumber, soybean, cabbage and carrot (Yang and Watts, 2005). This study attracted attentions from scientists and media, and was used to claim that nanoparticles can exert a negative effect on plants (Murashov, 2006). But the authors did not identify dissolution of nano-Al<sub>2</sub>O<sub>3</sub> in solution, thus, failed to clarify, if the phytotoxicity was from nano-Al<sub>2</sub>O<sub>3</sub> or aluminum ion in the aqueous solution. Therefore, this study aimed to provide new information about phytotoxicology of nanoparticles by investigating the

effect of bulk and nano-Al<sub>2</sub>O<sub>3</sub> on seed germination and seedling growth of medicinal plant, *Cynara scolymus*.

## MATERIAL AND METHODS

### Description of Materials

*Cynara scolymus* (var. Pishtaz) seeds were taken from the Pakan Bazr Company, Isfahan Province, Iran. Nanosized Al<sub>2</sub>O<sub>3</sub> powder was supplied by Nutrient Company. The size and topography of Al<sub>2</sub>O<sub>3</sub> nanoparticles (Figs. 1 and 2) were determined by scanning tunneling microscope (STM) in the Central Laboratory of Ferdowsi University of Mashhad, Iran. X-ray diffraction (XRD) pattern of Al<sub>2</sub>O<sub>3</sub> nanoparticles was shown in Fig. 3. XRD measurement showed that the used Al<sub>2</sub>O<sub>3</sub> nanoparticles were made by Al<sub>2</sub>O<sub>3</sub> and Corundum. Bulk Al<sub>2</sub>O<sub>3</sub> was supplied by merk Company.

### Experimental Design and Data Observation

In order to study the effect of different concentrations of bulk and nanosized Al<sub>2</sub>O<sub>3</sub> on *Cynara scolymus* germination, a randomized completely design with four replications was employed. The experimental treatments included four concentrations (10, 50, 100 and 500 ppm) of bulk and four concentrations (10, 50, 100 and 500 ppm) of nanosized Al<sub>2</sub>O<sub>3</sub> and untreated control (without any Al<sub>2</sub>O<sub>3</sub> types). The experiment was conducted in laboratory conditions with natural light and an average temperature of 25±1°C at the Faculty of Science, Mashhad Branch, Islamic Azad University, Mashhad, Iran, in 2014.

One hundred seeds of similar size were randomly selected and placed on moistened paper as four groups of seeds in Petri dishes, and then 10 ml of each 16-concentration treatment was added to each Petri dish. For the control, only distilled water was added to Petri dishes. Germination tests were performed according to the rule issued by the International Seed Testing Association.

All concentrations of Al<sub>2</sub>O<sub>3</sub> and the control were run at the same time and consequently under equal light and temperature conditions. The number of germinated seeds was noted daily for 7 days. Seeds were considered as germinated when their radicle showed at least 1 mm length. In this study, we used following germination parameters: Germination percentage (GP, %), Relative germination percentage

(RGP), Mean germination time (MGT), Germination index (GI) and Weighted germination index (WGI). Final percentage germination (GP) for each treatment was calculated after seven days. The germination index (GI) is based on number of seeds that germinated and the germination rate. These parameters were also calculated from the formulas proposed by (Figuroa and Armesto, 2001; Bu *et al.*, 2006; Wu and Du, 2007):

$$GP = 100 \times GN / SN \quad (1)$$

GN is the total number of germinated seed, SN is the total number of seeds tested:

$$RGP = GP \text{ treatment} / GP \text{ control} \times 100 \quad (2)$$

$$GI = (\sum(N-i) \times Gi \times 100) / (N \times GN) \quad (3)$$

GI is a synthetic measure designed to reflect the synthetical germination ability including germination rate and germination numbers. Where *i* is the number of days since the day of sowing and *G<sub>i</sub>* is the number of seeds germinated on day *i*.

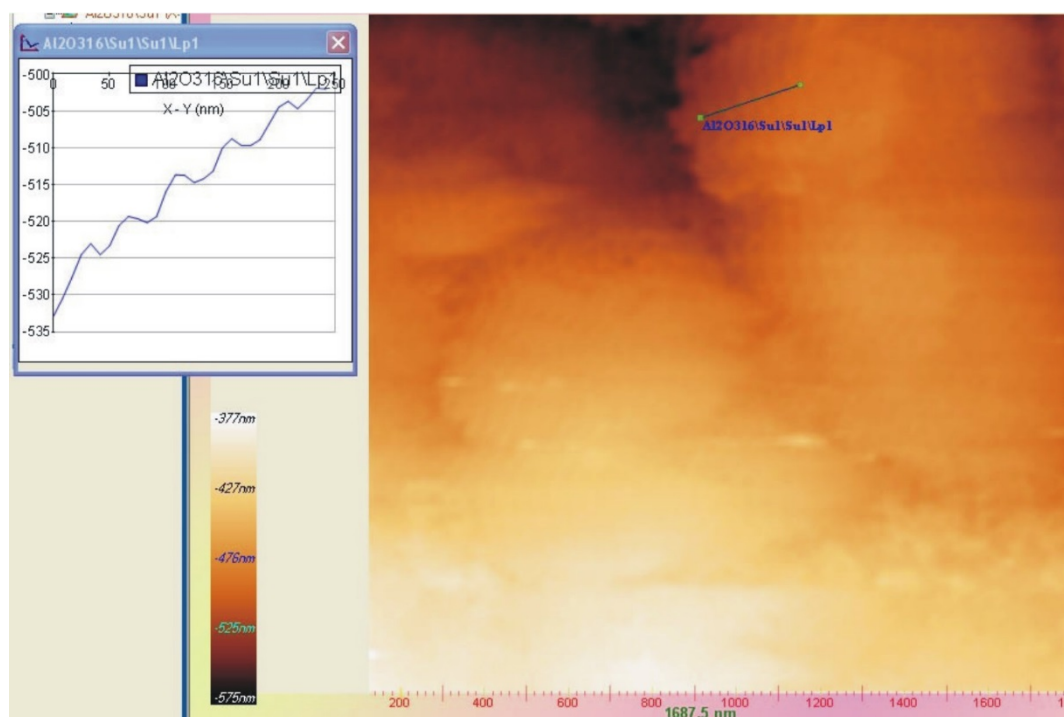
A weighted germination index (WGI) as described by Bu *et al* was calculated with maximum weight given to the seeds germinating early and less to those germinating late (Bu *et al.*, 2007):

$$WGI = [N \times n_1 + (N-1) \times n_2 + (N-2) \times n_3 + \dots] / N \times N' \quad (4)$$

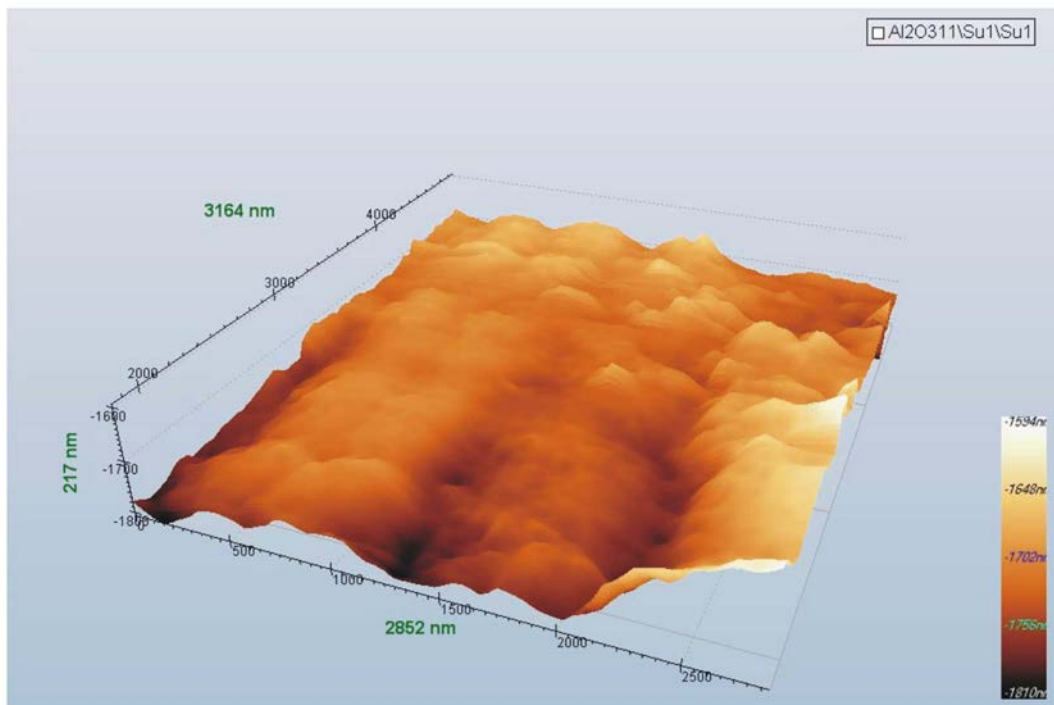
where *n*<sub>1</sub>, *n*<sub>2</sub>, ..., *n*<sub>60</sub> are the number of seeds that germinated on first, second, and subsequent days until the 60th day, respectively; *N* is total days of experiment; *N'* is the total number of seeds placed in incubation:

$$\text{Vigor index} = \text{germination\%} \times \text{seedling length (root + shoot)} \quad (5)$$

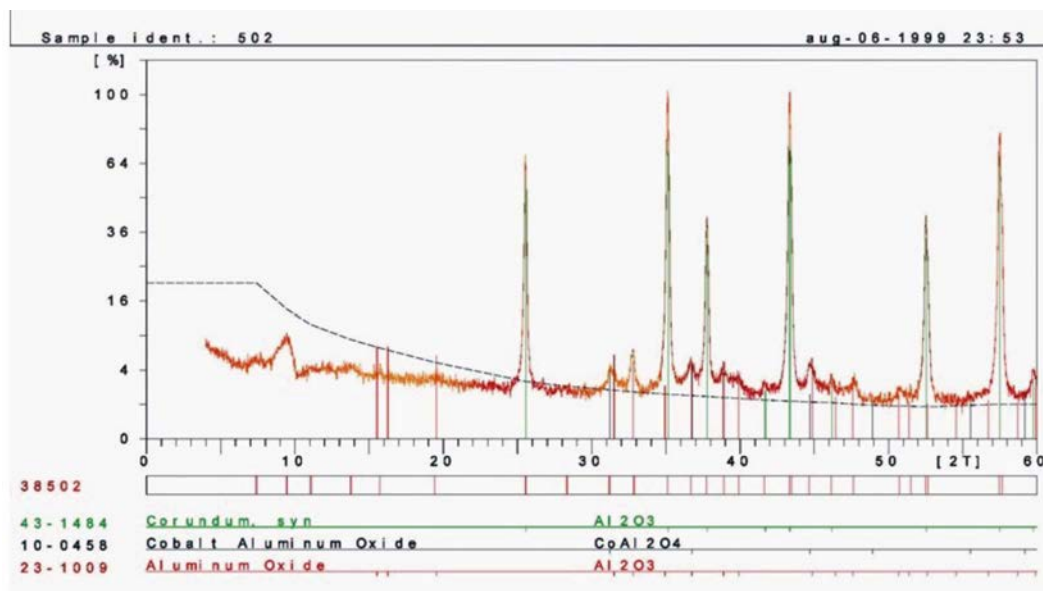
After an incubation period of 7 days, plumule and radical length of seedlings were measured using a ruler. In order for dry biomass to be weighed, the 7-day seedlings were first weighed; then, having been placed in oven at 80°C for 48 h, they were weighed for a second time.



**Fig. 1.** Image of nanosized Al<sub>2</sub>O<sub>3</sub> by STM



**Fig. 2.** Topographic image of nanosized  $\text{Al}_2\text{O}_3$  by STM



**Fig. 2. 3** XRD pattern of nano  $\text{Al}_2\text{O}_3$  particles

### TTC viability tests for root tips

2, 3, 5-triphenyltetrazolium chloride (TTC) was used as a histopathologic stain for testing the viability of root tips. The test was as follows: 5 mL of 0.5% solution of TTC was added to test tubes containing root tips, the temperature was kept at  $35 \pm 1^\circ\text{C}$ . After 5 h in the dark, the TTC solution was removed with a syringe and root tips were thoroughly rinsed with distilled water and then examined. The red colored root tips were considered to be viable and others were non-viable or dead (Shaymurat *et al.*, 2012).

### Statistical analysis

Significant differences for all statistical tests were evaluated at the level of  $P \leq 0.05$  with ANOVA. All data analyses were conducted using SPSS for Windows, Version 13.0.

## RESULTS

Once the *Cynara scolymus* seeds were plated, it took approximately three to five days for them to germinate. After seven days, the germination percentage of the *Cynara scolymus* seeds were calculated for each concentration of bulk and nano aluminum oxide. For the seeds grown on control media without any aluminum oxide, the germination percentage was 97.5%, therefore almost all of the *Cynara scolymus* seeds germinated. The lowest germination percentage (80%) was found in 500 ppm concentration nanosized Al<sub>2</sub>O<sub>3</sub> (Table 1). Although the highest germination rate (99.4%) was shown in the control treatment, but had no significant difference with other treatments, except 50 and 500 ppm bulk Al<sub>2</sub>O<sub>3</sub>. The lowest mean germination time (5.08 day) was found in 50 ppm concentration nanosized Al<sub>2</sub>O<sub>3</sub>, and the highest (5.24 day) was shown in the control treatment. Therefore 50 ppm concentration nanosized Al<sub>2</sub>O<sub>3</sub> treatment reduced mean germination time by 3% in comparison to untreated control. Although there was some variation between the mean germination times of seeds grown in media with differing concentrations of bulk and nanoparticles aluminum oxide, this variation was not statistically significant. In the media containing 10 and 500 ppm bulk Al<sub>2</sub>O<sub>3</sub>, the relative germination percentage (105.9 and 106.2,

respectively) were higher than other treatments and had significant difference with 50 and 500 ppm nanosized Al<sub>2</sub>O<sub>3</sub> treatments (Table 1). The highest germination index (29.2) was found in the control treatment and the lowest (25.2) was shown in 500 ppm bulk Al<sub>2</sub>O<sub>3</sub> treatment. Therefore 500 ppm concentration bulk Al<sub>2</sub>O<sub>3</sub> treatment reduced germination index by 14% in comparison to untreated control, while other treatments did not significantly reduce germination index in comparison with control (Table 1). Different concentrations of bulk and nano Al<sub>2</sub>O<sub>3</sub> did not significantly affect the weighted germination index of *Cynara scolymus* seeds. The effect of studied treatments on root length was not significant, but they had a significant effect on plumule length. plumule length at all of treatments of nano and bulk Al<sub>2</sub>O<sub>3</sub> was lower than control. The lowest plumule length was achieved at 500 ppm nanosized Al<sub>2</sub>O<sub>3</sub> (Table 1). All nano sized Al<sub>2</sub>O<sub>3</sub> treatments decreased seedling fresh biomass significantly. The lowest seedling fresh biomass was found in 50 and 500 ppm nano. Al<sub>2</sub>O<sub>3</sub> concentrations seedling fresh biomass significantly. Bulk Al<sub>2</sub>O<sub>3</sub> treatments have no significant effect on seedling fresh biomass. Experimental treatments affected seedling dry biomass significantly. The lowest seedling dry biomass (0.011 g) was found in 500 ppm concentration nanosized Al<sub>2</sub>O<sub>3</sub>, and the highest was shown in the control treatment (0.029 g). Therefore 500 ppm concentration nanosized Al<sub>2</sub>O<sub>3</sub> treatment reduced seedling dry biomass by 62% in comparison to untreated control, while 500 ppm concentration bulk Al<sub>2</sub>O<sub>3</sub> decreased seedling dry biomass by 34% in comparison with control (Table 2). Vigor index was affected significantly by bulk and nanosized Al<sub>2</sub>O<sub>3</sub> concentrations except 10 and 50 ppm bulk and 50 ppm nano treatments (Table 2). The lowest vigor index was shown in 500 ppm nano Al<sub>2</sub>O<sub>3</sub> that reduced vigor index by 84% in comparison with the control (Table 2). The TTC tests showed that the effects of bulk and nano Al<sub>2</sub>O<sub>3</sub> on root tips not varied with concentrations applied (Fig 4). For 24-h treatment all root tips were colored red.

**Table 1.** Effect of different concentrations of bulk and nanosized Al<sub>2</sub>O<sub>3</sub> on seed germination of *Cynara scolymus*

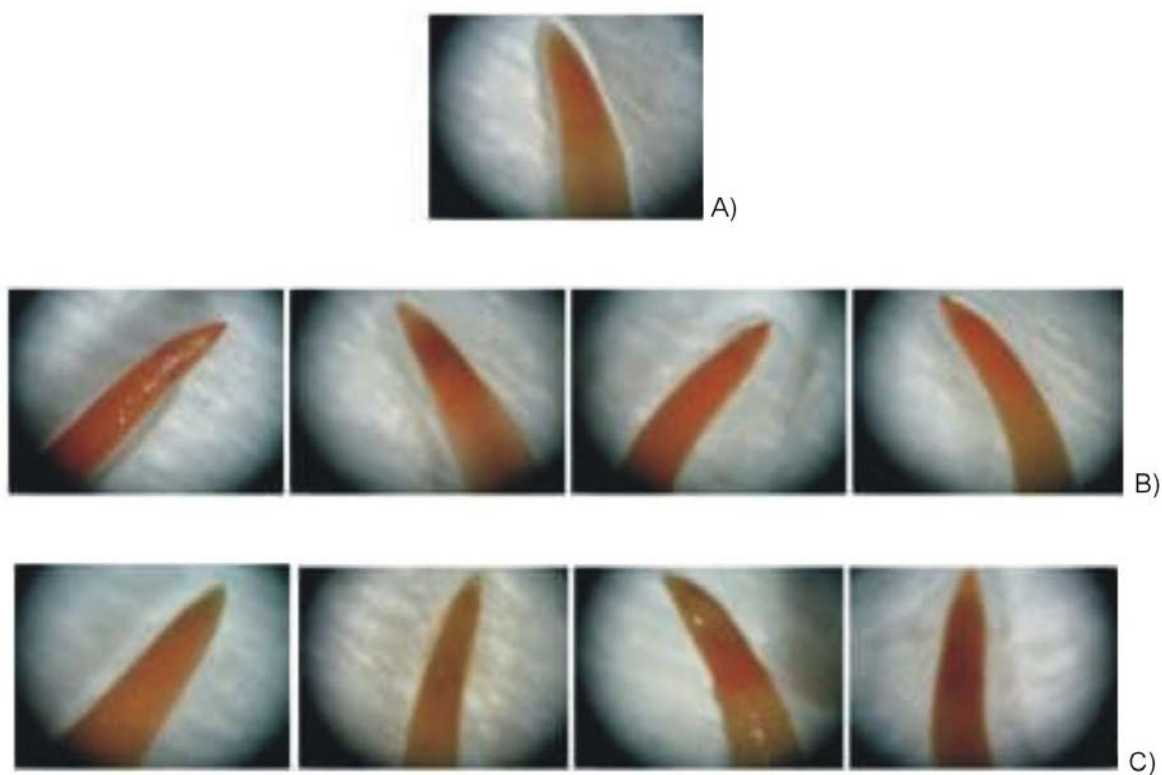
Concentration ppm	Germination %	RGP	Germination Rate %Day <sup>-1</sup>	MGT Day	GI	WGI
Bulk Al <sub>2</sub> O <sub>3</sub>						
10	90 abc	105.9 a	86.2 ab	5.17 a	26.1 ab	0.358 a
50	85 bc	100 ab	82.8 b	5.18 a	27.6 ab	0.361 a
100	90 abc	106.2 a	89.7 ab	5.20 a	25.6 ab	0.428 a
500	82.5 c	97.5 ab	83.9 b	5.22 a	25.2 b	0.392 a
Nano Al <sub>2</sub> O <sub>3</sub>						
10	95 ab	97.7 ab	96.8 ab	5.20 a	27 ab	0.412 a
50	87.5 abc	89.7 bc	93.8 ab	5.08 a	27.3 ab	0.415 a
100	90 abc	92.5 abc	90.1 ab	5.16 a	26.1 ab	0.404 a
500	80 c	82.2 c	85.22 ab	5.17 a	25.8 ab	0.400 a
Control	97.5 a	----	99.4 a	5.24 a	29.2 a	0.418 a

Means in each column followed by similar letters are not significantly different at the 5% probability level using Duncan's multiple range test

**Table 2.** Effect of bulk and nanosized Al<sub>2</sub>O<sub>3</sub> concentrations on seedling growth of *Cynara scolymus*

Concentration ppm	Plumule Length cm	Radicle Length cm	Seedling Fresh Biomass, g	Seedling Dry Biomass, g	Vigor Index
Bulk Al <sub>2</sub> O <sub>3</sub>					
10	0.47 cd	3.45 abc	0.155 ab	0.012 cd	457.3 a
50	0.70 ab	4.45 a	0.155 ab	0.015 cd	488.0 a
100	0.54 bcd	3.85 ab	0.156 ab	0.021 b	301.3 bc
500	0.52 bcd	3.02 bc	0.161 ab	0.017 bc	367.3 b
Nano Al <sub>2</sub> O <sub>3</sub>					
10	0.67 abc	2.52 c	0.148 b	0.012 cd	317.6 bc
50	0.47 cd	3.42 abc	0.142 b	0.015 cd	461.3 a
100	0.47 cd	2.43 c	0.147 b	0.012 cd	234.6 c
500	0.41 d	2.41 c	0.143 b	0.011 d	80.6 d
Control	0.82 a	3.13 bc	0.173 a	0.029 a	516.6 a

Means in each column followed by similar letters are not significantly different at the 5% probability level using Duncan's multiple range test



**Fig. 4.** TTC tests for different concentrations of bulk and nano-sized  $\text{Al}_2\text{O}_3$ , A (control), B (Bulk  $\text{Al}_2\text{O}_3$ (10, 50, 100 and 500 ppm)), C(Nano-sized  $\text{Al}_2\text{O}_3$ (10, 50, 100 and 500 ppm))

## DISCUSSION

Nanotoxicology, an emerging discipline, is receiving increasing attraction. Nanotoxicity has been the research focus of many publications, including several reviews (Oberdorster *et al.*, 2005; Nel *et al.*, 2006; Wiesner *et al.*, 2006). Some nanoparticles, such as fullerene, SWCNT and  $\text{TiO}_2$ , have been widely used as test materials to reveal their nanotoxicity mechanisms. However, available information on the is topic too scarce to search anytopic is too scarce to reach any consensus on nanotoxicity and its mechanism, particularly for the nanoparticle used in our study. Phytotoxicity in higher plants should be investigated in order to develop a comprehensive toxicity profile for nanoparticles. Seed germination and root elongation is a rapid and widely used acute phytotoxicity test with several advantages: sensitivity, simplicity, low cost and suitability for unstable

chemicals or samples (Munzuroglu and Geckil, 2002). Germination is normally known as a physiological process beginning with water imbibition by seeds and culminating in the emergence of the rootlet (Kordon, 1992). However, there are different definitions of seed germination according to its root length emergence of root, >1mm or >5mm (Kordon, 1992; Ren *et al.*, 1996; Munzuroglu and Geckil, 2002, Murata *et al.*, 2003). In this study, seeds showing emergence of radicle coming out of the seed coat were recorded as being germinated. Seed coat plays a very important role in protecting the embryo from harmful external factors. Seed coats can have selective permeability (Wierzbicka and Obidzinska, 1998). Pollutants, though having obviously inhibitory effect on root growth, may not affect germination if they can not pass through seed coats. This may explain that seed germination in this study was not greatly altered by nanoparticles. These findings agree with recent reports

stating that seed germination for different plant species was not affected by Al<sub>2</sub>O<sub>3</sub> (Yang and Watts, 2005; Lin and Xing, 2007; Lee *et al.*, 2010). It has also been reported that the process of seed germination in wheat is not affected by the presence of aluminum ion under experimental settings (Jamal *et al.*, 2006).

Our results also were consistent with a study which investigated the effects of five different types of nanomaterials (multi-walled carbon nanotubes, aluminum oxide, zinc oxide, aluminum, and zinc) on the seed germination rate and root length of five agriculturally important crops (radish, rape, ryegrass, lettuce, corn, and cucumber). They found that out of all five crops, that only the seed germination rate of ryegrass and corn were affected by nano-Zn and nano-ZnO, respectively. In short, Lin and Xing found that in response to 200 g·dm<sup>-3</sup> Al<sub>2</sub>O<sub>3</sub> nanoparticles that none of the germination rates from all 5 plants they tested were significantly affected (Lin and Xing, 2007). These results mirror our results because 2000 g·dm<sup>-3</sup> converts to the concentration of 0.2 g of Al<sub>2</sub>O<sub>3</sub> nanoparticles per 100 mL of media. This concentration fits approximately in the middle of the range of Al<sub>2</sub>O<sub>3</sub> nanoparticle concentrations that we used (0.1 g, 0.5 g, and 1 g of Al<sub>2</sub>O<sub>3</sub> nanoparticles per 100 mL of media). Lin and Xing state that the seed coat of the tobacco seeds was most likely not permeable to the aluminum oxide nanoparticles, therefore the germination rate was not affected greatly (Lin and Xing, 2007). Thus, it would not be until after the seedlings started to emerge from the seed coat, that they would be affected by aluminum oxide nanoparticles. As shown in Table 2, radicle growth was promoted at 10, 50 and 100 ppm of bulk Al<sub>2</sub>O<sub>3</sub>. The positive effect of Al<sub>2</sub>O<sub>3</sub> on root elongation in *Arabidopsis thaliana* has been reported by Lee *et al.* (2010). Improvement in wheat root growth has also been reported at low doses of Al (Aniol, 1984). On the other hand, the radicle length in presence of 10, 100 and 500 ppm nano Al<sub>2</sub>O<sub>3</sub> was reduced slightly (not significantly) as compared to control radicle length. This observation can be attributed to high Al content in the roots. The decrease in root elongation in the presence of nAl<sub>2</sub>O<sub>3</sub> for various plant species has been reported (Yang and Watts, 2005; Lin and Xing, 2007). In contrast, plumule length was affected by bulk and nano Al<sub>2</sub>O<sub>3</sub> (Table 2) which may be a consequence of high transportability

of this material to the shoot. Therefore, it may be concluded that the more uptake of these particles, the more adverse effects on the growth of seedlings.

The TTC tests showed that after 24-h treatment, all root tips were colored red that confirmed bulk and nano Al<sub>2</sub>O<sub>3</sub> could not cause the root tip cells to death. It was also found that the nano Al<sub>2</sub>O<sub>3</sub> roots were less red than bulk Al<sub>2</sub>O<sub>3</sub> treatments (Fig 4). These results showed that nano Al<sub>2</sub>O<sub>3</sub> treatments had more toxicity effects than bulk Al<sub>2</sub>O<sub>3</sub> treatments.

## CONCLUSIONS

Overall, we found through our study that bulk and nano sized Al<sub>2</sub>O<sub>3</sub> in high concentrations have a negative impact on the seed germination and seedling growth of *Cynara scolymus*. We saw an overall significant reduction in plumule length, seedling dry biomass, and vigor index of *Cynara scolymus* seedlings after being exposed to different concentrations of bulk and nano sized Al<sub>2</sub>O<sub>3</sub>, but no significant change in the mean germination time, weighted germination index and radicle length of the seedlings. The two common possibilities are either the bulk and nano sized Al<sub>2</sub>O<sub>3</sub> will adhere to the roots impeding any uptake of water and nutrients, or that the nanoparticles are up taken and translocated within the plant therefore causing toxicity internally. Aluminum oxide exists commonly in the soil as aluminum is one of the most abundant elements found in the earth's crust. It is not known how much of the aluminum oxide found in the soil exists as nanoparticles, therefore more research needs to be conducted to determine if the increasing use of aluminum oxide nanoparticles in industry is drastically affecting the concentrations of aluminum oxide that is already present in the soil. More research also needs to be conducted in order to elucidate the mechanisms of aluminum oxide nanoparticle toxicity and to determine the exact mode of phytotoxicity in plants.

## REFERENCES

- Aniol, A. (1984). Induction of aluminum tolerance in wheat seedlings by low doses of aluminum in the nutrient solution. Plant Physiol., 76, 551–555.



- Argonide. (2004). Nano aluminum powders for advanced rocket propellants.  
<http://www.argonide.com/propellants.html>.
- Brunner, T.J., Wick, P., Manser, P., Spohn, P., Grass, R.N., Limbach, L.K., Bruinink, A., Stark, W.J. (2006). In vitro cytotoxicity of oxide nanoparticles: comparison to asbestos, silica, and the effect of particle solubility. Environ. Sci. Technol., 40, 4374–4381.
- Bu, H., Chen, X.-L., Wang, Y., Xu, X., Liu, K., Du, G. (2007). Germination time, other plant traits and phylogeny in an alpine meadow on the eastern Qinghai-Tibet Plateau. Community Ecol., 8, 221–227.
- Figueroa, J., Armesto, J. (2001.) Community-wide germination strategies in a temperate rainforest of Southern Chile: ecological and evolutionary correlates. Aust. J. Bot., 49, 411–425.
- Hussain, S., Hess, K., Gearhart, J., Geiss, K., Schlager, J. (2005). In vitro toxicity of nanoparticles in BRL 3A rat liver cells. Toxicology in vitro, 19, 975–983.
- Jamal, S., Iqbal, M., Athar, M. (2006). Phytotoxic effect of aluminum and chromium on the germination and early growth of wheat (*Triticum aestivum*) varieties Anmol and Kiran. Int. J. Environ. Sci. Technol., 3, 411–416.
- Jia, G., Wang, H., Yan, L., Wang, X., Pei, R., Yan, T., Zhao, Y., Guo, X. (2005). Cytotoxicity of carbon nanomaterials: single-wall nanotube, multi-wall nanotube, and fullerene. Environ. Sci. Technol., 39, 1378–1383.
- Kordon, H. (1992). Seed viability and germination: a multi-purpose experimental system. J. Biol. Edu., 26, 247–251.
- Lam, C.-W., James, J.T., Mccluskey, R., Arepalli, S., Hunter, R.L. (2006). A review of carbon nanotube toxicity and assessment of potential occupational and environmental health risks. Crit. Rev. Toxicol., 36, 189–217.
- Lee, C.W., Mahendra, S., Zodrow, K., Li, D., Tsai, Y.C., Braam, J., Alvarez, P.J. (2010). Developmental phytotoxicity of metal oxide nanoparticles to *Arabidopsis thaliana*. Environ. Toxicol. Chem., 29, 669–675.
- Lin, D., Xing, B. (2007). Phytotoxicity of nanoparticles: inhibition of seed germination and root growth. Environ. Pollut., 150, 243–250.
- Munzuroglu, O., Geckil, H. (2002). Effects of metals on seed germination, root elongation, and coleoptile and hypocotyl growth in *Triticum aestivum* and *Cucumis sativus*. Arch. Environ. Con. Tox., 43, 203–213.
- Murashov, V. (2006). Comments on "Particle surface characteristics may play an important role in phytotoxicity of alumina nanoparticles" by Yang, L., Watts, D.J. Toxicology Letters, 2005, 158, 122–132. Toxicol. Lett., 164, 185–187; author reply 1886.
- Murata, M., Hammes, P., Zharare, G. (2003). Effect of solution pH and calcium concentration on germination and early growth of groundnut. J. Plant Nut., 26, 1247–1262.
- Navrotsky, A. (2003). Energetics of nanoparticle oxides: interplay between surface energy and polymorphism. Geochem. Trans., 4, 34–37.
- Nel, A., Xia, T., Mädler, L., Li, N. (2006). Toxic potential of materials at the nano level. Science, 311, 622–627.
- Oberdörster, G., Oberdörster, E., Oberdörster, J. (2005). Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. Environ Health Perspect 113, 823–839.
- Ren, L., Zeiler, L.F., Dixon, D.G., Greenberg, B.M. (1996). Photoinduced Effects of Polycyclic Aromatic Hydrocarbons on *Brassica napus* (Canola) during Germination and Early Seedling Development. Ecotoxicol. Environ. Safety, 33, 73–80.
- Sharma, P., Mishra, K.P. (2006). Aluminum-induced maternal and developmental toxicity and oxidative stress in rat brain: response to combined administration of Tiron and glutathione. Reprod. Toxicol., 21, 313–321.
- Shaymurat, T., Gu, J., Xu, C., Yang, Z., Zhao, Q., Liu, Y., Liu, Y. (2012). Phytotoxic and genotoxic effects of ZnO nanoparticles on garlic (*Allium sativum* L.): A morphological study. Nanotoxicol., 6, 241–248.
- Soto, K., Carrasco, A., Powell, T., Murr, L., Garza, K. (2006). Biological effects of nanoparticulate materials. Mater. Sci. Eng., C, 26, 1421–1427.
- Ward, R.J., Mccrohan, C.R. & White, K.N. (2006). Influence of aqueous aluminium on the immune system of the freshwater crayfish *Pacifastacus leniusculus*. Aquatic Toxicol., 77, 222–228.
- Wierzbicka, M., Obidzińska, J. (1998). The effect of lead on seed imbibition and germination in different plant species. Plant Sci., 137, 155–171.
- Wiesner, M.R., Lowry, G.V., Alvarez, P., Dionysiou, D., Biswas, P. (2006). Assessing the risks of manufactured nanomaterials. ACS Publications.
- Wu, G., Du, G. (2007). Germination is related to seed mass in grasses (Poaceae) of the eastern Qinghai-Tibetan Plateau, China. Nord J. Bot., 25, 361–365.
- Yang, L., Watts, D.J. (2005). Particle surface characteristics may play an important role in phytotoxicity of alumina nanoparticles. Toxicol. Lett., 158, 122–132.

## WPŁYW MASY I NANOCZĄSTEK Al<sub>2</sub>O<sub>3</sub> NA KIEŁKOWANIE I WZROST SIEWEK *Cynara scolymus*

### Streszczenie

Nanocząstki są stosowane na całym świecie, wywierając znaczny wpływ na środowisko i żywe organizmy. Rośliny jako organizmy ukorzenione są zawsze narażone na znaczne wahania stężeń nanocząstek. Zbadano wpływ różnych stężeń Al<sub>2</sub>O<sub>3</sub> w masie i nanocząsteczkach na kiełkowanie nasion i wzrost siewek *Cynara scolymus* w całkowicie losowym projekcie z czterema powtórzeniami. Zabiegi doświadczalne obejmowały cztery stężenia Al<sub>2</sub>O<sub>3</sub> w masie (10, 50, 100 i 500 ppm), cztery stężenia Al<sub>2</sub>O<sub>3</sub> w postaci nanocząstek (10, 50, 100 i 500 ppm) oraz kontrolę (bez Al<sub>2</sub>O<sub>3</sub>). Wyniki wskazują, że spośród wskaźników kiełkowania *Cynara scolymus* zabiegi nie miały wpływu na średni czas kiełkowania i ważony wskaźnik kiełkowania. Na długość pędu embrionalnego, długość korzonków, świeżą i suchą masę siewek istotnie wpływały stężenia Al<sub>2</sub>O<sub>3</sub> w masie i nanoskali. Długość rodnika przy stężeniu 50 ppm Al<sub>2</sub>O<sub>3</sub> w postaci nanocząstek była wyższa w porównaniu z kontrolą i masowym Al<sub>2</sub>O<sub>3</sub> przy stężeniach 10, 100 i 500 ppm. Stwierdzono, że traktowanie dużą ilością Al<sub>2</sub>O<sub>3</sub> ma bardziej hamujący wpływ na wskaźniki kiełkowania *Cynara scolymus* niż stosowanie Al<sub>2</sub>O<sub>3</sub> w postaci nanocząstek. W badaniach określono, że Al<sub>2</sub>O<sub>3</sub> w dużych ilościach i w wysokich stężeniach nanocząsteczek ma negatywny wpływ na kiełkowanie nasion i wzrost siewek *Cynara scolymus*. Zaobserwowano znaczne zmniejszenie długości pędu embrionalnego, suchej masy sadzonek i wskaźnika wigoru siewek *Cynara scolymus* po ekspozycji na różne stężenia Al<sub>2</sub>O<sub>3</sub> w rozmiarze masowym i nano, ale bez znaczących zmian w średnim czasie kiełkowania, ważonym wskaźniku kiełkowania i długości korzonków siewek. Wynika to albo z przywierania Al<sub>2</sub>O<sub>3</sub> do korzeni w rozmiarze masowym i nanometrowym, utrudniając wchłanianie wody i składników odżywczych, albo nanocząsteczki są pobierane i przemieszczane w obrębie rośliny, powodując w ten sposób toksyczność wewnętrzną.

**Słowa kluczowe:** biomasa siewek, *Cynara scolymus*, długość siewek, masa Al<sub>2</sub>O<sub>3</sub>, nanocząstki, wskaźniki kiełkowania