

EXPERIMENTAL PAPERS

Comparison of proanthocyanidins content in *Rhodiola kirilowii* and *Rhodiola rosea* roots – application of UPLC-MS/MS method

AGNIESZKA GRYSZCZYŃSKA^{1*}, ANNA KRAJEWSKA-PATAN², WALDEMAR BUCHWALD³,
BOGUSŁAW CZERNY^{4,5}, SEBASTIAN MIELCAREK¹, KAROLINA RUDZIŃSKA⁶,
PRZEMYSŁAW M. MROZIKIEWICZ^{1,7}

¹Department of Quality Control of Medicinal Products and Dietary Supplements
Institute of Natural Fibres and Medicinal Plants
Libelta 27
61-707 Poznan, Poland

²Department of Pharmacology and Experimental Biology
Institute of Natural Fibres and Medicinal Plants
Libelta 27
61-707 Poznan, Poland

³ Department of Botany, Breeding and Agricultural Technology
Institute of Natural Fibres and Medicinal Plants
Kolejowa 2
62-064 Plewiska/Poznań, Poland

⁴ Institute of Natural Fibres and Medicinal Plants
Libelta 27
61-707 Poznan, Poland

⁵Department of General Pharmacology and Pharmacoconomics
Pomeranian Medical University
Żołnierska 48
70-204 Szczecin, Poland

⁶109 Military Hospital Outpatient SPZOZ
Piotra Skargi 9-11
70-965 Szczecin, Poland

⁷Laboratory of Experimental Pharmacogenetics
Department of Clinical Pharmacy and Biopharmacy

University of Medical Sciences
Św. Marii Magdaleny 14
61-861 Poznań, Poland

*corresponding author: tel.: +4861 6659550, fax: +4861 6659551,
e-mail: agnieszka.gryszczynska@iwnirz.pl

S u m m a r y

The purpose of presented study was the comparison of flavan-3-ol content in the roots of *Rhodiola kirilowii* and *Rhodiola rosea* with use of UPLC-MS/MS method. Two kinds of extract were prepared: aqueous extract and 50% v/v ethanol extract. The worked out UPLC MS/MS method allowed to determine the content of five flavan-3-ols: (+)-catechin, (-)-epicatechin, (-)-epigallocatechin, (-)-epicatechin gallate (ECG) and (-)-epigallocatechin gallate (EGCG). The obtained results shown that the content of measured catechins was higher in *R. kirilowii* roots than in *R. rosea*. Both *Rhodiola* roots contain EGCG as a main proanthocyanidin compound – the content in *R. kirilowii* roots is about 0.14%. Our results indicate that application of ultra performance liquid chromatograph connected to a tandem mass spectrometer (UPLC MS/MS method) allows to determine the proanthocyanidins content in tested samples with satisfactory precision and can be used in the *Rhodiola* sp. phytochemical investigations.

Key words: *Rhodiola kirilowii*, *Rhodiola rosea*, flavan-3-ol content, UPLC-MS/MS method, (-)-epigallocatechin gallate (EGCG)

INTRODUCTION

Two species of *Rhodiola* L. genus (*Crassulaceae* family) are most frequently used in official and traditional medicine: *R. rosea* and *R. kirilowii*.

R. kirilowii (Regel.) Maxim grows in mountains at an altitude of 2000–5600 m in Asia: mainly Tien-Shan, Altaj, Pamir [1]. The main pharmacological activity of extracts from *R. kirilowii* is preventing high altitude reactions of human organism [2]. The roots contain: phenylethanoids as *p*-tyrosol and salidroside [3-5], phenylpropanoids [4, 5], catechins [4, 6-10], coumarins [11], phenolic acids [4-7, 10], phytosterols [4], tannins [10], cyanogenic glycosides [9], arbutin [9], terpenoids [8].

Rhodiola rosea L., roseroot (synonyms: Golden root, Arctic root), is a herbaceous perennial plant growing in Arctic and in the mountainous regions of Asia, North America and Europe. This plant shows some physiological and pharmacological properties: stimulates the central nervous system (CNS) [12-13], enhances physical and mental work performance [14-15], eliminates fatigue and possesses adaptogenic [16-17], cardioprotective [18], anticancer [19], antioxidant [20-22] and antimicrobial activities [23, 24]. Some activities of extracts have been proved in pharmacological and clinical studies [14, 15]. The roots of *R. rosea* contain: phenylpropanoids

– rosavin, rosarin, rosin [25], phenylethanoids – salidroside, p-tyrosol [21, 25], flavonoids – rodionin, rodiolin, rhodiosin, acetylrodalgin and tricin [26-28], phenolic acids [30], monoterpenes [30], phytosterols [31], tannins [30], cyanogenic glucoside – lotaustralin [32] and essential oils – n-decanol, geraniol [33].

Chemical constituents of both *Rhodiola* species, pharmacological activities of roots extracts and *in vitro* cultures have been investigated in the Institute of Natural Fibres and Medicinal Plants (previously: Research Institute of Medicinal Plants) for several years [4, 6-9, 13, 21, 23, 24, 34]. The purpose of present study was the comparison of flavan-3-ol content in the roots of these species using UPLC-MS/MS method.

MATERIAL AND METHODS

Plant material

The roots of *R. kirilowii* and *R. rosea* were harvested from field cultivation (in the Garden of Medicinal Plants, Institute of Natural Fibres and Medicinal Plants in Plewiska near Poznań, Poland) in October 2009. The roots were dried in the room temperature (about 22–24°C).

Preparation of plant extracts

Two kinds of extract were prepared: aqueous extract and 50% (v/v) ethanol extract.

Preparation of aqueous extract

The powdered dry roots were extracted with purified water for 3 h at 90°C (1:10 plant material to solvent ratio). After filtering, the extracts were frozen at -55°C and then lyophilized.

Preparation of 50% (v/v) ethanol extract

The powdered dry roots were extracted with 50% (v/v) ethanol using the percolation method at a 1:10 ratio of plant material to solvent. After evaporation of the alcohol under reduced pressure at a temperature of 40–45°C, the extracts were frozen at -55°C and then lyophilized.

The dry plant extracts were stored at a temperature of 20–25°C.

Standard substances

The following substances were used in the experiment for comparison: (+)-catechin, (-)-epicatechin, (-)-epigallocatechin, (-)-epicatechin gallate, (-)-epigallocatechin gallate (ChromaDex) and D-(-)-salicine (SIGMA).

Preparation of test samples: extraction of flavan-3-ols from dry plant materials and from plants extracts

The method of flavan-3-ol extraction from roots and extracts by P. Mammela [35] was used. An exact amount of ca. 0.5 g of dried and powdered (0.315) *R. kirilowii* roots, an exact amount of ca. 0.75 g of dried and powdered (0.315) *R. rosea* roots and ca. 0.1 g of dried and powdered extracts (aqueous and 50% v/v ethanol) from *R. kirilowii* and *R. rosea* roots were weighed out and placed in a 20 ml volumetric flasks. 15.0 ml of 80% v/v methanol was added and the solutions were subjected to ultrasounds for 60 min at a room temperature (20–25°C). Subsequently, the solutions were made up to the mark with the same solvent and filtered on a quantitative filter paper. The filtrates were concentrated to evaporate the methanol up to a volume of about 1/5 in a rotary evaporator in vacuum. The residues were extracted with 4 × 16.0 ml of diethyl ether. The combined ether extracts were dried with anhydrous sodium sulphate and evaporated to dryness in a rotary evaporator in vacuum. The dry residues were dissolved in 4.0 ml of 10% v/v methanol and then transferred quantitatively to 5 ml volumetric flasks. 0.023 ml of 0.5 mg/ml D-(-)-salicine (IS) was added to every flasks and the solutions were made up to the mark with 10% v/v methanol. The samples were filtered through a membrane filter of a 0.20 µm diameter.

LC-MS/MS assay

The validated assay using ultra performance liquid chromatograph connected to tandem mass spectrometer (UPLC-ESI MS/MS; Waters) was worked out in the Institute of Natural Fibres and Medicinal Plants [34]. The preparation of calibration curves for flavan-3-ols, the evaluation of precision, linearity and accuracy of this analytical method is described by Gryszczyńska et al. [34].

Statistical analysis

The results of the study were statistically verified determining the relative standard deviation (RSD), n=6.

RESULTS AND DISCUSSION

Flavan-3-ols (so called proanthocyanidins) have the antioxidative activity, thus, they protect against harmful effects of free radicals and reactive oxygen forms. They also show anticarcinogenic, anti-inflammatory, antiallergenic,

antimutagenic and antiaging activity as well as improve the function of liver [36]. There are also reports that catechins can prevent obesity. Tests performed on animals have confirmed their activity in hypercholesterolaemia [38]. According to the fact that supplements containing *R. rosea* extract are recommended for protection against many pathogenic agents as well as that using of *R. kirilowii* extract in the Far East ethno-medicine is connected with the protection of human against high altitude reactions, the determination of proanthocyanidins in the *Rhodiola* genus could help explain the activities of the extracts.

The content of flavan-3-ols was investigated in *R. kirilowii* and *R. rosea* roots and in the extracts prepared according to the method described above. The UPLC MS/MS method worked out in the Institute of Natural Fibres and Medicinal Plants [34], allows to determine the contents of five flavan-3-ols: (+)-catechin, (-)-epicatechin, (-)-epigallocatechin, (-)-epicatechin gallate (ECG), (-)-epigallocatechin gallate (EGCG). The MRM chromatograms showing the fragmentation of flavan-3-ols from *Rhodiola kirilowii* and *R. rosea* roots are demonstrated in figures 1 and 2. The obtained quantitative results are presented in tables 1 and 2.

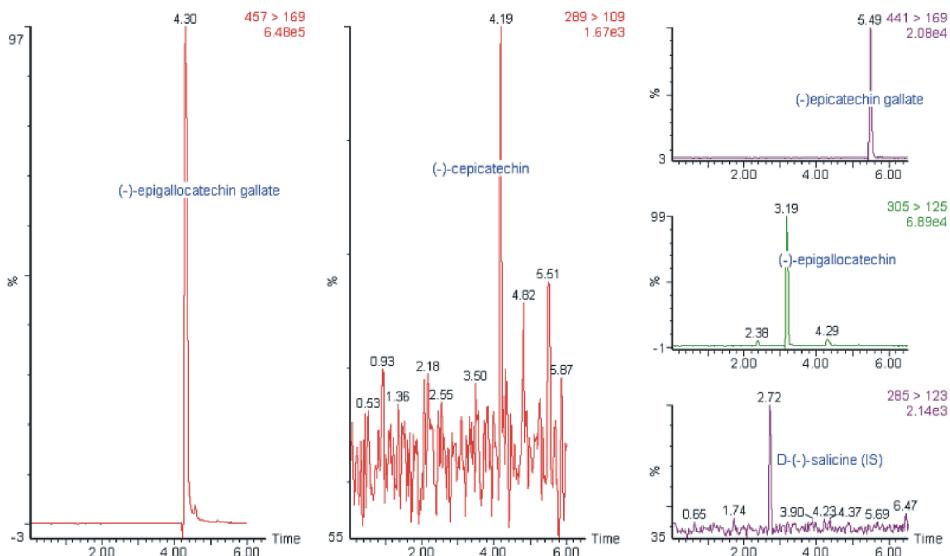


Figure 1.
MRM chromatogram showing fragmentation of flavan-3-ols from *Rhodiola kirilowii* roots.

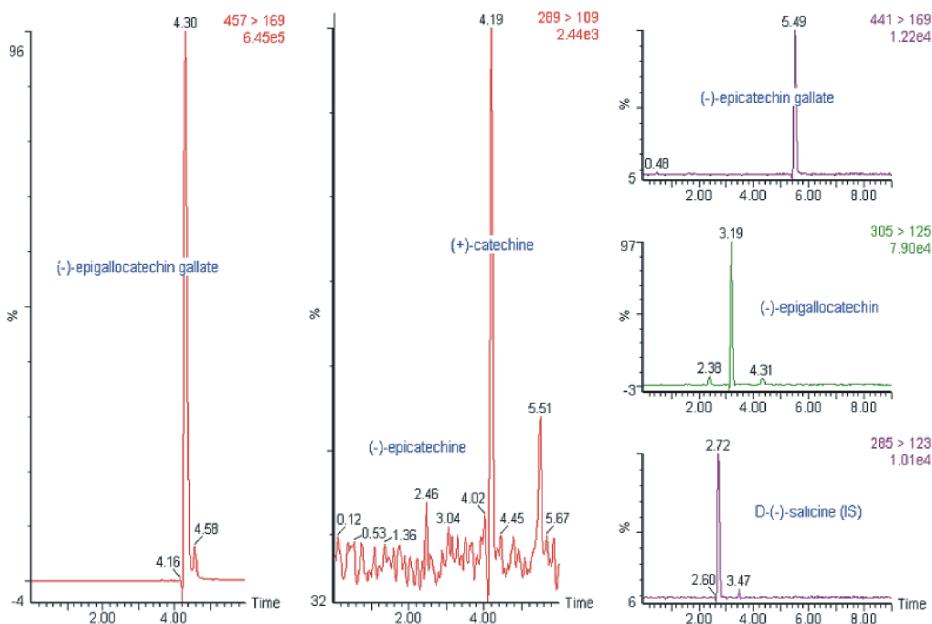


Figure 2.
MRM chromatogram showing fragmentation of flavan-3-ols from *Rhodiola rosea* roots

Table 1.

Content of catechins in *Rhodiola kirilowii* and *Rhodiola rosea* roots

Sample	(+)-Catechin		(-)-Epicatechin		(-)-Epigallo-catechin		(-)-Gallate epicatechin		(-)-Gallate epigallocatechin	
	Content [mg/100 g] ¹	RSD [%]*								
<i>R. kirilowii</i> root	0.097	2.06	0.288	2.08	19.584	7.53	5.294	4.55	135.435	2.00
<i>R. rosea</i> root	0.051	3.92	0.135	1.48	8.526	1.77	0.437	2.29	36.873	2.17

*- RSD – relative standard deviation (n=6)

¹ – the content in 100 g of powdered dry material

The obtained results show that the content of measured catechins was higher in *R. kirilowii* roots than in *R. rosea*. Both *Rhodiola* roots contain (-)-gallate epigallocatechin as a main proanthocyanidin compound – the content in *R. kirilowii* roots is about 0.14% (tab. 1).

(-)-Epicatechin and (-)-epigallo-catechin were found in the highest concentration in ethanol extract, whereas (+)-catechin and (-)-gallate epigallocatechin in the aqueous extracts (tab. 2).

Table 2.

Content of catechins in *Rhodiola kirilowii* and *Rhodiola rosea* extracts

Sample	(+)-Catechin		(-)-Epicatechin		(-)-Epigallo-catechin		(-)-Gallate epicatechin		(-)-Gallate epigallocatechin	
	Content [mg/100 g] ¹	RSD [%]*								
<i>R. kirilowii</i>										
50% ethanol extract	0.377	1.86	1.651	1.51	97.357	3.81	4.045	3.51	266.494	2.72
<i>R. kirilowii</i>										
aqueous extract	0.565	2.12	0.858	3.38	31.558	5.81	5.138	2.80	304.449	3.78
<i>R. rosea</i>										
50% ethanol extract	<LOD	-	0.613	1.63	24.978	5.85	1.589	0.63	133.407	4.10
<i>R. rosea</i>										
aqueous extract	0.381	2.62	0.392	2.55	3.081	7.14	0.955	4.19	299.702	3.67

* – RSD – relative standard deviation (n=6)

¹ – the content in 100 g of powdered dry material

Searched proanthocyanidins were reported in *R. kirilowii* roots by Wiedenfeld et al. [9], Zuo et al. [10] and Wong et al. [5]. The presence of (-)-gallate epigallocatechin (EGCG) in *R. kirilowii* roots was also demonstrated in our previous investigations on biological activities of extracts [6-8] or searching on tissue cultures of *R. kirilowii* [38] – the content varied from 68 to 1734 mg/100 g d.w. according to harvest time (as was determined by HPLC method) [38].

Proanthocyanidins in *R. rosea* were searched by Yousef et al. [36]. They found that roots contains (-)-epigallocatechin and its 3-O-gallate esters in different degrees of polymerization. In our previous investigation on *R. rosea* we have found that the content of (-)-gallate epigallocatechin (EGCG) was different in various periods of vegetation and varied from 141 to 399 mg/100 g d.w. [39], as it was determined by HPLC method.

The above mentioned determination of EGCG content in *R. kirilowii* and *R. rosea* roots [38, 39] and our results presented above indicate that *R. kirilowii* can be a better source of catechin as compared with *R. rosea* roots, especially if the content of (-)-gallate epigallocatechin is taken into consideration. In this case, the water extraction is recommended. The application of ultra performance liquid chromatograph connected to a tandem mass spectrometer (UPLC MS/MS method) allows the determination of the proanthocyanidins content in tested samples with satisfactory precision and can be used in the *Rhodiola* sp. phytochemical investigations.

ACKNOWLEDGEMENT

This research project was financed by the Ministry of Science and Higher Education under grant No. N N405 306136.

REFERENCES

1. Flora of China. Wu Zheng-yi Raven PH (ed.). Science Press (Beijing), Missouri Botanical Garden Press (St. Louis), 2001; 8:51-268.
2. Zhang ZH, Feng SH, Hu GD, Cao ZK, Wang LY. Effect of *Rhodiola kirilowii* (Regel.) Maxim on preventing high altitude reactions. A comparison of cardiopulmonary function in villagers at various altitudes. China J of Chinese Materia Medica 1989; 14(11):687-90.
3. Krasnov EA, Kuvaiev VB, Choružaya TG. Chemotaksonomic investigations of *Rhodiola* sp. Rast Res 1978; 14(2):153-160.
4. Krajewska-Patan A, Furmanowa M, Derger M, Łowicka A, Górska-Paukszta M, Mścisza A, Mielcarek S, Przybylak JK, Buchwald W, Mrozikiewicz PM. Zawartość związków biologicznie czynnych w hodowlach kalusa i w hodowlach zawiesinowych *Rhodiola Kirilowii* (Regel.) Maxim. Herba Pol 2006; 52(3):47-8.
5. Wong YC, Zhao M, Zong YY, Chan CY, Che CT. Chemical constituents and anti-tuberculosis activity of root of *Rhodiola kirilowii*. China J of Chinese Materia Medica 2008; 33(13):1561-5.
6. Mścisza A, Mielcarek S, Buchwald W, Krajewska-Patan A, Furmanowa M, Skopińska-Różewska E, Luczkowska T, Mrozikiewicz PM. Phytochemical study of *Rhodiola rosea*, *Rhodiola quadrifida* and *Rhodiola kirilowii* extracts. Basic Clin Pharmacol Toxicol 2005; 97(suppl I):41.
7. Buchwald W, Mścisza A, Krajewska-Patan A, Furmanowa M, Przybylak J, Luczkowska T, Mrozikiewicz PM. Contents of biological active compounds of *Rhodiola kirilowii* roots during the vegetation. Herba Pol 2005; 51(suppl. 1):105-6.
8. Mielcarek S, Mścisza A, Buchwald W, Krajewska-Patan A, Furmanowa M, Skopińska-Różewska E, Luczkowska T, Mrozikiewicz PM. Phytochemical investigation of *Rhodiola* sp. roots. Herba Pol 2005; 51(suppl 1):159-160.
9. Wiedenfeld H, Zych M, Buchwald W, Furmanowa M. New compounds from *Rhodiola kirilowii*. Sci Pharm 2007; 75:29-34.
10. Zuo G, Li Z, Chen L, Xu X. Activity of compounds from Chinese herbal medicine *Rhodiola kirilowii* (Regel) Maxim against HCV NS3 serine protease. Antiviral Res 2007; 76(1):86-92.
11. Zhang S, Wang J, Zhang H. Chemical constituents of Tibetan medicinal herb *Rhodiola kirilowii* (Reg.) Reg. China J of Chinese Materia Medica 1991; 16(8):483,512.
12. Saratikov A, Marina TF, Fisanova LL, Effect of golden root extract on processes of serotonin synthesis in CNS. J Biol Sci 1978; 6:142.
13. Krajewska-Patan A, Mikołajczak PL, Okulicz-Kozaryn I, Bobkiewicz-Kozłowska T, Buchwald W, Łowicka A, Furmanowa M, Dreger M, Górska-Paukszta M, Mścisza A, Mrozikiewicz PM. *Rhodiola rosea* extracts from roots and callus tissues – study on relationship between their chemical contents and CNS affecting pharmacological activity. 11th International Congress of Polish Herbal Committee. Poznań, June, 24-25 2005. Herba Pol 2005; 51(Suppl. 1):105-6.
14. Darbinyan V, Kteyan A, Panossian A Gabrielian E, Wikman G, Wagner H. *Rhodiola rosea* in stress induced fatigue: a double blind cross-over study of a standarized extract SHR-5 with a repeated low-dose regimen on the mental performance of healthy physicians during night duty. Phytomedicine 2000; 7(5):365-71.
15. Spasov AA Wilkman GK, Mandrikov VB, Mironova IA, Neumoin VV. A double - blind, placebo - controlled pilot study of the stimulating and adaptogenic effect of *Rhodiola rosea* SHR-5 extract on the fatigue of students caused by stress during an examination period with a repeated low-dose regimen. Phytomedicine 2000; 7(2):85-9.
16. Kelly GS. *Rhodiola rosea*: a possible plant adaptogen. Altern Med Rev 2001; 6(3):293.

17. Panossian A, Wagner H. Stimulating effect of adaptogens: an overview with particular reference to their efficacy following single dose administration. *Phytother Res* 2005; 19(10):819-38.
18. Maslova LV, Kondratev Blu, Maslov LN, Lishmanov JB. The cardioprotective and antiadrenergic activity of an extract of *Rhodiola rosea* in stress. *Eks Klin Farmakol* 1994; 57(6):61-63.
19. Uditsev SN, Shakhev VP. Decrease of cyclophosphamide haemotoxicity by *Rhodiola rosea* root extract in mice with Ehrlich and Lewis transplantable tumors. *Eur J Cancer* 1991; 27:1182.
20. Furmanowa M, Skopińska-Różewska E, Rogala E, Hartwich M. *Rhodiola rosea* L. *in vitro* culture – phytochemical analysis and antioxidant action. *Acta Soc Bot Pol* 1998; 67:69.
21. Furmanowa M, Kędzia B, Hartwich M, Kozłowski J, Krajewska-Patan A, Mścisz A, Jankowiak J. Phytochemical and pharmacological properties of *Rhodiola rosea* L. *Herba Pol* 1999; 45:108-13.
22. Battistelli M, De Sanctis R, De Bellis R, Cucchiari L, Dacha M, Gobbi P. *Rhodiola rosea* as antioxidant in red blood cells: ultrastructural and hemolytic behaviour. *Eur J Histochem* 2005; 49(3):243-54.
23. Furmanowa M, Starościk B, Lutomski J, Kozłowski J, Urbańska N, Krajewska-Patan A, Pietrosiuk A, Szypuła W. Antimicrobial effect of *Rhodiola rosea* L. roots and callus extracts on some strains of *Staphylococcus aureus*. *Herba Pol* 2002; 48:23.
24. Krajewska-Patan A, Kędzia B, Dreger M, Mścisz A, Buchwald W, Furmanowa M, Mrozikiewicz PM. Antimicrobial activity of *Rhodiola rosea* extracts. Proceeding of 7th Congress of the European Association for Clinical Pharmacology and Therapeutics; 2005, 25-29 June, Poznań, Poland. Abstract Book. Basic Clin Pharmacol Toxicol 2005; 97(Suppl.1):38.
25. Kir'yanov A, Bondarenko L, Kurkin V, Zapesochnaya G et al. Determination of biologically active constituents of *Rhodiola rosea* rhizomes. *Khim-Prir Soedin* 1991; 3:320.
26. Kurkin V, Zapesochnaya G, Klyaznina V. *Rhodiola rosea* rhizome flavonoids. *Khim Prir Soedin* 1982; 5:581.
27. Kurkin V, Zapesochnaya G, Shchavinskii A. Flavonoids of rhizomes of *Rhodiola rosea*. III. *Khim Prir Soedin* 1984; 5:657.
28. Zapesochnaya G, Kurkin V. Flavonoids of *Rhodiola rosea* rhizomes. I. *Khim Prirod Soed* 1983; 1:23.
29. Dubichev A, Kurkin W, Zapesochnaya G, Vorontsov V. HPLC study of *Rhodiola rosea* rhizomes. *Khim Prir Soedin* 1991; 2:188.
30. Kurkin V, Zapesochnaya G. Chemical composition and pharmacological properties of *Rhodiola* sp. Plants Review. *Khim-Farm Zh* 1986; 20(10):1231.
31. Kurkin V, Zapesochnaya GG, Kir'yanov AA et al. Quality of raw *Rhodiola rosea* L. material. *Khim-Farm Zh* 1989; 23(11): 1364-67.
32. Akgul Y, Ferreira D, Abourashed EA, Khan IA. Lotaustralin from *Rhodiola rosea* roots. *Fitoterapia* 2004; 75(6):612-4.
33. Rohloff J. Volatiles from rhizomes of *Rhodiola rosea* L. *Phytochemistry* 2002; 59:655:61.
34. Gryszczyńska A, Mielcarek S, Buchwald W. The determination of flavan-3-ol content in the root of *Rhodiola Kirilowii*. *Herba Pol* 2011; 51(1): 27-37.
35. Mammella P. Phenolics in selected European hardwood species by liquid chromatography-electrospray ionization mass spectrometry. *Analyst* 2001; 126:1535-1538.
36. Yousef GG, Grace MH, Cheng DM, Belolipov IV, Raskin I, Lila MA. Comparative phytochemical characterization of three Rhodiola species. *Phytochemistry* 2006; 67:2380-2391.
37. Murakami I, Nakamura T, Ishibashi Y, Shibuya R, Ayano E, Morita-Murase Y, Nagata Y, Kanazawa H. Simultaneous determination of catechins and procyanidins in bottled tea drinks by LC/MS. *Chromatography* 2006; 27(1):27-33.
38. Krajewska-Patan A, Dreger M, Buchwald W, Górska-Paukszta M, Mielcarek S, Baraniak M, Furmanowa M, Mrozikiewicz PM. The obtaining of the enriched *Rhodiola Kirilowii* callus biomass by exogenous supplementation with p-tirosol and cinnamyl alcohol. *Pamiętnik Puławski* 2009;151/I:183-192.
39. Buchwald W, Mścisz A, Krajewska-Patan A, Furmanowa M, Mielcarek S, Mrozikiewicz PM. Contents of biologically active compounds in *Rhodiola rosea* roots during the vegetation period. *Herba Pol* 2006;52(4):39-43.

PORÓWNANIE ZAWARTOŚCI PROANTOCYJANIDYN W KORZENIACH *RHODIOLA KIRILOWII*
I *RHODIOLA ROSEA* Z WYKORZYSTANIEM METODY UPLC-MS/MS

AGNIESZKA GRYSZCZYŃSKA^{1*}, ANNA KRAJEWSKA-PATAN², WALDEMAR BUCHWALD³,
BOGUSŁAW CZERNY^{4,5}, SEBASTIAN MIELCAREK¹, KAROLINA RUDZIŃSKA⁶,
PRZEMYSŁAW M. MROZIKIEWICZ^{1,7}

¹Zakład Badania Jakości Produktów Leczniczych i Suplementów Diety
Instytut Włókien Naturalnych i Roślin Zielarskich
ul. Libelta 27
60-707 Poznań

²Zakład Farmakologii i Biologii Doświadczalnej
Instytut Włókien Naturalnych i Roślin Zielarskich
ul. Libelta 27
60-707 Poznań

³Zakład Botaniki, Hodowli i Agrotechniki
Instytut Włókien Naturalnych i Roślin Zielarskich
ul. Kolejowa 2
62-064 Plewiska k/Poznania

Instytut Włókien Naturalnych i Roślin Zielarskich
ul. Libelta 27
60-707 Poznań

⁵Zakład Farmakologii Ogólnej i Farmakoekonomiki
Wydział Nauk o Zdrowiu, Pomorski Uniwersytet Medyczny
ul. Żołnierska 48
70-204 Szczecin

⁶109 Szpital Wojskowy SPZOZ
ul. Piotra Skargi 9-11
70-965 Szczecin, Poland

⁷Pracownia Farmakogenetyki Doświadczalnej
Katedra i Zakład Farmacji Klinicznej i Biofarmacji
Uniwersytet Medyczny w Poznaniu
ul. Św. Marii Magdaleny 14
61-861 Poznań

*autor, do którego należy kierować korespondencję: tel.: +4861 6659550,
faks: +4861 6659551, e-mail: agnieszka.gryszczynska@iwnirz.pl

Streszczenie

Celem przedstawionych badań było porównanie za pomocą opracowanej metodyki wykorzystującej ultrasprawny chromatograf cieczowy sprzężony z tandemowym spektrometrem mas (Waters) zawartości flavan-3-oli w korzeniach *Rhodiola kirilowii* i *Rhodiola rosea*. Badano wodne i alkoholowo-wodne (50% EtOH) wyciągi z korzeni. Opracowana metoda UPLC MS/MS pozwoliła na określenie stężeń pięciu flavan-3-oli: (+)-katechiny, (-)-epikatechiny, (-)-epigalokatechiny, galusanu (-)-epikatechiny (ECG) oraz galusanu (-)-epigalokatechiny (EGCG). Otrzymane wyniki wskazują, że zawartość tych katechin jest wyższa w korzeniach *R. kirilowii* niż w korzeniach *R. rosea*. Oba surowce zawierają galusan (-)-epigallokatechiny jako główną proantocyjanidynę – jej zawartość w korzeniach *R. kirilowii* wynosi ok. 0,14%. Uzyskane przez nas wyniki wskazują, że metoda analityczna z wykorzystaniem ultrasprawnego chromatografu cieczowego sprzężonego z tandemowym spektrometrem mas pozwala z zadawalającą precyzją oznaczyć zawartości proantocyjanidyn w analizowanych próbkach i może być stosowana w badaniach rodzaju *Rhodiola*.

Słowa kluczowe: *Rhodiola kirilowii*, *Rhodiola rosea*, zawartość flavan-3-oli, UPLC-MS/MS, galusan epigalokatechiny (EGCG)